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5 Abstract

6 This study was to investigate effects of different phytogenic feed additives (PFA) in grower finishing 7 pigs with stressed by high stocking density. A total of 84 growing pigs [(Landrace × Yorkshire) × Duroc] 8 with initial body weight (BW) of 28.23 ± 0.21 kg were used for 10 weeks (4 replicate pens with 3 pigs 9 per pen). The dietary treatment consisted of basal diets in animal welfare density (PC, Positive control), 10 basal diet in high stocking density (NC, negative control), NC + 0.04% bitter citrus extract (PT1), NC 11 + 0.01% microencapsulated blend of thymol&carvacrol (PT2), NC + 0.10% mixture of 40% bitter citrus extract and 10% microencapsulated blend of thymol and carvacrol (PT3), NC + 0.04% premixture of 12 grape seed and grape marc extract, green tea and hops (PT4), and NC + 0.10% fenugreek seed powder 13 (PT5). The reduction of space allowance significantly decreased (P < 0.05) growth performance 14 (average daily gain, average daily feed intake, feed efficiency) and nutrient digestibility (dry matter, 15 crude protein). Also, the fecal score of NC group increased (P < 0.05) compared with other groups. In 16 blood profiles, lymphocyte decreased (P < 0.05), and neutrophil, cortisol, TNF- α increased (P < 0.05) 17 18 when pigs were in high stocking density. Basic behaviors (feed intake, standing, lying) were inactive (P < 0.05) and singularity behavior (biting) were increased (P < 0.05) under high stocking density. 19 However, PFA groups alleviated the negative effects such as reducing growth performance, nutrient 20 21 digestibility, increasing stress indicators in blood and animal behavior. In conclusion, PFA 22 groupsimproved the health of pigs with stressed by high stocking density and PT3 is the most effective.

- 23
- 24 Keywords: Pig, Robustness, Additive, Stress, Plant Extract, High Stocking Density

Introduction

26 Recently, there has been increased interest in using natural and safe feed additives to enhance robustness 27 for pigs [1,2]. Phytogenic feed additives (PFA) are plant-derived compunds such as leaves, bark, seeds, roots, flowers, twigs, tree herbs, and fruits [3]. According to the European Council, PFAs can be 28 categorized as sensory and flavoring compounds and generally feels safe as substitutes for antibiotics 29 30 [3]. PFAs have been recognized as the latest feed additives and antibiotics alternatives for livestock 31 [4,5]. Previous studies have reported that PFA complex including sunflower, thyme, and garlic can 32 improve growth performance in monogastric animals [6-8]. Rahal et al [9] have also reported that dietary PFA supplementation has immunomodulatory effects such as immunoglobulin secretion, 33 cytokine, lymphocyte expression, phagocytosis, and histamine release. Essential oils such as thymol, 34 cavacrol, cymene, terpinene reduce the pathogenic microbial load, but also promote digestive enzymes 35 thereby affecting nutrient digestibility [10-12]. Other studies have shown dietary herbs (i.e., onion, 36 fenugreek seed, and anise seed) enhanced economical efficiency to farms by improving the growth and 37 38 health of mono-gastric animals [7,13,14]. High stocking density is the most significant caused by inducing stress during growing-finishing periods. Stress caused high stocking density can reduce feed 39 intake, thereby causing low body weight gain [15-17]. Also, this stress can increase aggressive and 40 negative social behavior such as fighting, feeder occupying, tail biting [18,19] and the incidence of 41 42 body lesions [20-22]. Supplementaion of Scutellaria baicalensis L. roots mitigated negative behavior 43 caused by heat stress in mono-gastric animals [23,24]. However, studies on the relationship between 44 high stocking density and PFA have not been reported. In addition, there are few studies searching for effective PFA against stress derived from high stocking density. Therefore, the objective of this study 45 46 was to explore effective PFA against environmental stress and the exact mechanism alleviated by PFA 47 in a stress situation for grower-finishing pigs.

Marterial and Methods

The experimental protocol for this study was reviewed and approved by the Institutional Animal Care
and Use Committee of Chungbuk National University, Cheongju, Korea (approval CBNUA-1530-2101).

