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8     **Abstract**

9     The objective of this experiment was to evaluate the physiochemical characteristics of three tertiary  
10    hybrids (crossbreeds) of pigs, with and without coffee supplementation. A total of fifty pigs of  
11    different mixed breeds Landrace × Yorkshire × Duroc (LYD), Yorkshire × Berkshire (YB), and  
12    Yorkshire × Woori (YW); 113.45 kg ±3.33 kg) at age 190 days old were employed to measure the  
13    effect of spent coffee grounds from Gangneung-Si area of South Korea on the meat quality of pigs in  
14    the pigsty at the Kangwon National University Teaching and Research Farm using the 2 × 2 factorial  
15    arrangements. Our result shows that the fat percentage was higher (P<0.05) in YB and YW. pH was  
16    higher (p<0.05) in the YB breed. Meat colour a\* was higher (p<0.05) in the YB and YW breeds. Meat  
17    colour b\* was higher (p<0.05) in YW. Water holding capacity was higher (p<0.05) in the YB and YW  
18    breeds. Drip loss was lower (p<0.05) in YB and YW. Cooking loss was higher (p<0.05) in LYD and  
19    YW breeds. The fatty acid components such as linolenic (C18:2), myristic (C14:0), and palmitoleic  
20    (C16:1) were higher (p<0.05) in the YB. Palmitic (C16:0), stearic (C18:0), and arachidic (C20:0) was  
21    higher (p<0.05) in YW. Lignoceric (C24:0) was higher (p<0.05) in LYD and YW. Unsaturated fatty  
22    acid (UFA) was higher (p<0.05) in YB and YW, while Polyunsaturated fatty acid (PUFA) was higher  
23    (p<0.05) in YB. Monosaturated fatty acid (MUFA) / PUFA was higher (p<0.05) in LYD. Saturated  
24    fatty acid (SFA) was higher (p<0.05) in YW. UFA and MUFA were higher (p<0.05) in the YB.  
25    MUFA / PUFA were higher (p=0.05) in YB. We concluded from our results that YW and YB had  
26    close meat qualities in terms of firmness and flavour compared to LYD as the physiochemical  
27    characteristics of meat were improved. SCG supplemented at 0.5% had no detrimental effect on the  
28    parameters measured.

29

30

31    **Keywords:**

32    Korean Woori pig, coffee waste, water holding capacity, meat quality, fat firmness, cooking loss

33

34

## Introduction

35

36 Coffee is the most common global beverage after water, with millions of tons being produced  
37 worldwide [1]. However, coffee residues and by-products are major environmental contaminants,  
38 especially in regions where coffee is produced in large amounts. From the 20<sup>th</sup> century till date, many  
39 efforts have been made to develop viable ways of converting coffee waste into other valuable  
40 products, such as feeds, biogas, pectic enzymes, and proteins [2]. Extensive research has also been  
41 conducted to investigate the possible benefits of using coffee residue.

42 One of the primary by-products of coffee is spent coffee grounds (SCG), which are derived from  
43 brewing coffee seeds. Manufacturing 1 kilogram (kg) of green coffee generates approximately 0.65 kg  
44 of SCG, whereas producing 1 kg of instant brewed coffee generates approximately 2 kg of wet SCG  
45 [3]. Using coffee by-products, especially SCG, as an animal feed source was initially unpopular  
46 because of their poor starch quality, which is comparable to that of low-quality hay. However, a  
47 growing body of research has examined its application as a dietary additive for swine, poultry, and  
48 other livestock because of its constant availability, high volume, and low price [3]. For instance,  
49 coffee by-products may be used as livestock feed supplements to reduce the cost of production. In  
50 particular, SCG has been found to be a viable source of protein and lipids, and it contains minute  
51 quantities of polyunsaturated fatty acids [4]. However, SCG is an unpalatable by-product of instant  
52 coffee production and contains diuretics, tannins, and caffeine. Therefore, SCG supplements should  
53 not exceed 2.5% of any feed [5]. Using by-products from coffee in various farm animal diets may  
54 help reduce the cost of production and waste management; however, the effect of SCG on the meat  
55 quality of different pig breeds has not been extensively studied. The consumption of pork is  
56 substantially increasing in South Korea. However, the Duroc (D) and other popular breeds that are  
57 commonly used as breeding stock are sourced from the United States of America and Canada.  
58 However, in most cases, this results in high foreign exchange costs and other expenses for breeding  
59 companies [6]. To reduce these costs, it is essential to develop other breeds, including local Korean  
60 pig breeds. It is also important to note that breed plays a significant role in meat quality [7], and  
61 creating high-quality pork is crucial for boosting revenues in the pig industry [8]. Landrace (L),  
62 Yorkshire (Y), and D three-way hybrids (crossbreeds) (LYD) are frequently used in Korea [9]. Due to  
63 their large litter size, rapid growth, and high meat production, L pigs are frequently employed as  
64 fattening pigs, whereas D pigs have an excellent growth rate and high fat content. However,  
65 crossbreeds primarily developed for meat production, such as LYD, may produce pork of poor quality,  
66 with unstable fat firmness and poor water-holding capacity (WHC) [10]. Traditional Korean pig  
67 breeds have better meat qualities, such as firm fat tissue, good texture, and a unique flavour that can  
68 please Korean customers, but they have low economic value because of their low feed efficiency,  
69 growth rate, and output rate [11].

