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<b>Article Title (within 20 words without abbreviations)</b>	Evaluation of forage production, feed value, and ensilability of proso millet ( <i>Panicum miliaceum</i> L.)
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5 Running title: Proso millet as conserved forage

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7 **Evaluation of forage production, feed value, and ensilability of**  
8 **proso millet (*Panicum miliaceum* L.)**

9

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25 **Abstract**

26 Whole-plant corn (*Zea mays* L.) and sorghum-sudangrass hybrid (*Sorghum bicolor* L.) are major  
27 summer crops that can be fed as direct-cut or silage. Proso millet is a short-season growing crop with  
28 distinct agronomic characteristics that can be productive in marginal lands. However, information is  
29 limited about the potential production, feed value, and ensilability of proso millet forage. We evaluated  
30 proso millet as a silage crop in comparison with conventional silage crops. Proso millet was sown on  
31 June 8 and harvested on September 5 at soft-dough stage. Corn and sorghum-sudangrass hybrid were  
32 planted on May 10 and harvested on September 10 at the half milk-line and soft-dough stages,  
33 respectively. The fermentation was evaluated at 1, 2, 3, 5, 10, 15, 20, 30, and 45 days after ensiling.  
34 Although forage yield of proso millet was lower than corn and sorghum-sudangrass hybrid, its relative  
35 feed value was greater than sorghum-sudangrass hybrid. Concentrations of dry matter (DM), crude  
36 protein, and water-soluble carbohydrate decreased commonly in the ensiling forage crops. The DM  
37 loss was greater in proso millet than those in corn and sorghum-sudangrass hybrid. The *in vitro* dry  
38 matter digestibility declined in the forage crops as fermentation progressed. In the early stages of  
39 fermentation, pH dropped rapidly, which was stabilized in the later stages. Compared to corn and  
40 sorghum-sudangrass hybrid, the concentration of ammonia-nitrogen was greater in proso millet. The  
41 count of lactic acid bacteria reached the maximum level on day 10, with the values of 6.96, 7.77, and  
42 6.95 log<sub>10</sub> cfu/g fresh weight for proso millet, corn, and sorghum-sudangrass hybrid, respectively. As  
43 ensiling progressed, the concentrations of lactic acid and acetic acid of the three crops increased and  
44 lactic acid proportion became higher in the order of sorghum-sudangrass hybrid, corn, and proso millet.  
45 Overall, the shorter, fast-growing proso millet comparing with corn and sorghum-sudangrass hybrid  
46 makes this forage crop an alternative option, particularly in areas where agricultural inputs are limited.  
47 However, additional research is needed to evaluate the efficacy of viable strategies such as chemical  
48 additives or microbial inoculants to minimize ammonia-nitrogen formation and DM loss during

49 ensiling.

50 **Key words:** Proso millet, Corn, Sorghum-sudangrass hybrid, Silage, Conservation

51 **INTRODUCTION**

52 South Korea is a country with scarce agricultural resources. About two thirds of its land area is  
53 mountains and hills. The cultivated area only accounts for 22% of the total land area. It has one of the  
54 lowest per capita cultivated land areas in the world. The livestock industry accounts for almost 40% of  
55 total agricultural production in South Korea [1]. With the development of the livestock industry, the  
56 forage industry has attracted increasing attention. The forage industry is the basis for the survival and  
57 development of the livestock industry. However, South Korea's current self-produced feed resources  
58 are relatively limited, and some feeds must still be imported from overseas. As the most basic  
59 production source of animal products, problems with the feed supply will affect the sustainable  
60 development of the whole livestock industry. To stabilize the livestock industry and agricultural  
61 production, the production of high-quality forage would reduce feed costs and have an import  
62 substitution effect.

63 Corn and sorghum-sudangrass hybrids are the two most common forage crops that are used mainly  
64 as summer-season forages in dairy and beef rations. They have low production costs, high yield, and a  
65 relatively high nutritional value. Proso millet (*Panicum miliaceum* L.) is a short growing, summer  
66 season crop (60 to 100 days) with unique agronomic properties such as high tolerance to heat and  
67 drought conditions, and is cultivated in abundance in Asian and African countries [2-4]. Proso millet  
68 crop has the potential to remain productive in areas with marginal lands and limited agricultural inputs,  
69 where cultivation of major crops such as corn is restricted [5-7]. Proso millet could be a viable  
70 alternative to main summer forages in areas where cultivation of corn or sorghum-sudangrass is  
71 restricted due to a longer growing season or poor agricultural conditions [8].

72 Ensiling has long been recognized as a simple and effective method of preserving moist forage,

73 ensuring a continuous supply of forage to animals [9, 10]. To our knowledge, few studies have  
74 investigated the fermentation dynamics of proso millet forage. The purpose of this research was to  
75 provide basic information about the ensiling feasibility of forage from proso millet in comparison to  
76 commonly cultivated summer crops (whole-plant corn and sorghum-sudangrass hybrid).

## 77 **MATERIALS AND METHODS**

### 78 **Crop establishment and management**

79 Establishment of experimental plots was made at the experimental site of Seoul National University,  
80 Pyeongchang Campus (located at 37° 32' 40" N, 128° 26' 33" E, average altitude is about 550 m above  
81 sea level) during the summer season of 2019. A detailed description of meteorological data including  
82 temperature and precipitation throughout the growing season (May to September, 2019) is illustrated  
83 in Figure 1. During the growing season, temperature ranged from 15.9 to 26.8°C (average = 21.5°C).  
84 Soil analysis on the 0–15-cm soil depth of the experimental site showed that it was slightly acidic (pH  
85 6.55; soil:water suspension = 1:5), with 14.1% organic matter, 0.12% total nitrogen, and a cation  
86 exchange capacity of 16.5 cmol(+)/kg. Concentration of exchangeable cations including Ca, K, Mg,  
87 and Na averaged 1.75, 4.01, 0.92, and 0.10 mg/kg, respectively. For the three crops, nitrogen,  
88 phosphorus, and potassium fertilizers were applied at a rate of 200, 150, and 150 kg/ha, respectively.  
89 After preparation of seedbed, seeds were sown manually and grown on 3 replicate plots/each crop.  
90 Each plot was 3 m × 5 m in size. Proso millet (*Panicum miliaceum* L. var. Geumsilchal) was planted  
91 on June 8 at a seeding rate of 20 kg/ha, and harvested on September 5. Sorghum-sudangrass hybrid  
92 (*Sorghum bicolor* L. var. Turbo-gold) was sown at a seeding rate of 40 kg/ha on May 10 and harvested  
93 on September 10. Corn (*Zea mays* L. var. Gwangpyeongok) was sown on May 10 at a plant-to-plant  
94 distance of 20 cm and an inter-row spacing of 75 cm. Whole-crop corn was harvested on September  
95 10. Sorghum-sudangrass hybrid and proso millet were harvested when they reached soft-dough stage

96 of the seedhead. Whole-crop corn was harvested at about the half milk-line stage, which is a reliable  
97 visionary criterion indicating the optimum time to harvest whole plant for silage making [11]. This  
98 was accomplished by splitting the corn ear in the center and visually inspecting the kernel milkline.  
99 Whole-crop corn was fractionated into cob (containing kernel and rachis) and stover component that  
100 was consisted of the remaining components of the plant after cob removal [12]. These fractions were  
101 separately weighed and approximately 1-kg representative subsamples were collected for dry matter  
102 (DM) determination. The proportion of these fractions in the whole plant was then calculated. Forage  
103 yield was determined by manually harvesting the forage material in the whole plot and calculating the  
104 fresh forage yield, which was then converted to units of fresh and DM/hectare.