53

54 Preparation of phytogenic feed additives

PFA1 is a bitter citrus extract (BioFlavex[®] GC, HTBA, Beniel, Spain) that is rich in 25-27% naringin 55 and 11-15% neohesperidin. PFA2 is a microencapsulated blend of thymol and carvacrol (AviPower[®] 2, 56 VetAgro SpA, Reggio, Emmilia, Italy) that contains 7% of thymol and 7% carcacrol. PFA3 is a mixture 57 of PFA1, PFA2 and excipient in ratio of 4:1:5. It contains 0.7% thymol, 0.7% carvacrol, $10 \sim 10.8\%$ 58 naringin and $4.4 \sim 6\%$ neohesperidin. PFA4 is a premixture of grape seed & grape marc extract, green 59 tea and hops (AntaOx[®]FlavoSyn, DR. Eckel GmbH, Niederzissen, Germany) containing more than 10% 60 of flavonoids. PFA5 is fenugreek seed powder containing 12% saponin (Fenugreek Seed Powder, P&D 61 62 Export, Jaguar, India). All PFAs materials were provided by EUGENE BIO Co., (Suwon, South Korea).

63

64 Animals, housing, and experimental design

65 A total of 84 crossbred LYD ([Landrace × Yorkshire] × Duroc) mixed-sex growing pigs at 10 weeks 66 of age (average body weight 28.23 ± 2.89 kg) were used in a 10-week feeding trial. Pigs were allotted 67 to one of seven treatments in a completely randomized block design based on initial body weight (BW). 68 Treatments were as follow:PC (positive control; basal diet in animal welfare density), NC (negative control; basal diet in high stocking density), PT1 (basal diet with 0.05% PFA1 in high stocking density), 69 70 PT2 (basal diet with 0.04% PFA2 in high stocking density), PT3 (basal diet with 0.10% PFA3 in high 71 stocking density), PT4 (basal diet with 0.04% PFA4 in high stocking density), PT5 (basal diet with 0.05% 72 PFA5 in high stocking density). All pigs were housed in an environmentally controlled room. There are two types of room area. In growing pig periods, animal welfare stocking density is $0.55m^2/pig$, high 73 stocking density is 0.40 m^2/pig and in finshing pig periods, animal welfare stocking density is 1.00 74 m^2/pig , high stocking density is 0.60 m^2/pig . Each pen consisted of a stainless steels self-feeder and 75

nipple drinker at one-side. There are 4 replicate pens with 3 pigs per pen during the experiment period.
Basal diet was mostly consisted with corn and soybean meal and were formulated to meet or exceed
National Research Council (2012) recommendations (Table 1). During the experimental period, each
pen was equipped with a self-feeder and nipple drinker to allow *ad libitum* access to feed and water.

80

81 Sampling and measuremets

82 *Growth performance*

To calculate average daily gain (ADG), pig's BW was individually measured at the 09:00 on an empty
stomach at start of grower (0 weeks), end of grower and start of finisher (4 weeks), end of the finisher
(10 weeks). Feed intake and wasted feed were recorded daily to calculate average daily intake (ADFI).
Feed efficiency (G:F) was calculated by ratio of body weight gain and feed intake.

87

88 Nutrient digestibility

Apparent total tract digestibility (ATTD) of dry matter (DM) and nitrogen (N) were estimated using 89 0.2% of chromic oxide as an inert indicator (Fenton & Fenton, 1979). Crude proteins (CP) were 90 measured from the nitrogen. Pigs were fed diets mixed with chromic oxide on 4th week and 10th week. 91 Fresh fecal grab samples collected via rectal massage from each pig, and these samples were stored in 92 a freezer at -20°C until analyzed. All feed and fecal samples were analyzed for DM and N following 93 94 the procedures outlined by the AOAC (2005) methods. N was determined with a Kjeltec 2300 nitrogen 95 analyzer (Foss Tecator AB, Hoeganaes Sweden) and Chromium was analyzed via UV absorption 96 spectrophotometry (Shimadzu UV-1201, Shimadzu, Kyoto, Japan) following the method described by 97 Williams, David, & Iismaa (1962). The ATTD of DM and N were calculated with indirect ratio methods using the following formula: Coefficient of apparent total tract digestibility= $\{1-[(Nf \times Cd)/(Nd \times Cf)]\}$ 98 99 \times 100. Where: Nf = nutrient concentration in faces (% DM), Nd= nutrient concentration in diet (% 100 DM), Cf = chromium concentration in faeces (% DM), Cd = chromium concentration in diets (% DM).