70 Therefore, the purpose of the current study was to evaluate the digestibility, antioxidant activity, meat  
71 colour, WHC, pH, cooking loss (CL), cooking moisture, drip loss (DL), and various fatty acid

72 characteristics of the belly and loin of different crossbreeds of pigs when fed diets supplemented with  
73 and without SCG.

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75

76

## Materials and Methods

77 The Institutional Animal Care and Use Committee of Kangwon National University approved the  
78 animal care and experimental techniques utilized in this study (Ethical code: KW- 220413-1)

79

### 80 Test animals, feed, and experimental design

81 A total of fifty pigs of different mixed breeds LYD, Yorkshire × Berkshire (YB), and Yorkshire ×  
82 Woori (YW); 113.45 kg ±3.33 kg) at age 190 days old were employed to measure the effect of coffee  
83 extract from Gangneung-Si South Korea on the meat quality of pigs in the pigsty at the Kangwon  
84 National University Teaching and Research Farm, and pigs with good conformity were selected for  
85 this experiment. The corn-soybean feed utilized was made available *ad libitum* and designed to meet  
86 or surpass the National Research Council's nutrient standards [12] and was supplied in a powdered  
87 form (Table 1). 2 × 2 factorial arrangements were used with a section containing the breeds without  
88 coffee LYD, YB, and YW, and breeds YB and YW with coffee, with the experiment containing ten  
89 pigs per treatment and one pig per replicate. The SCG with nutrient composition as shown in Table 2  
90 was oven dried at 105°C for 24h to reduce the moisture content. It was then stored in airtight bags at  
91 room temperature and eventually premixed at 0.5% per 20 kg of the corn-soybean feed.

92 All test animals were killed and used fresh directly from the processing unit. The left side of the  
93 carcasses' loins and bellies were used to gauge several parameters of meat quality. The loins and  
94 bellies were used for all samples, and the muscles' extra fat and bone were removed.

95

### 96 Proximate analysis of pork

97 The proximate analysis was examined following the AOAC 2012 [13] standard. Weight loss after 12  
98 hours at 105°C in a drying oven was used to determine the moisture content (AOAC method  
99 950.46B) (SW-90D, Sang Woo Scientific Co, Korea). The Soxhlet model was used to evaluate the fat  
100 content (AOAC method 960.69) by employing a solvent evaporator system (Soxtec® Avanti 2050

101 Auto System, Foss Tecator AB, Sweden), and the Kjeldahl nitrogen analyzer (Kjeltec® 2300  
102 Analyzer Unit, Foss Tecator AB, Sweden) was used to determine the protein content (AOAC method  
103 981.10).

104

#### 105 **Antioxidant activity**

106 The samples were pre-treated and examined following the Cayman kit handbook (Enzyme activity  
107 assay, Cayman Chemical, Ann Arbor, MI, USA). Belly and loin samples were analyzed for the  
108 concentrations of MDA (Cat. #10006438, Cayman Chemical, Ann Arbor, MI, USA). According to the  
109 instructions in the Cayman kit's manufacturer's manual, a microplate reader (Power Wave XS,  
110 BIoTeK, Winooski, VT, USA) was employed to examine the absorption detection [14].

111

#### 112 **Meat quality**

##### 113 **Meat colour**

114 In order to measure the meat's colour, the belly and loin muscles were separated from the skin, tendon,  
115 and fat. A Chroma Meter CR-400 device (Minolta Co, Osaka, Japan) was used to measure the colour  
116 in line with the CIE L\* (lightness), a\* (redness), and b\* standards (yellowness) which is the  
117 International Commission on Illumination Standards.