#### 105 **Silage preparation**

106 At harvest, four whole-crop plants from center rows in each plot were randomly selected and  
107 chopped into approximately 2–3 cm long pieces using a chopper (Richi Machinery Co., Ltd, Henan,  
108 China). The chopped crops were grouped into separate piles per each plot for silage experiment. The  
109 representative allotments were also collected for quality assessment of fresh biomass before ensiling.  
110 Ensiling was made by packing approximately 600 g chopped material into plastic film bags (28 cm ×  
111 36 cm). The bags were vacuum-sealed (Zhejiang Hongzhan Packing Machinery Co., Ltd, Wenzhou  
112 city, China) and stored in a dark and dry condition at room temperature (about 22°C). Bags were  
113 randomly opened on days 1, 2, 3, 5, 10, 15, 20, 30, and 45 of ensiling for quality assessment of silage  
114 fermentation. Silos were weighed at designated openings for DM loss determination [13]. Number of  
115 replicate silos for each crop at each opening was 3. Therefore, the design arrangement for the three  
116 forage types in the silage trial was as follows: 3 forage types × 9 silo openings × 3 replications, resulting  
117 in formation of a total of 81 silos. At each silo opening, the ensiled material inside each silo was  
118 emptied, mixed thoroughly and divided into 3 representative portions. The first portion was dried  
119 (65°C) to a constant weight and used for the chemical composition analysis. The second portion was

120 stored in a freezer at  $-80^{\circ}\text{C}$  (TSE400D, Thermo Fisher Scientific, Waltham, MA, USA) for  
121 quantification of organic acids and ammonia nitrogen. The third subsample was used for enumeration  
122 of microbial population in ensiled biomass.

### 123 **Analytical analyses**

124 A 10-g fresh silage sample was placed into a 250 mL conical flask and covered with 100 mL distilled  
125 water. The flasks were shaken for 1 h on a mechanical shaker (Green Sseriker, Vision Scientific,  
126 Gyeonggi-Do, Korea) and stored in refrigerator for 24 h. The conical flasks were shaken by hand every  
127 2 hours during refrigeration. The mixture was filtered through a filter paper (Whatman No. 6,  
128 Advantech, Zurich, Switzerland). Silage pH was determined in the filtrate with a pH meter (AB 150,  
129 Fisher Scientific International, Inc., Pittsburgh, PA, US). A 1.5 mL portion of the filtrate was used for  
130 analysis of the organic acid concentration using high performance liquid chromatography (HPLC,  
131 Agilent Technologies, Santa Clara, CA, US) equipped with a refractive index detector [8]. Ammonia  
132 nitrogen ( $\text{NH}_3\text{-N}$ ) was analyzed via the method described by Broderick and Kang [14]. The spread-  
133 plate method [15] was used to enumerate the population of microorganisms. In brief, a 10-g sample  
134 was diluted with 90 mL sterilized saline solution (8.50 g/L NaCl) and shaken for 1 h. Lactic acid  
135 bacteria (LAB), molds, and total microorganisms were enumerated on Rogosa, and Sharpe  
136 agar medium, potato dextrose agar, and plate count agar media, respectively. The limit of detection  
137 was  $2 \log_{10}$  CFU/g fresh mass.

138 Dry matter concentration in ensiled material was determined in triplicate at  $65^{\circ}\text{C}$  in a forced drying  
139 oven for 72 h. The dried samples were ground to pass through a 1 mm screen (Thomas Scientific, Inc.,  
140 Swedesboro, NJ, USA) for nutrient composition analysis. Total nitrogen was quantified via the Dumas  
141 method [16], and crude protein (CP) was calculated as nitrogen  $\times$  6.25. Acid detergent fiber (ADF)  
142 and neutral detergent fiber (NDF) were measured following the method of Van Soest et al. [17]. Water-  
143 soluble carbohydrate (WSC) was analyzed via a modification of the anthrone method proposed by

144 Yemm and Willis [18].

145 ***In vitro* dry matter digestibility**

146 *In vitro* DM digestibility (IVDMD) was performed in triplicate using an Ankom Daisy<sup>II</sup> incubator  
147 (ANKOM Technologies, Inc., Fairport, NY, USA) [19], as described by Goering and Van Soest [20].  
148 Ground samples (0.5–0.6 g) were weighed into F57 filter bags and sealed using a heat sealer. Samples  
149 were evenly distributed on both sides of the digestion jars. Then, 1330 mL buffer solution A and 266  
150 mL buffer solution B were added to each jar. Two ruminally cannulated Holstein steers were selected  
151 and their rumen fluid was collected before the morning feed and passed through four layers of  
152 cheesecloth. Then, 400 mL rumen fluid was added to the buffer solution and samples. The digestion  
153 jar was purged with CO<sub>2</sub> gas for 30 s and then closed with a lid. The jars were incubated at 39°C for  
154 48 h. Undigested NDF residues in original bags were extracted using an ANKOM<sup>2000</sup> fiber analyzer.

155 **Statistical analysis**

156 Field experiment was arranged in a completely randomized block design with three replications. Data  
157 were subjected to analysis of variance (ANOVA) using the general linear model (GLM) in SPSS (IBM  
158 SPSS Statistics, Version 24.0 Armonk, NY, INM Corp). Individual plot was regarded as the  
159 experimental unit in the model for analysis of data from the field experiment (Table 1). Individual silo  
160 served as the experimental unit in the model for analysis of data from silage experiment. Prior to  
161 statistical analysis, microbial data (Table 4) were logarithmically transformed. Mean treatment  
162 differences were obtained by Duncan's multiple range tests, with a statistical significance level of 5%.

163



## RESULTS AND DISCUSSION

164

### 165 **Forage quality and yield**

166 Yield and forage quality of experimental forage crops are presented in Table 1. Forage DM  
167 concentration was greatest in proso millet (303 g/kg), intermediate in corn (277 g/kg), and lowest in  
168 sorghum-sudangrass hybrid (193 g/kg). Whole-plant corn had the highest relative feed value (RFV) of  
169 117, which was 20 and 40 units higher on average than proso millet and sorghum-sudangrass hybrid,  
170 respectively. A forage crop with an RFV between 103 and 124 is considered a high-quality forage [21],  
171 indicating the superiority of corn over sorghum-sudangrass hybrid and proso millet forage. Similar to  
172 our observations, Jahansouz et al. [22] also reported a similar trend in fresh forage yield. Concentration  
173 of total digestible nutrients was highest in corn (667 g/kg DM), intermediate with proso millet (631  
174 g/kg DM), and lowest with sorghum-sudangrass hybrid (541 g/kg DM). In general, the forage nutritive  
175 value of proso millet is comparable to the value reported by Kim et al. [23] harvesting “Geumsilchal”  
176 variety in reclaimed lands located in Sihwa (Korea).

177 Forage yield was significantly different by forage types, with proso millet producing the least DM.  
178 The forage DM yield was greater in the order of sorghum-sudangrass hybrid (23.5 t/ha), corn (18.7  
179 t/ha), and proso millet (7.68 t/ha). Forage yield of proso millet (fresh or DM basis) agrees with the  
180 values reported by Shin et al. [24]. Calamai et al. [4] also reported that total dry biomass in proso millet  
181 averaged 6.43 t/ha. Data of NDF and CP concentration of these forage crops is previously reported [8].  
182 Neutral detergent fiber was highest in sorghum-sudangrass hybrid, intermediate in proso millet, and  
183 lowest in corn. No difference existed in CP concentration among crops, averaging 58 g/kg DM.