101

102 Fecal score

During experiment, each pig fecal score was measured by same person before daily feeding. The fecal was scored according to its moisture content and shape. Normal feces are 0-point, soft feces are 1-point, mild diarrhea are 2-point and severe diarrhea are 3-point (Marquardt et al., 1999). The score was calculated by averaging each group with the average value of the daily fecal score of each pig.

107

108 *Blood sample*

For the serum profile, at each pen, one pig was randomly selected to collect blood samples through 109 venipuncture at the end of 4th week, and 10th week. At the time of collection, blood samples were 110 collected both whole blood and serum in nonheparinized tubes and vacuum tubes containing K₃EDTA 111 (Becton Dickinson Vacutainer systems, Franklin Lake, NJ, U.S.A.), respectively. White blood cells 112 (WBC) and WBC including lymphocyte, neutrophil, basophil concentration in whole blood were 113 114 measured using an automatic blood analyzer (ADVIA 120, Bayer, NY, USA). After collection, serum samples were centrifuged 3,000g for 15 min at 4 °C. Samples were stored at -20°C in the refrigerator 115 116 until analysis. Serum cortisol levels were assessed using enzyme-linked immunosorbent assay kits (LDN GmbH & Co., Nordhorn, Germany) following to the manufacturer's protocol. Tumor necrotizing 117 factor-alpha (TNF-α) and interukine-6 (IL-6) concentration was analyzed with ELISA kit (Quantikine, 118 R&D systems, Minneapolis, MN, USA) and they were measured at 450 nm. 119

120

121 *Pig behavior*

Collection of each pig image data was recorded by using six-day/night infrared cameras (QNB-7080 122 123 RH, Hanwha, Seoul, Korea) installed 3m above each pen. A total of 28 pig behaviors were analyzed by 124 randomly selecting one pig from each pen. Observers collected data based on results of Yang et al. 125 (2018), and only one person made all observations and video analysis to see consistent results. The pig 126 behavior analysis was classified for the following criteria (A) Feed intake: the act of eating with the 127 head in the feed bin, or similar behavior. (B) Standing: the act of standing still with the forelimbs and hindlimbs extended perpendicular to the floor, or similar behavior. (C) Lying: the act of lying with the 128 whole body on the floor, lying with the head, front legs, hind legs and abdomen all touching the floor. 129

130 (D) Sitting: Two front legs are spread vertically to the floor, two rear legs and two hips are sitting on 131 the floor, like a dog sitting on the floor, or something like that. (E) Drinking water: the act of drinking water for 10 seconds by putting your mouth in a drinking nipple. (F) Posture transition (lying→standing) 132 A behavior that changes from lying down to standing, in which the two front legs are stretched first, 133 and the hind legs are naturally stretched out. (G): Posture transition (standing \rightarrow lying): A behavior that 134 135 changes from a standing behavior to a lying behavior, in which the two front legs are bent to the floor first, and then the two hind legs are naturally folded and lying down. (H) Rooting: the act of repeating 136 similar behaviors, such as scratches, itching, or something on the nose and front legs. (I) Biting: The 137 act of biting another pig's ears, mouth, and tail with teeth and then biting again or doing similar things. 138

139

140 Statistical analysis

141 All data were analyzed by one-way ANOVA using SPSS software (ver. 20.0; IBM, USA), and the

142 differences among treatments were examined by Tukey's multiple range test, which were considered to

143 be significant at P < 0.05, unless otherwise stated.