118

##### 119 **Water holding capacity (WHC in %)**

120 The evaluation of WHC was done by depositing 0.5 g of the specimen on a round plastic plate in a  
121 tube (Millipore Ultra free-MC; Millipore, Bedford, MA). The specimens were then subjected to a  
122 temperature of 80°C in a water bath for 20 minutes and then cooled afterward to 23°C. The specimen  
123 was then centrifuged (2000 g) for 10 minutes at 4°C to determine WHC using the formula,  $WHC =$   
124  $(\text{moisture content} - \text{water loss}) / \text{moisture content} \times 100$

125

##### 126 **pH**

127 A 5 g sample of pork belly and loin was homogenised in 45 mL of distilled water using a DIAX 900,  
128 Heidolph, Kelheim, Germany homogenizer for 15 seconds. The pH meter (Orion 230A, Thermo

129 Fisher Scientific, Waltham, MA, USA) was then used to measure the pH level, and the homogenized  
130 samples were filtered using Watman no. 2 (Hillsboro, OH, USA) after the pH procedure

131

132

### 133 **Cooking loss (%)**

134 3 g of pork belly and loin were heated to 85°C for 20 minutes in a water bath using an airtight  
135 polyethylene container before being allowed to cool to 25°C. Using the formula (sample weight  
136 before cooking/sample weight after cooking)/sample weight before cooking × 100, we calculated the  
137 CL afterward.

138

### 139 **Drip loss**

140 At 24 hours postmortem, belly and loin samples were cut and weighed immediately, then recorded as  
141 the initial drip loss weight. The samples were kept in netting and hung in a pressurized bag to prevent  
142 subsequent contact between the samples and the bag. The loin and belly samples were gently removed  
143 from the bag and allowed to dry after 6-, 12-, 24-, and 48-hours postmortem period respectively. Drip  
144 loss was calculated as the percentage of the final sample weight following dripping over the initial  
145 sample weight for the different hours as stated above.

146

### 147 **Fatty acid analysis (%)**

148 Following the methodology of Folch et al. [15], total fat was examined by adding 5 g of lipids from  
149 the samples to a solution of chloroform/methanol in (2:1) with butylated hydroxytoluene. A potassium  
150 hydroxide methanol solution was used to create fatty acid methyl esters (FAMES), which were then  
151 extracted using water and hexane. Anhydrous Na<sub>2</sub>SO<sub>4</sub> was used to dehydrate the top hexane layer that  
152 contained FAMES. The extracted hexane was then dried and transferred into a vial for testing. Gas  
153 chromatography (Agilent 7890N, Agilent Technologies, Korea) with a flame ionisation detector and  
154 capillary column (30 m, 0.32 mm id, 0.25 μm, Omega wax 320, Supelco, USA) was used to separate  
155 and measure FAMES via an inlet with a 100:1 split ratio. The carrier gas was high-purity nitrogen, and  
156 the flow rate was 1 ml/min. After holding the oven temperature at 180°C for five minutes, it was

157 raised to 200°C at a rate of 2.5°C/min and maintained for 25 minutes. Temperatures for the injector  
158 and detector were 25°C and 26°C, respectively.

159

## 160 **Statistical method**

161 The data were accumulated using a 2 × 2 factorial arrangement in a completely randomised design  
162 which was analysed using the statistical analysis system (SAS) and general linear model (GLM) (SAS  
163 Inst. Inc., Cary, NC, USA). The main effects for differentiating treatments were LYD, YB, and YW,  
164 with the parameters, treatments, and individual pigs serving as the repeated experimental unit. The  
165 Tukey test was employed for post hoc testing, and the disparity was considered statistically significant  
166 when the *p*-value was less than 0.05 ( $p < 0.05$ ) in the experimental units

167

## 168 **Results**

### 169 **Proximate analysis and MDA**

170 Table 3 shows a comparison of the belly and loin of different breeds of pigs fed diets with and  
171 without SCG supplementation. In the belly, the fat percentage was higher ( $p < 0.05$ ) in the YB breed  
172 than in the LYD and YW breeds. In the loin, the fat percentage was higher ( $p = 0.050$ ) in the YW breed  
173 than in the LYD breed. There were no significant differences in protein, moisture, and MDA across  
174 the breeds. There were also no significant effects on the measured parameters for the YB and YW  
175 breeds when their diets were supplemented with SCG.

176

### 177 **pH and meat colour**

178 The pH and meat colour in the belly and loin are shown in Table 4 of our study. In the belly, the pH  
179 was higher ( $p < 0.05$ ) in the YB than in the YW breed. Meat colour  $a^*$  was higher ( $p < 0.05$ ) in the YB  
180 and YW breeds compared to that in the LYD breed. There were no significant differences in meat  
181 colour ( $L^*$  and  $b^*$ ) in the belly. In the loin, the pH was higher ( $p < 0.05$ ) in the YB breed than in the  
182 LYD breed. Meat colour  $b^*$  was higher in the YW group ( $p < 0.05$ ) than in the YB group. There was  
183 no significant difference in meat colour,  $L^*$ , and  $a^*$ , in the loins. There were also no significant effects  
184 on the measured parameters for the YB and YW breeds when their diets were supplemented with  
185 SCG.