### 184 **Chemical composition during ensiling**

185 Changes in DM loss and chemical composition of the three forage crops during ensiling are reported  
186 in Table 2. As ensiling progressed, DM loss occurred in all crops, with proso millet losing the most

187 DM than corn or sorghum-sudangrass hybrid, most likely because a higher number of epiphytic molds  
188 existed on proso millet biomass. Loss of DM was faster in proso millet during the first day of  
189 fermentation, which may be justified by the significantly greater population of total microorganisms  
190 in fresh mass of proso millet than in corn or sorghum-sudangrass hybrid (Table 4). Microbial  
191 degradation of nutrients into carbon dioxide and water could possibly explain loss of DM with ensiling  
192 [25, 26]. Crude protein concentration displayed a downward trend during the ensiling process, which  
193 is suggestive of protein degradation with ensiling. A downward trend was also observed in NDF  
194 concentration of all forage crops with ensiling. From day 0 to 45, NDF concentration of proso millet  
195 decreased from 607 to 591 g/kg DM, which is less than the corresponding values in corn and sorghum-  
196 sudangrass hybrid. Chen et al. [26] suggested that hemicellulose degradation during the ensiling  
197 process is mainly responsible for NDF reduction with ensiling. This loss could be due to a combination  
198 of enzymatic and acid hydrolysis of the more digestible cell-wall fractions during the fermentation [10,  
199 27]. After 45 days of ensiling, ADF concentration of proso millet silage declined by about 20 g/kg DM.  
200 Similar decreases also occurred for corn and sorghum-sudangrass hybrid.

### 201 **Fermentation quality during ensiling**

202 Changes in silage pH as a function of fermentation time are illustrated in Figure 2. The day-0 pH  
203 of corn crop (5.80) was generally lower than proso millet or sorghum-sudangrass hybrid (mean 6.05),  
204 which is in agreement with the mean values (5.50 to 6.0) reported for the different forages after  
205 chopping [28, 29]. Silage pH of corn and sorghum-sudangrass hybrid fell rapidly to below 5 within 24  
206 hrs of ensiling, but it took 3 days for proso millet pH to decline below this value. During the late phase  
207 of ensiling, silage pH remained stable and was significantly lower in corn than in proso millet or  
208 sorghum-sudangrass hybrid ( $p < 0.05$ ), possibly due to the higher population of LAB in corn silage  
209 biomass (Table 4). During the 45-day ensiling period, silage pH of corn, proso millet, and sorghum-  
210 sudangrass hybrid decreased by 1.94, 1.65, and 2.04 units, respectively. Buffering capacity, WSC

211 concentration, and moisture level have been identified as critical parameters influencing the  
212 ensilability of forages if epiphytic LAB exist in sufficient numbers [30]. Buffering capacity was lowest  
213 in corn (24.2 mEq/kg DM), intermediate in proso millet (32 mEq/kg DM), and highest in sorghum-  
214 sudangrass hybrid (55.5 mEq/kg DM) [8]. Forages with higher buffering capacity require more acids  
215 for pH reduction. This supports the faster pH reduction in corn plant at the initial phase of ensiling  
216 than proso millet or sorghum-sudangrass hybrid.

217 Time-course of silage ammonia-nitrogen development, expressed as a proportion of total N is  
218 illustrated in Figure 3. Initial NH<sub>3</sub>-N (g/kg total N) level before ensiling was highest in corn (35),  
219 intermediate in proso millet (30), and lowest in sorghum-sudangrass hybrid (14.4). Ammonia-N  
220 concentration increased in three forage crops as ensiling progressed, with proso millet exhibiting the  
221 highest rise. This indicates that protein fractions in proso millet were degraded to a greater extent  
222 during ensiling, perhaps because of accelerated rate of proteolysis and deamination [31]. The NH<sub>3</sub>-N  
223 concentration of less than 70 g/kg total N indicates successful silage fermentation, whereas amounts  
224 greater than 100 g/kg total N have been linked to poor silage fermentation [32]. This criterion indicates  
225 more degradation of protein in proso millet than corn and sorghum-sudangrass hybrid. The rapid  
226 acidification of silage mass is known to inhibit growth and activity of undesirable microorganisms as  
227 well as proteolytic activity [10, 33]. The higher NH<sub>3</sub>-N concentration in proso millet silage could be  
228 attributed to its higher pH during ensiling, which was likely insufficient to effectively suppress  
229 enzymes and microorganisms involved in protein degradation during fermentation.

230 Concentration of WSC in silage mass over the course of the 45-d fermentation is presented in Figure  
231 4. Initial WSC concentration (before ensiling) was higher in proso millet than in corn or sorghum-  
232 sudangrass hybrid (170 vs. mean 141 g/kg DM). An initial WSC concentration between 60 and 80 g/kg  
233 DM has been suggested as an adequate amount to promote an efficient silage fermentation [34]. This  
234 indicates that the forage crops evaluated in this study contained sufficient WSC to promote a good-  
235 quality silage fermentation. The exhaustion of WSC was faster in corn plant as ensiling progressed,

236 reaching a minimum of 6.70 g/kg DM after 3 days of ensiling, after which WSC concentration  
237 decreased slightly until day 45 of ensiling (5.20 g/kg DM). Proso millet experienced a comparatively  
238 slower rate of decline in WSC during ensiling, decreasing to 18.2 g/kg DM on day 15 of ensiling and  
239 reaching a mean value of 5.9 g/kg DM after 45 days of ensiling.

240 During the ensiling fermentation, LAB consume WSC as a readily available source of energy and  
241 primarily convert it to lactic acid, which is associated with silage mass acidification and inhibition of  
242 the activities of undesirable microorganisms [26]. Variations in WSC consumption rates amongst  
243 forage crops during the early phase of ensiling might be ascribed to differences in microbial activity  
244 and plant enzymes in the crops prior to ensiling. In general, WSC supplies the energy required to drive  
245 silage fermentation [35]. A sufficient quantity of WSC has been identified as an important factor in  
246 fast acidification during the initial phase of ensiling, which is associated with DM loss reduction and  
247 improvement of silage quality [10]. In our experiment, the faster reduction of WSC in corn compared  
248 to proso millet forage represented a faster decline in silage pH, which was associated with less DM  
249 loss and NH<sub>3</sub>-N production during ensiling.

250 The IVDMD of the experimental forage crops as a function of ensiling duration are illustrated in  
251 Figure 5. Before ensiling, IVDMD of proso millet and sorghum-sudangrass hybrid was not different,  
252 averaging 643 g/kg DM, which was approximately 16% less than corn (746 g/kg DM). All crops  
253 experienced a decline in IVDMD with ensiling. Previous studies have identified that ADF and NDF  
254 concentrations correlate negatively with IVDMD [36]. This supports findings of the current study  
255 because corn had less NDF and ADF fractions than proso millet or sorghum-sudangrass hybrid,  
256 resulting in the higher digestibility of corn than the other two crops.