Results

145 *Growth performance*

There was no difference between treatment groups in the initial BW of pigs (Table 2). During the 146 growing period (0-4 weeks), PT3 group significantly increased (P < 0.05) ADG and G:F ratio than NC 147 148 group. During the finishing period (4-10 weeks), NC group significantly decreased (P < 0.05) ADG and ADFI than PC group. PFA groups ADG significantly higher (P < 0.05) than NC group. The PT3-149 PT4 group ADFI significantly higher (P < 0.05) than NC group. During entire experimental period (0-150 10 weeks), NC group significantly decreased (P < 0.05) ADG, ADFI and G:F ratio than PC group. PFA 151 152 groups significantly higher (P < 0.05) ADFI than NC group. The PT3-PT4 groups significantly 153 increased (P < 0.05) ADG and G:F ratio than NC group.

154

155 *Nutrient digestibility*

During the growing period (0-4 weeks), the ATTD of DMsignificantly increased (P < 0.05) in PT1-PT3 groups compared PC group (Table 3). The ATTD of CP significantly decreased (P < 0.05) in NC group compared toPC group. However, PFA groups significantly increased (P < 0.05) CP digestibility than NC group. During the finishing period (4-10 weeks), PFA groups decreased (P < 0.05) ATTD of DM and CP compared to NC group. The PT3-PT4 groups CP digestibility numerically increased (P < 0.05) than other PFA groups.

162

163 *Fecal score*

During the growing period (0-4 weeks), NC group showed significantly higher (P < 0.05) fecal score than PC group (Table 4). However, PFA groups significantly decreased (P < 0.05) fecal score compared to NC group. During finishing period (4-10 weeks), the difference of diarrhea incidence was not observed among all treatment groups.

168

169 Blood profile

During the growing period (0-4 weeks), there were no significant difference (P > 0.05) on WBC,

171 Basophil, and IL-6 among treatment groups (Table 5). The NC group significantly decreased (P < 0.05) 172 lymphocyte and increased (P < 0.05) neutrophil, cortisol, and TNF- α level in blood compared with PC group. However, PFA groups significantly alleviated (P < 0.05) these negative effects by stress with 173 stocking density and was similar with the level of PC group. During the finishing period (4-10 weeks), 174 there were no significant difference (P > 0.05) on WBC among treatment groups. NC group significantly 175 176 sdecreased (P < 0.05) lymphocyte and significantly increased (P < 0.05) neutrophil, cortisol, and TNF- α level in blood compared to PC group. However, PFA groups significantly increased (P < 0.05) 177 lymphocyte and significantly decreased (P < 0.05) neutrophil, cortisol, and TNF- α compared with NC 178 179 group. PT3 group showed (P < 0.05) the lowest results in neutrophil, cortisol, and IL-6 among PFA 180 groups.

181

182 Animal behavior

The effects of different PFA on animal behavior were shown in Table 6, Table 7, Figure 1. During the growing period (0-4 weeks), there are no significant difference (P > 0.05) in basic behavior and most of singularity behavior. The NC group had significantly higher (P < 0.05) biting frequency than PC group. However, PFA groups had significantly alleviated (P < 0.05) biting frequency compared with NC group. Among PFA groups, PT3 group showed the lowest biting frequency.

During the finishing period (4-10 weeks), NC group showed (P < 0.05) more lying time and less feed intake and standing time than PC group. Feed intake time significantly increased (P < 0.05) in PFA groups than NC group. Standing time significantly increased (P < 0.05) in PT2-PT5 group than NC group. Lying time significantly decreased (P < 0.05) in PFA groups than NC group. Especially, PT3-PT5 groups showed similar result with PC group. In singularity behavior, there are no significant difference (P > 0.05) in treatment groups. But NC group showed numerically high number of biting than other treatment groups.