186

### 187 **Meat quality**

188 Table 5 shows the result for meat quality in our study. In the belly, the WHC was higher ( $p < 0.05$ ) in  
189 the YB and YW breeds than in the LYD breed. The DL6 was lower ( $p < 0.05$ ) in the YB and YW  
190 breeds than in the LYD breed. Among the breeds, there were no significant differences in the CL, DL  
191 12, 24, and 48 in the belly. In the loin, the CL was higher ( $p < 0.05$ ) in the LYD and YW breeds than in  
192 the YB breed. There were no significant differences in WHC, DL6, 12, 24, and 48 in the loin. There

193 were also no significant effects on the measured parameters for the YB and YW breeds when their  
194 diets were supplemented with SCG.

195

### 196 **Fatty acid composition**

197 The fatty acid composition was shown in Table 5 and Table 6 of our experiment. In the belly, the  
198 linolenic (C18:2) content was higher ( $p<0.05$ ) in the YB than in the LYD breed. However, there were  
199 no significant differences in belly lauric (C12:0), myristic (C14:0), palmitic (C16:0), palmitoleic  
200 (C16:1), stearic (C18:0), oleic (C18:1), linolenic (C18:3), arachidic (C20:0), or lignoceric (C24:0)  
201 content across breeds. In the loin, the C14:0 content was higher ( $p<0.05$ ) in the YB breed than in the  
202 YW or LYD breeds. The C16:0 content was higher ( $p<0.05$ ) in the YW breed than in the LYD or YB  
203 breeds. The C16:1 content was higher ( $p<0.05$ ) in the YB breed than in the YW and LYD breeds. The  
204 C18:0 content was significantly higher ( $p<0.05$ ) in the YW breed than in the YB breed. The C20:0  
205 content was higher ( $p<0.05$ ) in the YW breed than in the LYD or YB breeds. The C24:0 levels were  
206 higher ( $p<0.05$ ) in the LYD and YW breeds than in the YB breed. The levels of unsaturated fatty  
207 acids (UFA) were higher ( $p<0.05$ ) in the YB and YW breeds than in the LYD breed (Table 5). The  
208 levels of polyunsaturated fatty acids (PUFA) were higher ( $p<0.05$ ) in the YB breed than in the LYD  
209 breed. The monosaturated fatty acid (MUFA) / PUFA ratio was higher ( $p<0.05$ ) in the LYD breed  
210 than in the YW and YB breeds. There were no significant differences in the levels of saturated fatty  
211 acids (SFA) and MUFAs in the belly across the breeds. In the loin, the levels of SFA were higher  
212 ( $p<0.05$ ) in the YW breed than in the LYD or YB breeds. The levels of UFA and MUFA were higher  
213 in the YB breed than in the LYD and YW breeds. In addition, the levels of MUFA and PUFA were  
214 higher ( $p<0.05$ ) in the YB breed than in the YW breed. Finally, there was no significant difference in  
215 the PUFA content of the loins when all breeds were compared. There were also no significant effects  
216 on the measured parameters for the YB and YW breeds when their diets were supplemented with  
217 SCG.

218

### 219 **Discussion**

220 In our study, the fat content responsible for meat firmness and flavour was higher in the YB and YW  
221 breeds compared with LYD. Certain fats and fatty acids contribute immensely to several aspects of  
222 meat quality. They are essential to the nutritional content and juiciness of meat, although they have  
223 been reported to be harmful by several studies based solely on their nature and the category of fat to  
224 which they belong [16,17]. The firmness and taste of pork are primarily determined by its moisture  
225 content and adipose tissue composition, which is important because different fats have different  
226 melting points. Our results are similar to those of Kim et al. [18] who found that the fats responsible  
227 for firmness and flavour were higher in the Korean native Woori breed than in the LYD breed. The fat  
228 content in the YW and YB breeds is thought to be due to the comparatively high intramuscular fat  
229 content of the native Korean pig and the YB breed [19,20].

230 Pork is naturally acidic, and its pH affects the interaction of proteins in fresh and processed pork [21],  
231 Thus, pH is a fundamental component of meat quality. Factors such as breed, nutrition, and  
232 management affect the final pH of meat [22,23]. Several protein characteristics, such as solubility,  
233 function, colour, and water-binding capacity, are directly affected by pH [24], and extremely low pH  
234 values usually have a negative impact on these qualities [25]. In our study, the YB breed had the  
235 highest pH values in the belly and loin compared to the YW breed (in the belly) and the LYD breed  
236 (in the loin), which could explain the increase in  $a^*$  in the YB breed. Lonergan et al. (2008) reported  
237 that the pH in the muscle and other denatured meat drops rapidly and encourages the production of  
238 sarcoplasmic protein, a water-soluble protein in the muscle that can impact meat colour and WHC.  
239 However, this process may be slowed if the meat is chilled. Preventing the denaturation of myoglobin,  
240 which gives meat its colour, is crucial because consumer preference is related to meat colour. It has  
241 also been found that a lighter colour results from a reduced final pH and vice versa [24,26]. This may  
242 explain the high  $a^*$  meat colour in the belly of the YB breed and the  $b^*$  meat colour seen in the YW  
243 breeds. As previously mentioned, pH also greatly contributes to the WHC, which determines the meat  
244 quality parameters DL and CL. The ability of pork to bind water is due to the activity of several  
245 proteins [27].