### 257 **Organic acids formation during ensiling**

258 Formation of lactic acid and acetic acid as a function of ensiling duration is illustrated in Table 3.  
259 Butyric acid was undetectable during the 45-day ensiling period, which indicates a well-fermented

260 silage and a lack of clostridial activity during ensiling process [10, 26, 29]. High silage pH, typically  
261 greater than 4.5, low DM concentration, and high buffering capacity have been identified as probable  
262 factors which contribute to clostridia growth and proliferation during ensiling [32, 37]. This suggests  
263 that among the forage types evaluated in this experiment, proso millet had a greater susceptibility to  
264 clostridial activity and, thus butyric acid production. However, such an effect was not observed in this  
265 experiment and the absence of butyric acid detection during silage fermentation of proso millet  
266 indicates its low susceptibility to putrefaction by clostridial fermentation.

267 Lactic acid formation increased as silage fermentation progressed, and the magnitude of this  
268 increase was generally greater in sorghum-sudangrass hybrid, intermediate in corn, and lowest in proso  
269 millet. During the 45-day ensiling period, lactic acid concentration displayed an upward trend and  
270 reached a maximum on day 45, with values of 42.5 g/kg DM for proso millet, 67.7 g/kg DM for corn,  
271 and 127 g/kg DM for sorghum-sudangrass hybrid. Lactic acid is typically found in concentrations  
272 ranging from 20 to 40 g/kg DM in commonly used silages [29], which indicates that all forages in the  
273 present experiment underwent an adequate lactic acid fermentation. Similar to lactic acid production,  
274 acetic acid also increased with ensiling, the rate of its production was generally larger in the earlier  
275 phase of silage fermentation. During ensiling process, acetic acid was usually lower in proso millet  
276 than in corn and sorghum-sudangrass hybrid. Acetic acid concentration in sorghum-sudangrass hybrid  
277 reached a maximum concentration of 100 g/kg DM on day 45 of silage fermentation. The higher lactic  
278 acid and acetic acid production in sorghum-sudangrass hybrid during silage fermentation could be  
279 explained by its higher moisture concentration than the other two crops, which accelerates microbial  
280 activity and acid production during the ensiling process. This explanation is supported by the findings  
281 of a previous study identifying that a lower moisture level limits silage fermentation [38]. Although no  
282 consistent trend was seen in lactic acid: acetic acid ratio, there was a general downward trend for each  
283 crop, which is likely indicative of a shift from homo- to hetero-fermentative pattern. This observation  
284 is consistent with results reported by Shao et al. [39, 40]. The higher ratio of lactic acid: acetic acid in

285 corn is most likely suggestive of the dominance of homofermentative LAB during the ensiling process.

#### 286 **Microbial composition during ensiling**

287 Changes in microbial population as a function of ensiling duration are shown in Table 4. The pre-  
288 ensiling population of LAB, mold, and total microorganisms is presented in our companion paper [8].  
289 Briefly, the highest LAB count was detected in corn (6.15 log<sub>10</sub> cfu/g), followed by proso millet (5.91  
290 log<sub>10</sub> cfu/g), and sorghum-sudangrass hybrid (5.88 log<sub>10</sub> cfu/g). An LAB count of 5.0 log<sub>10</sub> cfu/g  
291 biomass has been suggested as a minimum number to enable the dominance of the epiphytic LAB  
292 during ensiling [41, 42]. This suggests that the forage crops had sufficient epiphytic LAB population  
293 to initiate an efficient silage fermentation. Number of mold was highest on proso millet biomass (4.53  
294 log<sub>10</sub> cfu/g fresh mass), which was 0.23 and 1.23 log<sub>10</sub> cfu/g fresh mass greater than corn and sorghum-  
295 sudangrass hybrid, respectively. Forage species, maturity stage, weather, and field wilting have all been  
296 identified as factors causing differences in the population of epiphytic microorganisms in forage crops  
297 [43]. During the 45-day of fermentation, LAB count was generally lower in proso millet than corn or  
298 sorghum-sudangrass hybrid. Number of LAB increased during the early ensiling period and peaked on  
299 day 10 of ensiling. Low pH and the exhaustion of fermentable substrates have been identified as the  
300 primary factors contributing to the decline of LAB population as ensiling proceeds [44].

301 Mold was always present in each crop during fermentation, with a lower number existing on corn  
302 than proso millet or sorghum-sudangrass hybrid. The lower mold population in corn biomass is likely  
303 related to the rapid acidification (lower pH) of corn silage, inhibiting the growth of undesirable  
304 microorganisms [26, 42]. Another factor inhibiting mold growth during ensiling is a high acetic acid  
305 concentration [45]. Less formation of acetic acid and lactic acid (higher pH) during ensiling  
306 fermentation of proso millet could possibly explain the higher mold number in proso millet biomass  
307 during ensiling. The count of total microorganisms was generally higher in corn than in sorghum-  
308 sudangrass hybrid or proso millet. Total microorganisms reached the maximum number on day 10 of

309 ensiling, and then followed a downward trend, which could be explained by pH reduction at this time  
310 point, limiting the growth of microorganisms.

311

## **CONCLUSION**

312 Silage fermentation of proso millet forage resulted in a significant increase in ammonia nitrogen  
313 generation and a larger loss of dry matter when compared to corn or sorghum-sudangrass hybrid,  
314 perhaps because of its higher buffering capacity and silage pH. However, butyrate was undetectable  
315 during its ensiling fermentation. Further research is needed to optimize the fermentation quality of  
316 proso millet forage, possibly by using the appropriate silage additives to minimize ammonia-nitrogen  
317 formation during fermentation, as well as to promote greater lactate production, which is associated  
318 with a further decline in silage pH and mold growth inhibition, and thus with a reduction in dry matter  
319 loss. Despite the lower productivity (less forage production per unit of cultivated land) than corn and  
320 sorghum-sudangrass hybrid, nutrient value of proso millet was comparable to sorghum-sudangrass  
321 hybrid. Proso millet could be harvested in a shorter period of time, making it a potential summer crop  
322 in situations where cultivation of other major summer crops is limited.

323

## **CONFLICT OF INTEREST**

324 We certify that there is no conflict of interest with any financial organization regarding the material  
325 discussed in the manuscript.

326

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479 **Table 1.** Forage yield and forage quality of proso millet, corn, and sorghum-sudangrass hybrid.

Items	Forage type			SEM	<i>p</i> value
	Proso millet	Corn	Sorghum-sudangrass hybrid		
Dry matter, g/kg	303 <sup>a</sup>	277 <sup>b</sup>	193 <sup>c</sup>	11.7	<0.01
TDN, g/kg DM	631 <sup>b</sup>	677 <sup>a</sup>	541 <sup>c</sup>	16.2	<0.01
RFV	97 <sup>b</sup>	117 <sup>a</sup>	77 <sup>c</sup>	7.63	<0.01
Yield, tons/ha					<0.01
Fresh matter	25.4 <sup>c</sup>	67.6 <sup>b</sup>	121.7 <sup>a</sup>	8.97	<0.01
Dry matter	7.69 <sup>c</sup>	18.7 <sup>b</sup>	23.5 <sup>a</sup>	1.41	<0.01

480 <sup>a-c</sup> means with different letter within each row differ (*p* < 0.05). SEM = standard error of mean.

481 TDN = total digestible nutrients. For proso millet and sorghum-sudangrass hybrid, TDN was calculated  
 482 according to the following equation: [889 – (0.79 × ADF, g/kg DM)]. For corn plant, TDN was calculated using  
 483 the following equation: [878.4 – (0.70 × ADF, g/kg DM)] [46].

484 RFV = relative feed value calculated according to the following equation: [(dry matter intake × digestible dry  
 485 matter)/1.29], where dry matter intake = 120/(NDF%) and digestible dry matter = 88.9 – (0.779 × ADF%) [47].  
 486

487 **Table 2.** Dry matter (DM) concentration, DM loss and chemical composition during ensiling. Values were expressed as g/kg DM, unless otherwise  
 488 stated.