Discussion

196 *Growth performance*

197 High stocking density can disturb the movement of animals due to limited feeding environment (space, feeders, and drinkers). Moreover, high stocking density can interfere with airflow and generate heat 198 199 energy [12]. It can result in difficulty in evacuating body temperature, poor air quality, reduced access 200 to feed and water, and poor performance of animals due to increased ammonia levels [28-30]. High heat 201 energy and poor air quality are known to cause heat stress and adverse effects on growth rate, feed 202 consumption, mortality, and health [31-33]. Similarly, our study showed that pigs under high stocking density (i.e., 0.40 m²/ growing pig, 0.60 m²/finishing pig) had reduced ADG and ADFI by 16.38% and 203 11.24%, respectively, than those under welfare density (i.e., $0.55 \text{ m}^2/\text{growing pig}$, $1.0 \text{ m}^2/\text{finishing pig}$) 204 during the whole period (grower: 28-56 kg, finisher: 56-103 kg). Spicer and Aherne [34] have also 205 reported that daily gain and daily feed are reduced 8.47% and 13.15%, respectively, when group size is 206 decreased from 0.72 m²/pig to 0.35 m²/pig. Stress-induced heat and high stocking densities can reduce 207 208 growth performance by damaging cellular structure, increasing intracellular water imbalance, and increasing free radical concentration [35]. However, our study revealed that pigs under high stocking 209 density with supplementation of PFA showed improvement (i.e., BW decreased 11.61%) in growth 210 performance compared to those in the unsupplemented group. Many researchers reported that dietary 211 supplementation of PFA such as Korean pine extract, cinnamon, turmeric, essential oils, and rosemary 212 213 can improve growth performance with reducing stress response [35-38]. In our study, PFA 214 supplementation under our high stocking density showed no difference in ADFI between treatments in the growing period, but significantly increased with PFA supplementation in the finishing period This 215 216 is consistent with previous studies suggesting that PFA is effective for intake when supplied long-term 217 [39]. Moreover, PFA3 could improve the flavor of feed and increase the palatability of feed intake in pigs [40-43]. Therefore, using natural products with polyphenols (suitable structure for free radical 218 219 scavenging activity) can effectively alleviate stress caused by low space allowance and heat through 220 their antioxidant activity with improved low feed intake, thereby increasing growth performance.

195

222 Nutrient digestibility

223 High stocking density can negatively affect nutrient digestibility and growth performance. During the whole experiment periods, nutrient digestibility (DM, CP) showed improvement in the treatment group 224 225 added with PFA than that in the control group without PFA under high stocking density. PFA can also 226 enhance nutrient digestibility and absorption [44,45]. It has been reported that the addition of essential 227 oils to monogastric animals can enhance the activity of trypsin, maltase and pancreatic amylase and 228 increases glucose absorption in the small intestine [46]. Therefore, the addition of PFA can stimulate 229 the secretion of mucus in the intestine, thereby reducing the adhesion of pathogens and stabilizing 230 intestinal microbial symbiosis [47]. It can be seen that improved digestive tract function is associated 231 with increased nutrient digestibility. It can also be said that the antibacterial action of PFA contributes 232 to the increase of nutrient digestibility. PFAs such as carvacrol, thymol, anetol, oregano, anise, and 233 citrus essential oil have antibacterial activity against intestinal microbes when ingested. Among them, phenolic substances are the most active compounds [48,49]. PT2 and PT3 have a phenolic structure in 234 235 our experiment. It was shown that the digestibility of DM and CP was higher than the high stocking density throughout the experiment period. Fiesel et al [50] reported an increase in nutrient digestibility 236 due to the antioxidant effect of polyphenols and an increased absorbable surface of the intestine. As the 237 experiment progressed, the digestibility deviation of DM and CP increased according to the presence 238 239 or absence of PFA in the feed under high stocking density. In this experiment, it was confirmed that the 240 digestibility was gradually improved when PFA was used, leading to improved performance of pigs. In 241 particular, it was found that the digestibility was significantly improved by flavonoids, a common 242 component of PT3-PT5 additives. A previous study has shown that flavonoids have DM and CP 243 synergistic effects [51]. Therefore, it can be concluded that the use of flavonoid additives can increase 244 the digestibility of nutrients, as it can improve nutrient availability by boosting immunity and 245 antibacterial action in pigs.