246 The two myofibrillar proteins actin and myosin are water-binding proteins that are affected by pH  
247 (Choi et al., 2019). In our study, we found that the WHC was higher in the bellies of the YB and YW  
248 breeds, which had a lower DL6. However, the WHC for the YB breed was 72.36 with a higher belly  
249 pH, whereas that for the YW breed was 70.53 with a lower belly pH. The YB breed had a higher belly  
250 pH, and its DL6 was as low as 1.14. The YW breed had a higher belly pH, and its DL6 was 1.48. The  
251 reduced CL in the YB loin can be attributed to its increased pH, suggesting that a combination of high  
252 temperatures and low pH can trigger the denaturation of myosin [21]. Denatured myosin is less  
253 functional as a result of minute water-binding protein availability. Although the pH values for the  
254 bellies in the YB and YW breeds in our study were almost incomparable, they had similar red meat  
255 colours, WHC, and DL. These attributes in the YW breed can be ascribed to the ability of native  
256 Korean pigs to produce higher levels of myosin and to the configuration of their firm and red muscle  
257 fibres. A higher myoglobin concentration and the presence of enough fat to serve as an energy source  
258 for the muscle cells may lead to a higher WHC, red colour, and reduced DL6 [28,29].

259 Fats and fatty acids are essential components of the pork nutritional profile and substantially  
260 contribute to the practical aspects of meat quality and human health after consumption [16].  
261 Consuming high quantities of SFA, such as C14:0, C16:0, and C20:0, can increase the risk of  
262 developing type 2 diabetes and cardiovascular disease. Therefore, they should be moderately ingested  
263 [30-32]. We found that C14:0 levels were higher in YB loins, whereas C16:0 and C20:0 levels were  
264 higher in YW loins. In our study, we found that C18:0, which has the greatest impact on meat  
265 firmness, was significantly higher in the loin. This concentration correlates strongly with the melting  
266 temperature of lipids and the firmness of carcass fat, which has a melting point of approximately 70°C  
267 [33]. Although C18:0 is regarded as a neutral fatty acid, it increases the level of high-density

268 lipoprotein cholesterol and reduces that of low-density lipoprotein (LDL) cholesterol, which makes it  
269 harder for the body to produce more cholesterol [34]. This process leads to the absorption, transport,  
270 and removal of cholesterol from the body via the liver. Many UFAs, especially omega-3 fatty acids  
271 such as C16:1, are also beneficial to human health and are linked to the flavour and general  
272 desirability of meat [35,36]. Consuming UFAs helps prevent disorders such as arteriosclerosis and  
273 hypertension, and they are so abundant in meat that they affect its flavour [18]. In our study, C16:1  
274 was higher in the loin of the YB breed, whereas UFA levels were higher in the belly of the YB and  
275 YW breeds. In the YB breed, UFA levels were highest in the loin. Essential fatty acids, such as C18:2,  
276 also have a positive impact on the heart when they are consumed. Several clinical trials have shown  
277 that C18:2 decreases both total and LDL cholesterol levels when it was substituted for saturated fat.  
278 The present study found that C18:2 levels were higher in the YB breed. Another group of significant  
279 fatty acids that contributes immensely to the taste of pork are MUFAs; however, an increase in  
280 MUFAs is accompanied by a proportional decrease in PUFAs [11]. However, PUFAs have been  
281 reported to beneficially affect human and animal health [37,38]. Diets rich in PUFAs may reduce  
282 inflammation and lower the risk of chronic diseases, such as arthritis, cancer, and heart disease. Our  
283 study showed that PUFA levels were higher in YB and YW bellies, whereas MUFA levels were  
284 higher in YB loins. Increases in MUFA and PUFA levels are related to changes in fat oxidation,  
285 softness, texture, and rancidity. All of these result from the proportional increase in the double bonds  
286 of fatty acids, and they can lead to a decline in the melting point and oxidative stability of fats [39].  
287 Our study showed that MUFA and PUFA levels were higher in the belly of the LYD breed, whereas  
288 MUFA levels were higher in the loin of the YB breed.