Items	Forage type	Ensiling days									SEM
		1	2	3	5	10	15	20	30	45	
DM, g/kg	Proso millet	284.4 <sup>aA</sup>	284.1 <sup>aA</sup>	278.6 <sup>abA</sup>	278.9 <sup>abA</sup>	272.2 <sup>bcA</sup>	270.2 <sup>cA</sup>	269.6 <sup>cA</sup>	266.8 <sup>cA</sup>	266.4 <sup>cA</sup>	3.22
	Corn	275.2 <sup>aB</sup>	274.6 <sup>aB</sup>	274.7 <sup>aA</sup>	272.5 <sup>aA</sup>	272.1 <sup>aA</sup>	271.0 <sup>aA</sup>	268.4 <sup>abA</sup>	264.1 <sup>bcA</sup>	260.7 <sup>cA</sup>	2.82
	Sorghum-sudangrass hybrid	190.3 <sup>aC</sup>	187.5 <sup>abC</sup>	183.1 <sup>abcB</sup>	180.0 <sup>bcB</sup>	174.3 <sup>cB</sup>	177.7 <sup>cB</sup>	176.7 <sup>cB</sup>	178.4 <sup>bcB</sup>	174.7 <sup>cB</sup>	3.82
DM loss	Proso millet	19.0 <sup>cA</sup>	19.30 <sup>cA</sup>	24.8 <sup>bcA</sup>	24.50 <sup>bcA</sup>	31.2 <sup>abA</sup>	33.20 <sup>aA</sup>	33.8 <sup>aA</sup>	36.6 <sup>aA</sup>	37.0 <sup>aA</sup>	2.95
	Corn	2.14 <sup>dB</sup>	2.73 <sup>dB</sup>	2.63 <sup>dB</sup>	4.78 <sup>cdC</sup>	5.20 <sup>cdC</sup>	6.28 <sup>cdC</sup>	8.94 <sup>bcC</sup>	13.2 <sup>abB</sup>	16.6 <sup>aB</sup>	2.11
	Sorghum-sudangrass hybrid	2.50 <sup>dB</sup>	5.30 <sup>dcB</sup>	9.70 <sup>bcC</sup>	12.8 <sup>abB</sup>	18.5 <sup>aB</sup>	15.1 <sup>aB</sup>	16.1 <sup>aB</sup>	15.6 <sup>aB</sup>	18.1 <sup>aB</sup>	2.23
Crude protein	Proso millet	62.3 <sup>aA</sup>	61.0 <sup>abA</sup>	59.8 <sup>abA</sup>	58.3 <sup>abA</sup>	60.3 <sup>abA</sup>	59.9 <sup>abA</sup>	57.9 <sup>bA</sup>	59.6 <sup>abA</sup>	57.1 <sup>bA</sup>	1.46
	Corn	57.4 <sup>aB</sup>	56.5 <sup>aB</sup>	58.4 <sup>aA</sup>	54.6 <sup>abB</sup>	54.5 <sup>abB</sup>	53.40 <sup>abB</sup>	52.7 <sup>abB</sup>	53.0 <sup>abB</sup>	50.8 <sup>bbB</sup>	2.12
	Sorghum-sudangrass hybrid	53.4 <sup>aC</sup>	48.9 <sup>abC</sup>	49.5 <sup>abB</sup>	49.6 <sup>abBC</sup>	48.4 <sup>abC</sup>	46.2 <sup>bcC</sup>	46.6 <sup>bcC</sup>	46.6 <sup>bcC</sup>	46.2 <sup>bcC</sup>	1.99
ADF	Proso millet	324.9 <sup>bbB</sup>	327.3 <sup>bbB</sup>	324.3 <sup>bbB</sup>	342.4 <sup>abB</sup>	344.4 <sup>abB</sup>	340.1 <sup>abB</sup>	347.1 <sup>abB</sup>	330.1 <sup>bbB</sup>	345.8 <sup>abB</sup>	4.76
	Corn	260.5 <sup>aC</sup>	256.2 <sup>abC</sup>	251.5 <sup>bcC</sup>	243.4 <sup>dcC</sup>	248.9 <sup>bcC</sup>	249.2 <sup>bcC</sup>	246.1 <sup>cdC</sup>	241.5 <sup>dcC</sup>	252.0 <sup>bcC</sup>	3.21
	Sorghum-sudangrass hybrid	419.1 <sup>A</sup>	419.7 <sup>A</sup>	420.4 <sup>A</sup>	414.9 <sup>A</sup>	420.5 <sup>A</sup>	422.1 <sup>A</sup>	427.6 <sup>A</sup>	413.2 <sup>A</sup>	415.1 <sup>A</sup>	4.98
NDF	Proso millet	608.5 <sup>abB</sup>	610.8 <sup>abB</sup>	604.3 <sup>abB</sup>	606.6 <sup>abB</sup>	601.8 <sup>abB</sup>	610.5 <sup>abB</sup>	602.5 <sup>abB</sup>	586.0 <sup>bbB</sup>	590.5 <sup>bbB</sup>	6.01
	Corn	496.1 <sup>aC</sup>	491.0 <sup>aC</sup>	467.6 <sup>bcC</sup>	445.3 <sup>cC</sup>	454.4 <sup>cC</sup>	455.8 <sup>cC</sup>	455.6 <sup>cC</sup>	445.5 <sup>cC</sup>	449.2 <sup>cC</sup>	4.37
	Sorghum-sudangrass hybrid	674.7 <sup>aA</sup>	673.1 <sup>aA</sup>	668.0 <sup>aA</sup>	665.0 <sup>aA</sup>	666.2 <sup>aA</sup>	669.8 <sup>aA</sup>	671.7 <sup>aA</sup>	635.0 <sup>baA</sup>	640.1 <sup>baA</sup>	5.45

489 ADF: acid detergent fiber, NDF: neutral detergent fiber. Values with different lowercase letters within each row show significant difference among ensiling days  
 490 with the same forage type. Values with different capital letters within each column show significant differences among forage types in the same ensiling day ( $p$   
 491  $< 0.05$ ).

492 SEM = standard error of mean.

493 **Table 3.** Concentrations of lactic acid and acetic acid as a function of ensiling days.