246

247 Fecal score

248 In high stocking density, the frequency of diarrhea was increased during the growing period, although

249 it showed no significant difference during the finishing period. Many studies have found that diarrhea 250 in pigs is more likely to be induced by stress [52-54]. Actually, the frequency of diarrhea is increased 251 in weaned pigs during stress [55]. Diarrhea has been found intermittently in growing pigs [56]. When pigs get stressed, their immunity is lower and pathogens in the intestine are activated. Intestinal 252 pathogens can suppress unnecessary energy loss such as reduced feed intake and G:F ratio known to 253 254 interfere with immune system activation. In addition, intestinal pathogens can inhibit homeostasis of 255 the epithelial barrier, causing secretory diarrhea due to intestinal damage through osmotic stress or 256 inflammatory diarrhea by increasing inflammatory cytokines. However, in our study, the frequency of 257 diarrhea was significantly reduced when PFAs were fed to pigs in a stressful situation. These results 258 indicate that PFA can improve fecal status by improving intestinal health, and further studies on fecal 259 microflora should be conducted. When pigs are fed with natural products reduces the frequency of 260 diarrhea due to stress as the natural product's antibacterial action improves intestinal health and increased digestibility [57,58]. Many researchers have checked diarrhea scores of weaning pigs, but not 261 262 those of growing to finishing pigs. In the present study, complete diarrhea was not found even in the growing period, although a lot of soft feces were observed for pigs under a high stocking density 263 condition. The difference between growing period and finishing period is that as pigs grow, their 264 immune system gets better, and their gut health improves. Therefore, we can confirm a meaningful 265 266 diarrhea score even in pigs during the growing period. Thus, it is necessary to check the status of feces.

267

268 Blood profile

In the present study, there were no significant differences in blood profile between the entire experiment period WBC or growing period basophil and IL-6 of pigs between treatment groups. However, pigs fed with PFAs under a high stocking density condition had better blood results than those without addition of additives under a high stocking density condition. Pigs with high stocking density are subjected to critical psychological, social, and environmental stresses. High stocking density can also cause chronic severe stress that affects immunity and health [60]. Lymphocytes show various immunological responses including modulation of immune defense and immunoglobulin [61]. In our study, 276 lymphocytes were decreased during stress situation, but returned to PC levels when PFAs were added. 277 According to Dhabhar [62], in stressful situations, lymphocyte counts are decreased due to changes 278 induced by trafficking or redistribution of lymphocytes to other body compartments of glucocorticoids. 279 This result was similar to our study. In our study, the number of neutrophils was increased when pigs 280 were stressed. This number was then decreased after supplementation with PFA in our study. It has been 281 reported that stressful situations cause decreasing lymphocytes and increasing neutrophils in the blood 282 [63]. As a result, it was possible to confirm the indirect change caused by supplementation of PFA to 283 relieve stress. Cortisol, a steroid hormone, or glucocorticoid produced by the adrenal gland and released 284 in response to stress, is often used as a physiological marker to quantify animal stress [64]. It is well 285 known that cortisol can regulate intermediary metabolism, immunity, and growth [65,66]. A poor 286 welfare situation can cause animals to be extremely stressed. In this study, cortisol level was increased 287 under high stocking density compared with animal welfare density (space decreased in growing pig 27.27%, in finshing pig 40%). This result was agreement with the results of Jang et al. [67] that reported 288 289 decreasing space allowance (decreasing 28.13%) induced increasing cortisol level (2.3µg/dL to 4µg/dL). 290 However, PFAs supplementation alleviated high cortisol level in blood caused by high stocking density. Li et al [67] observed that flavonoids, which are physiologically active substances of PFA, down-291 regulated immune responses by mediated viruses and the T-cell, thereby reducing psychological stress. 292 293 This observation suggests that PFA mitigates the increased cortisol concentration by high stocking 294 density.

295 Pro-inflammatory cytokines such as TNF- α and IL-6 are potential outputs of the cellular immune system 296 and can indirectly reflect immune responses due to the activation of T-cells [68,69]. This study showed 297 that high stocking density increased pro-inflammatory cytokine level. These results suggest that the 298 environmental stress caused by a limited space allowance can induce a cellular immune response. When 299 stressed out, pro-inflammatory cytokines are secreted to promote cortisol secretion and suppress growth 300 hormone secretion [70,71]. Excessive pro-inflammatory cytokines can induce fever, inflammation, 301 tissue destruction [72], and in some cases, even shock and death (Dinarello, 2000). Thus, the immune 302 system is activated due to high stocking stress, which shifts nutrient distribution priorities from growth 303 to host defense [68,70,74].