289 In addition to the qualities of breeds stated above, the supplementation of SCG had no significant  
290 effect on all parameters measured. The level of responsiveness by SCG is greatly proportional to the  
291 administered dosage, thus, it is paramount to establish the most ideal level of SCG supplementation in  
292 diets [40]. We, therefore, propose that the inability of coffee to significantly improve the meat quality  
293 of pork in our study may be attributed to the dosage employed. However, our result suggests that the  
294 supplementation of SCG up to 0.5% had no detrimental effect on the pig's meat quality.

295

## 296 **Conclusion**

297 Our result confirmed that YW and YB had similar meat qualities rather than LYD in terms of pork fat  
298 firmness and flavour as the physiochemical characteristics of meats including higher unsaturated fat  
299 composition, pH, meat redness, WHC, and reduced drip loss which ultimately improves pork firmness,  
300 texture, consumer-preferred colour, and flavour were observed. It was also noticed that the  
301 supplementation of SCG at 0.5% had no detrimental effect on the parameters measured.

302 We, therefore, recommend that employing the Korean Woori pig as a breeding alternative can help  
303 ameliorate the dependence on exotic breeds including Landrace, Berkshire and Duroc in terms of  
304 meat quality such as pork fat firmness, colour, and flavour. We also recommend that SCG can be  
305 supplemented in finishing pig's diet up to 0.5% with no adverse effects.

306

307

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ACCEPTED

Table 1. Ingredient and chemical composition of basal diets (as-fed).

Item	CON
Ingredient, %	
Corn	74.43
Soybean meal (44%)	20.29
Beef tallow	0.23
Molasses (sugar beet)	3.00
L-Lysine (78.8%)	0.05
Limestone	0.70
DCP	0.60
Salt	0.30
Choline	0.05
Mineral	0.15
Vitamin	0.15
Phytase	0.05
Total	100.0
ME	3300
CP	14.00
Ca	0.54
P(Total)	0.47
P(STTD)	0.24
Lys	0.73
Met	0.23
MetCys	0.47
Phe	0.69
Thr	0.48
Trp	0.15
Val	0.66

Supplied per kilogram of diet: 8000 IU vitamin A, 1500 IU vitamin D3, 16 mg vitamin E, 1.0 mg vitamin B1, 8.0 mg vitamin B2, 1.6 mg vitamin B6, 0.03 mg vitamin B12, 1.0 mg vitamin K3, 16 mg pantothenic acid, 30 mg niacin, 0.06 mg biotin, 0.26 mg folic acid and 4.8 mg ethoxyquin.

Supplied per kilogram of diet: 150 mg Fe as ferrous sulfate, 96 mg Cu as copper sulfate, 72 mg Zn as zinc sulfate, 46.5 mg Mn as manganese sulfate, 0.9 mg I as calcium iodate, 0.9 mg Co as cobalt sulfate and 0.3 mg Se as sodium selenite.

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Table 2. Nutrient composition of spent coffee grounds in %	
Item	
Crude protein	7.65
Ether extract	0.28
Crude fibre	26.68
Ash	1.99
Caffeine	1.76

ACCEPTED

Table 3. Comparing the proximate analysis, and antioxidant activity of different breeds of pigs with and without coffee supplementation

Coffee						SEM	Breeds	P-value
	-			+				
Species	LYD	YB	YW	YB	YW			YB vs YW
<b>Belly</b>	0	1	3	2	4		0 vs 1 vs 3	1+2 vs 3+4
Protein	14.56	14.11	14.30	14.35	14.70	0.711	0.433	0.821
Fat (%)	32.23 <sup>b</sup>	35.07 <sup>a</sup>	33.02 <sup>b</sup>	35.27	32.90	0.445	<0.001	0.735
Moisture (%)	48.74	45.03	47.61	48.53	47.46	1.085	0.459	0.084
MDA	4.06	3.22	3.98	3.57	3.70	0.229	0.064	0.182
<b>Loin</b>								
Protein	22.12	21.16	21.66	21.38	21.71	0.252	0.147	0.944
Fat (%)	3.46 <sup>b</sup>	3.70 <sup>ab</sup>	3.96 <sup>a</sup>	3.72	3.99	0.114	0.050	0.950
Moisture (%)	70.47	72.40	71.32	71.15	71.94	0.579	0.805	0.118
MDA	0.87	0.64	0.79	0.68	0.74	0.070	0.131	0.285

<sup>1</sup>LYD, Landrace × Yorkshire × Duroc; YB, Yorkshire × Berkshire; YW, Yorkshire × Woori; (-), without SCG; (+), with 0.5% SCG