Organic acids	Forage type	Ensiling days									SEM
		1	2	3	5	10	15	20	30	45	
Lactic acid (LA), g/kg DM	Proso millet	10.1 <sup>eB</sup>	14.1 <sup>deB</sup>	23.2 <sup>cB</sup>	21.6 <sup>cdB</sup>	29.6 <sup>bcC</sup>	21.8 <sup>cdC</sup>	40.0 <sup>aB</sup>	37.0 <sup>abC</sup>	42.5 <sup>aC</sup>	3.64
	Corn	17.6 <sup>eB</sup>	28.6 <sup>dA</sup>	33.6 <sup>dAB</sup>	42.9 <sup>cA</sup>	44.3 <sup>cB</sup>	48.0 <sup>cB</sup>	57.5 <sup>bA</sup>	62.0 <sup>abB</sup>	66.7 <sup>aB</sup>	2.98
	Sorghum-sudangrass hybrid	31.3 <sup>eA</sup>	36.1 <sup>deA</sup>	39.7 <sup>deA</sup>	45.2 <sup>dA</sup>	67.9 <sup>cA</sup>	71.0 <sup>cA</sup>	69.3 <sup>cA</sup>	98.4 <sup>bA</sup>	126.6 <sup>aA</sup>	5.49
Acetic acid (AA), g/kg DM	Proso millet	5.67 <sup>eC</sup>	11.1 <sup>deB</sup>	14.7 <sup>dC</sup>	14.1 <sup>dC</sup>	26.3 <sup>cC</sup>	25.1 <sup>cC</sup>	62.7 <sup>aA</sup>	49.0 <sup>bB</sup>	41.7 <sup>bB</sup>	3.37
	Corn	10.3 <sup>dB</sup>	14.8 <sup>dB</sup>	22.6 <sup>cB</sup>	27.1 <sup>cB</sup>	37.6 <sup>bB</sup>	34.5 <sup>bB</sup>	26.0 <sup>cB</sup>	48.4 <sup>aB</sup>	38.2 <sup>bB</sup>	2.24
	Sorghum-sudangrass hybrid	15.9 <sup>eA</sup>	57.6 <sup>cdA</sup>	49.6 <sup>dA</sup>	63.3 <sup>cA</sup>	54.8 <sup>cdA</sup>	77.5 <sup>bA</sup>	61.6 <sup>cA</sup>	83.2 <sup>bA</sup>	100.3 <sup>aA</sup>	4.63
LA/AA	Proso millet	1.78 <sup>a</sup>	1.27 <sup>bB</sup>	1.58 <sup>abA</sup>	1.53 <sup>abA</sup>	1.13 <sup>b</sup>	0.87 <sup>cdB</sup>	0.64 <sup>dC</sup>	0.76 <sup>dB</sup>	1.02 <sup>bcB</sup>	0.15
	Corn	1.71 <sup>bc</sup>	1.93 <sup>abA</sup>	1.48 <sup>cdA</sup>	1.58 <sup>bcdA</sup>	1.18 <sup>d</sup>	1.39 <sup>cdA</sup>	2.21 <sup>aA</sup>	1.28 <sup>dA</sup>	1.75 <sup>bcA</sup>	0.21
	Sorghum-sudangrass hybrid	1.97 <sup>a</sup>	0.62 <sup>bC</sup>	0.80 <sup>bB</sup>	0.71 <sup>bB</sup>	1.24 <sup>ab</sup>	0.92 <sup>bB</sup>	1.13 <sup>abB</sup>	1.18 <sup>abA</sup>	1.26 <sup>abB</sup>	0.45

494 Values with different lowercase letters within each row show significant difference among ensiling days with the same forage type. Values with different capital  
 495 letters within each column show significant differences among forage types in the same ensiling day ( $p < 0.05$ ).

496 SEM = standard error of mean.

497

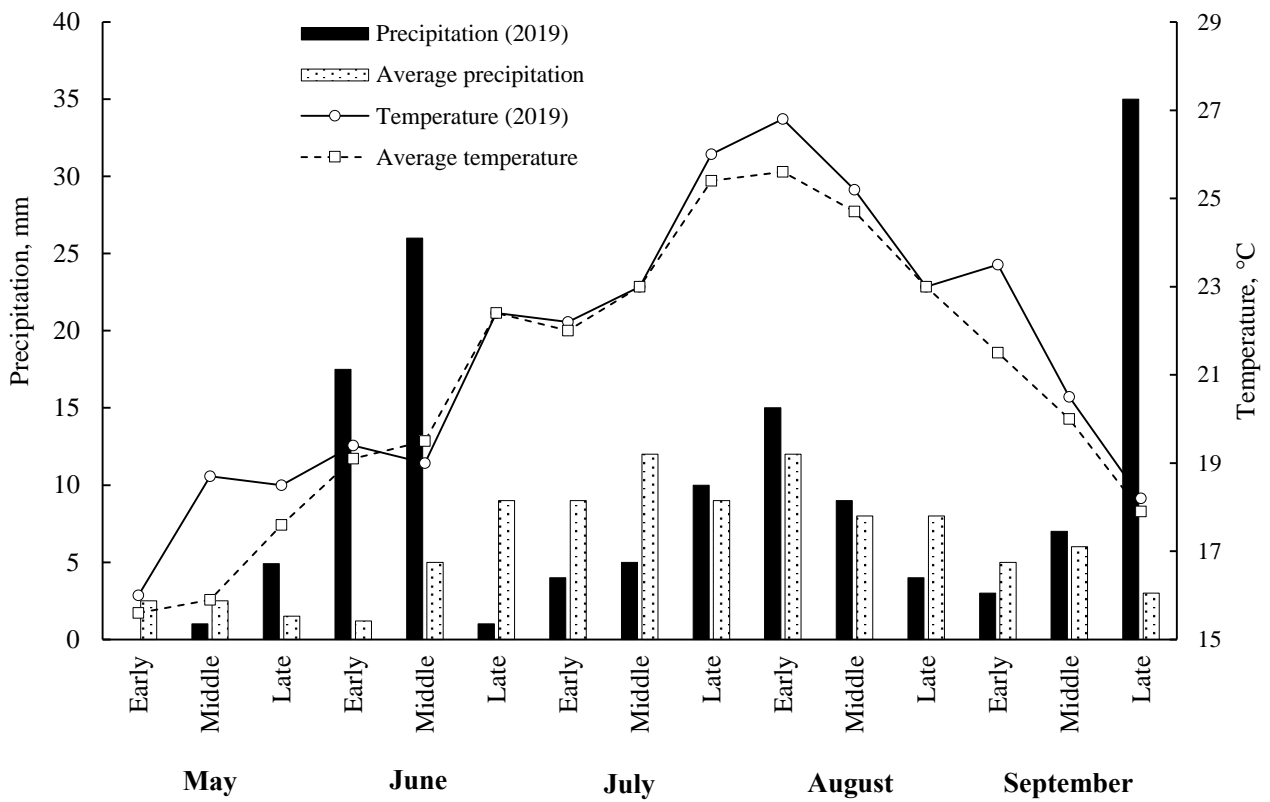
498 **Table 4.** Number of lactic acid bacteria, mold and total microorganisms as a function of ensiling days.

Microbial count <sup>1</sup>	Forage type	Ensiling days									SEM
		1	2	3	5	10	15	20	30	45	
Lactic acid bacteria	Proso millet	6.48 <sup>bb</sup>	6.88 <sup>aa</sup>	6.94 <sup>ab</sup>	6.93 <sup>ab</sup>	6.96 <sup>ab</sup>	6.48 <sup>bb</sup>	5.78 <sup>cc</sup>	5.78 <sup>cb</sup>	5.34 <sup>db</sup>	0.11
	Corn	6.84 <sup>fa</sup>	6.85 <sup>fa</sup>	7.23 <sup>cdA</sup>	7.30 <sup>ca</sup>	7.77 <sup>aa</sup>	7.61 <sup>abA</sup>	7.04 <sup>ea</sup>	6.60 <sup>ga</sup>	6.08 <sup>ha</sup>	0.08
	Sorghum-sudangrass hybrid	5.08 <sup>dc</sup>	5.68 <sup>cb</sup>	6.60 <sup>bc</sup>	6.53 <sup>bc</sup>	6.95 <sup>ab</sup>	6.60 <sup>bb</sup>	6.95 <sup>ab</sup>	6.62 <sup>ba</sup>	5.89 <sup>ca</sup>	0.12
Molds	Proso millet	3.49 <sup>da</sup>	4.21 <sup>ca</sup>	4.30 <sup>bcA</sup>	4.20 <sup>ca</sup>	5.00 <sup>abA</sup>	5.38 <sup>aa</sup>	4.34 <sup>bcA</sup>	4.04 <sup>cb</sup>	4.80 <sup>ba</sup>	0.30
	Corn	3.18 <sup>cb</sup>	3.00 <sup>cc</sup>	4.00 <sup>bb</sup>	3.00 <sup>cc</sup>	4.05 <sup>bb</sup>	4.00 <sup>bc</sup>	3.85 <sup>bb</sup>	4.67 <sup>aa</sup>	4.18 <sup>abB</sup>	0.41
	Sorghum-sudangrass hybrid	3.48 <sup>cdA</sup>	3.30 <sup>db</sup>	3.85 <sup>bb</sup>	3.70 <sup>bcB</sup>	3.29 <sup>dc</sup>	5.11 <sup>ab</sup>	3.60 <sup>bcC</sup>	3.31 <sup>dc</sup>	3.00 <sup>ec</sup>	0.13
Total microorganisms	Proso millet	7.43 <sup>ba</sup>	7.51 <sup>ba</sup>	7.44 <sup>ba</sup>	7.79 <sup>ab</sup>	7.86 <sup>ab</sup>	7.26 <sup>bcA</sup>	7.04 <sup>cb</sup>	6.30 <sup>dc</sup>	6.60 <sup>eb</sup>	0.10
	Corn	7.05 <sup>cb</sup>	7.29 <sup>ca</sup>	7.18 <sup>cb</sup>	8.10 <sup>ba</sup>	8.85 <sup>aa</sup>	7.11 <sup>caB</sup>	7.85 <sup>ba</sup>	7.04 <sup>cb</sup>	7.12 <sup>ca</sup>	0.19
	Sorghum-sudangrass hybrid	6.57 <sup>cc</sup>	6.88 <sup>bb</sup>	6.48 <sup>cc</sup>	7.01 <sup>bc</sup>	7.40 <sup>ac</sup>	6.95 <sup>bb</sup>	7.04 <sup>bb</sup>	7.32 <sup>aa</sup>	6.51 <sup>cb</sup>	0.08

499 <sup>1</sup> Microbial count was expressed as the logarithmic number of colony-forming units per gram fresh mass. Values with different lowercase letters within each  
500 row show significant difference among ensiling days with the same forage type. Values with different capital letters within each column show significant  
501 differences among forage types in the same ensiling day ( $p < 0.05$ ).

502 SEM = standard error of mean.



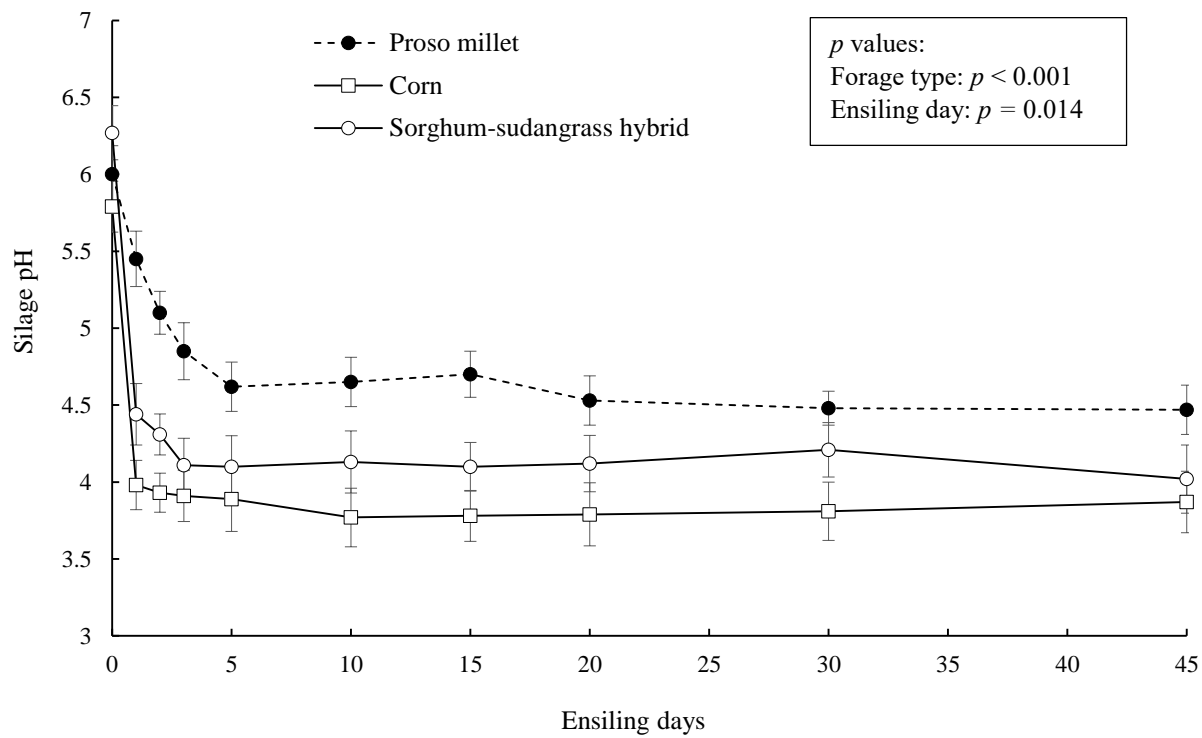


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505

506 **Figure 1.** Temperature and precipitation during the growing season (May to September, 2019) and  
 507 comparison with the average climatic normal. The data were obtained from the Korean Meteorological  
 508 Administration.

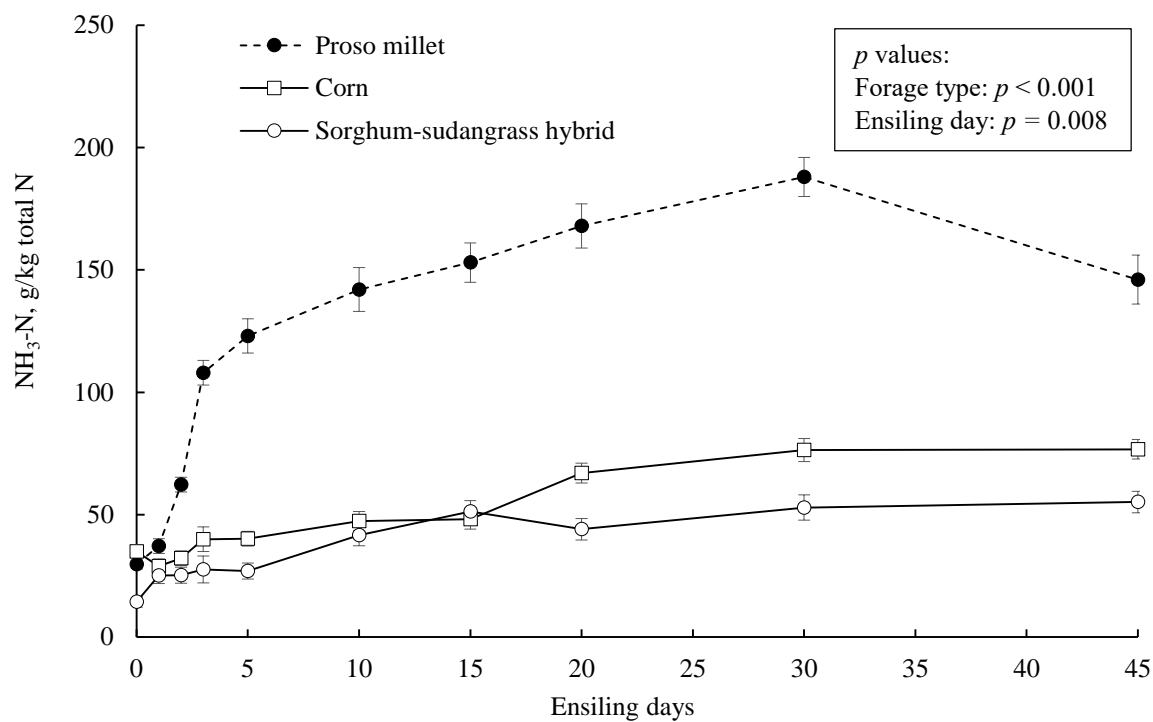
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510

511 **Figure 2.** The pH value of proso millet, corn, and sorghum-sudangrass hybrid as a function of ensiling  
 512 days. Bars indicate standard error.