<sup>2</sup>SEM, standard error of means

<sup>3</sup>MDA, malondialdehyde

Table 4. Comparing the pH and meat color of different breeds of pig with and without coffee supplementation

Coffee							<i>P</i> -value	
	-			+		SEM	Breeds	YB vs YW
Species	LYD	YB	YW	YB	YW			
<b>Belly</b>	0	1	3	2	4			
pH	5.97 <sup>ab</sup>	6.10 <sup>a</sup>	5.76 <sup>b</sup>	6.06	5.67	0.082	<0.001	0.754
Meat color L*	51.56	48.87	50.14	50.75	51.04	0.930	0.285	0.065
Meat color a*	13.74 <sup>b</sup>	18.43 <sup>a</sup>	17.43 <sup>a</sup>	18.28	17.18	0.495	0.044	0.921
Meat color b*	2.70	3.37	3.58	3.19	3.42	0.485	0.681	0.978
<b>Loin</b>								
pH	5.79 <sup>b</sup>	6.13 <sup>a</sup>	5.89 <sup>ab</sup>	6.07	5.89	0.061	0.005	0.634
Meat color L*	51.69	53.12	54.11	53.13	54.03	0.737	0.164	0.944
Meat color a*	16.66	17.37	17.47	17.79	17.84	0.344	0.814	0.950
Meat color b*	3.82 <sup>ab</sup>	3.09 <sup>b</sup>	4.86 <sup>a</sup>	3.47	4.03	0.527	0.049	0.285

<sup>1</sup> LYD, Landrace × Yorkshire × Duroc; YB, Yorkshire × Berkshire; YW, Yorkshire × Woori; (-), without SCG; (+), with 0.5% SCG

<sup>2</sup> SEM, standard error of means

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Table 5. Comparing the meat quality of different breeds of pig with and without coffee supplementation (%)

Coffee						SEM	<i>P</i> -value	
	-			+			Breeds	YB vs YW
Species	LYD	YB	YW	YB	YW			
<b>Belly</b>	0	1	3	2	4		0 vs 1 vs 3	1+2 vs 3+4
WHC	67.48 <sup>b</sup>	72.36 <sup>a</sup>	70.53 <sup>a</sup>	73.20	71.37	0.735	0.008	0.999
Cooking loss	25.35	24.86	25.93	25.67	26.13	1.200	0.426	0.753
Drip loss 6	1.91 <sup>a</sup>	1.14 <sup>b</sup>	1.48 <sup>b</sup>	1.07	1.31	0.103	0.003	0.559
Drip loss 12	2.05	1.74	2.28	1.94	2.08	0.226	0.154	0.389
Drip loss 24	0.96	0.74	0.94	0.90	0.97	0.111	0.273	0.596
Drip loss 48	0.67	0.54	0.45	0.58	0.54	0.085	0.478	0.775
<b>Loin</b>								
WHC	81.26	83.03	81.85	83.13	81.99	0.700	0.074	0.980
Cooking loss	31.13 <sup>a</sup>	27.84 <sup>b</sup>	30.15 <sup>a</sup>	27.99	30.45	0.799	0.005	0.926
Drip loss 6	3.23	1.97	2.04	1.99	2.01	0.244	0.835	0.912
Drip loss 12	1.29	1.20	1.35	1.05	1.04	0.119	0.495	0.397
Drip loss 24	1.31	1.02	1.09	1.14	1.17	0.094	0.614	0.814
Drip loss 48	0.54	0.65	0.57	0.56	0.54	0.110	0.645	0.770

<sup>1</sup> LYD, Landrace × Yorkshire × Duroc; YB, Yorkshire × Berkshire; YW, Yorkshire × Woori; (-), without SCG; (+), with 0.5% SCG

<sup>2</sup> SEM, standard error of means

<sup>3</sup> WHT, water holding capacity

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Table 6. Comparing various fatty acid composition of different breeds of pigs with and without coffee supplementation (%)