513

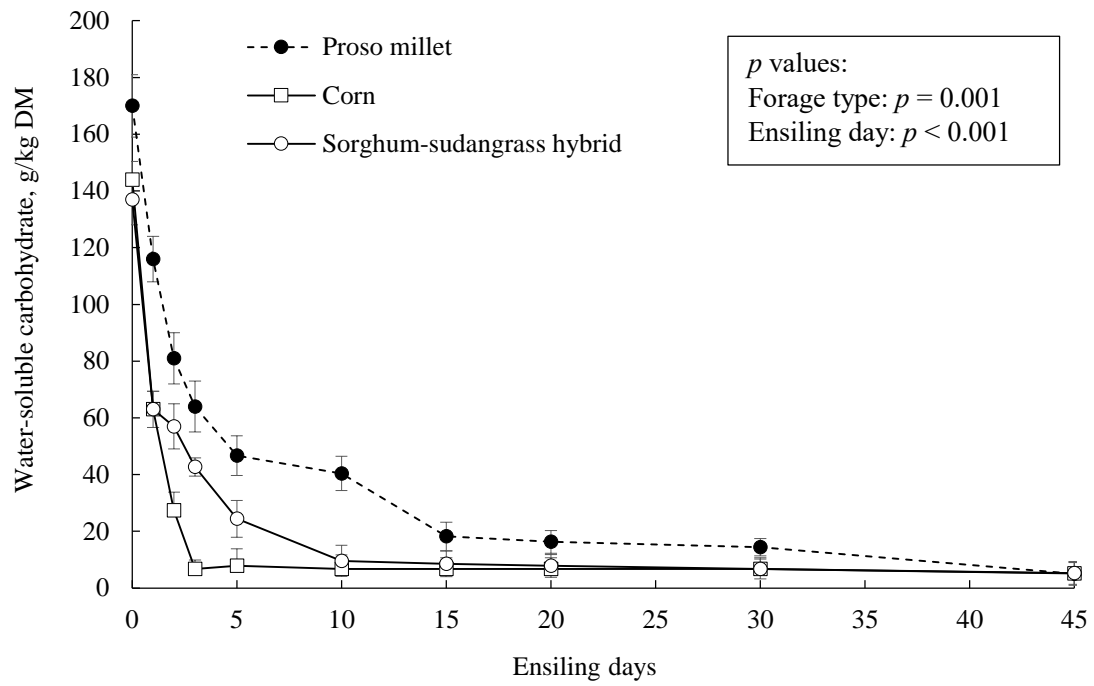


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516

517 **Figure 3.** Ammonia-nitrogen concentration of proso millet, corn, and sorghum-sudangrass hybrid as a  
 518 function of ensiling days. Bars indicate standard error.

519

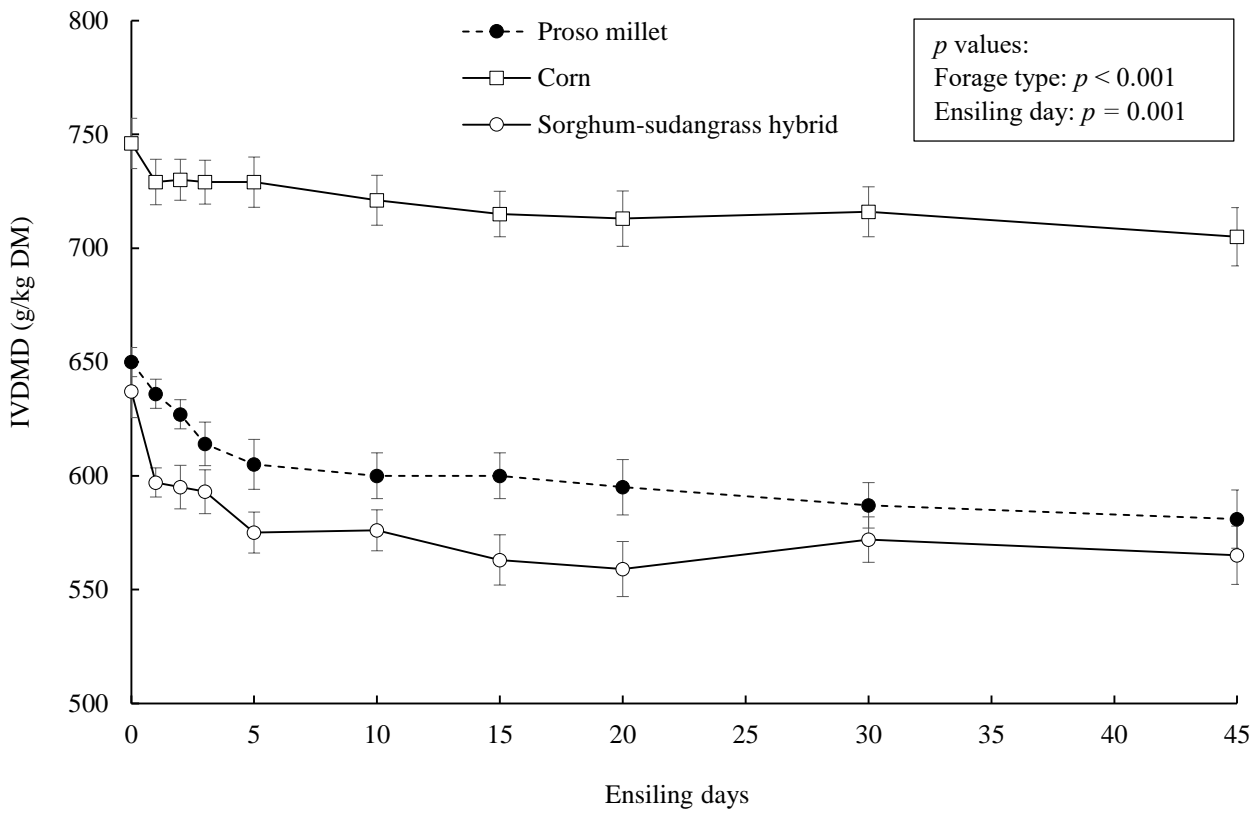


521

522 **Figure 4.** Water-soluble carbohydrate concentration of proso millet, corn, and sorghum-sudangrass  
 523 hybrid as a function of ensiling days. Bars indicate standard error.

524

525



526

527

528 **Figure 5.** *In vitro* dry matter digestibility (IVDMD) of proso millet, corn, and sorghum-sudangrass  
529 hybrid as a function of ensiling days. Bars indicate standard error.