Coffee							<i>P</i> -value	
	-			+		SEM	Breeds	YB vs YW
Species	LYD	YB	YW	YB	YW			
<b>Belly</b>	0	1	3	2	4			
Lauric (C12:0)	0.155	0.197	0.167	0.177	0.182	0.014	0.350	0.195
Myristic (C14:0)	0.400	0.528	0.402	0.535	0.487	0.049	0.093	0.438
Palmitic (C16:0)	29.253	31.565	32.353	31.087	31.613	0.528	0.162	0.776
Palmitoleic (C16:1)	2.720	3.180	3.062	3.097	3.052	0.066	0.210	0.567
Stearic (C18:0)	11.363	12.827	13.173	12.962	13.007	0.375	0.572	0.663
Oleic (C18:1)	37.810	40.565	39.580	40.877	39.875	0.748	0.200	0.991
Linoleic (C18:2)	10.305 <sup>b</sup>	13.468 <sup>a</sup>	11.750 <sup>ab</sup>	13.528	12.123	0.456	0.002	0.731
Linolenic (C18:3)	0.728	0.645	0.663	0.650	0.665	0.038	0.623	0.961
Arachidic (C20:0)	0.167	0.172	0.165	0.172	0.168	0.007	0.514	0.827
Lignoceric (C24:0)	1.140	1.105	1.088	1.127	1.118	0.019	0.522	0.830
<b>Loin</b>								
Lauric (C12:0)	0.103	0.145	0.152	0.148	0.158	0.006	0.193	0.791
Myristic (C14:0)	1.507 <sup>b</sup>	2.048 <sup>a</sup>	1.498 <sup>b</sup>	2.002	1.523	0.078	<0.001	0.592
Palmitic (C16:0)	23.148 <sup>b</sup>	23.138 <sup>b</sup>	24.130 <sup>a</sup>	23.187	24.053	0.386	<0.001	0.801
Palmitoleic (C16:1)	3.028 <sup>b</sup>	6.152 <sup>a</sup>	3.037 <sup>b</sup>	6.118	3.140	0.096	<0.001	0.507
Stearic (C18:0)	12.182 <sup>ab</sup>	11.732 <sup>b</sup>	12.712 <sup>a</sup>	11.870	12.485	0.308	0.021	0.572
Oleic (C18:1)	42.055	42.813	41.850	43.107	42.508	0.457	0.089	0.680
Linoleic (C18:2)	10.982	11.752	11.243	11.558	11.277	0.208	0.060	0.573
Linolenic (C18:3)	0.572	0.548	0.527	0.535	0.520	0.029	0.548	0.913
Arachidic (C20:0)	0.210 <sup>b</sup>	0.208 <sup>b</sup>	0.288 <sup>a</sup>	0.213	0.297	0.017	<0.001	0.912
Lignoceric (C24:0)	1.447 <sup>a</sup>	1.268 <sup>b</sup>	1.543 <sup>a</sup>	1.287	1.532	0.075	0.003	0.846

<sup>1</sup>LYD, Landrace × Yorkshire × Duroc; YB, Yorkshire × Berkshire; YW, Yorkshire × Woori; (-), without SCG; (+), with 0.5% SCG

<sup>2</sup>SEM, standard error of means

Table 7. Comparing various saturated, unsaturated, monounsaturated, and polyunsaturated fatty acids characteristics of different breeds of pigs with and without coffee supplementation (%)

Coffee						SEM	<i>P</i> -value	
	-			+			Breeds	YB vs YW
Species	LYD	YB	YW	YB	YW		0 vs 1 vs 3	1+2 vs 3+4
<b>Belly</b>	0	1	3	2	4			
SFA	42.475	46.388	47.347	46.052	46.570	0.722	0.273	0.741
UFA	51.558 <sup>b</sup>	57.858 <sup>a</sup>	55.053 <sup>a</sup>	58.152	55.715	0.845	0.005	0.827
MUFA	40.528	43.747	42.640	43.973	42.927	0.737	0.155	0.968
PUFA	11.032 <sup>b</sup>	14.112 <sup>a</sup>	12.412 <sup>ab</sup>	14.178	12.788	0.487	0.002	0.726
MUFA / PUFA	3.742 <sup>a</sup>	3.107 <sup>c</sup>	3.470 <sup>b</sup>	3.123	3.367	0.159	0.028	0.644
<b>Loin</b>								
SFA	38.597 <sup>b</sup>	38.535 <sup>b</sup>	40.322 <sup>a</sup>	38.697	40.038	0.512	<0.001	0.496
UFA	56.635 <sup>b</sup>	61.260 <sup>a</sup>	56.653 <sup>b</sup>	61.313	57.438	0.504	<0.001	0.450
MUFA	45.082 <sup>b</sup>	48.963 <sup>a</sup>	44.885 <sup>b</sup>	49.222	45.643	0.479	<0.001	0.605
PUFA	11.553	12.298	11.770	12.093	11.797	0.211	0.052	0.568
MUFA / PUFA	3.910 <sup>ab</sup>	3.990 <sup>a</sup>	3.820 <sup>b</sup>	4.078	3.873	0.085	0.037	0.837

<sup>1</sup>LYD, Landrace × Yorkshire × Duroc; YB, Yorkshire × Berkshire; YW, Yorkshire × Woori; (-), without coffee; (+), with 0.5% SCG

<sup>2</sup>SEM, standard error of means

<sup>3</sup>SFA, saturated fatty acid; UFA, unsaturated fatty acid; MUFA, monosaturated fatty acids; PUFA, polyunsaturated fatty acids