In addition, TNF- α and IL-6 content is reduced through improved gut microbiota, antioxidant, and antiinflammatory effects, due to improved digestibility of nutrients, alleviating stress response, and strengthening immunity [75,76]. However, PFA was effective methods to alleviate negative effects of a high stocking density in our study. Other researchers also reported essential oil and herb extract reduced pro-inflammatory cytokines [77,78].

Therefore, PFA is effective in relieving stress, and PT3 group showed the highest effect among PFAs group. The reason the PT3 group outperformed the others was due to the construction of the PFA group. Flavonoids and terpenoids (carbacrol and thymol) may protect cells from the harmful effects of autoxidation.

313

314 Animal behavior

A high stocking density equates to a reduced floor space allowance. Decreasing floor space allowance 315 per pig increases the frequency of contact, social tension, and aggression [79-82]. In addition, when 316 heat production per unit floor area is increased, heat stress will occur and induce oxidative stress [82,22]. 317 If this stress is not well managed in pigs, it can increase their susceptibility to stress and hence reduce 318 319 their immune and health status. Throughout our study, animal behavior at high stocking density 320 improved when fed with PFA. The biting frequency was increased in NC but decreased after PFA 321 treatment similar to PC. Among all treatment groups, PT3 group showed the lowest biting frequency. 322 Greene et al [35] has reported that biting as a representative form of aggressive behavior can occur in pigs under chronic stress. This is consistent with our study. When ingesting phenolic compounds as 323 324 components of PT3 group, it is possible to restore redox homeostasis and prevent oxidative stress by 325 improving the activity of antioxidant enzymes SOD, CAT, GPx, and GR [84]. Therefore, the effect of 326 adding PFA3 not only can help pigs cope with biting behavior caused by stress, but also can overcome it. During the finishing period, basic behaviors (eating, standing, lying down) were more active when 327 fed with PFA added in high stocking density. In addition, the feed intake increased during PFA feeding 328 329 in growth performance. Feed intake is an important indicator because it is related to body weight, ADG,

330 ADFI, and G: F ratio. Pigs with a high stocking density face difficulty in feeding due to competition in 331 the feeder. In this study, PC group showed less time than other treatments in feed intake. Therefore, the number of trips to the feeder is directly related to intake and can affect growth performance. Also, 332 333 standing and lying time were similar to NC group. They were more active than PC group. Especially, PT3-PT5 groups are more activated than others. Pearce and Paterson [85] have reported that observation 334 335 of the behavior of standing motionless in a narrow space is a behavior that pigs do to cope with stress at a high stocking density. As stress increased, the amount of physical activity decrease. It can be seen 336 337 that when the standing time decrease, the lying time increases at the same time. This indicates that there is a close relationship between basic behavior and growth performance. Through this experiment, it can 338 339 be seen that when pigs get stressed, their basic behaviors (standing, lying, and feeding) were affected at the same time. 340

Conclusion

343	Dietary supplementation of PFA improves the growth performance, nutrient digestibility, immunity,
344	fecal score, and animal behavior in grower-finishing pigs. As a result, lymphocytes, neutrophils, cortisol,
345	IL-6, and TNF- α in the blood, bites, and basic behaviors were improved, indicating that stress was
346	reduced and strengthened. The diarrhea index improved because of getting healthier, which means less
347	damage to the intestines and increased digestibility. Due to these positive effects, growth performance
348	was improved, and it was found that PFA is an effective additive for stress due to high stocking density.
349	Among them, the most effective and additional advantages were found when using PFA3 (mixture of
350	PFA1 40%, PFA2 10% and excipient 50%) rather than using PFA1 (bitter citrus extract) and PFA2
351	(microencapsulated blend of thymol and carvacrol) separately.

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Items	Grower 0-4w	Finisher 4-10w
Ingredients (%)		
Corn	65.10	72.38
Soybean meal	23.90	17.40
Wheat bran	7.00	6.00
Soybean oil	1.00	1.00
L-Lysine	0.10	0.28
DL-Methionine	0.04	0.04
L-T-hreonine	0.03	0.03
Dicalcium phosphate	1.00	1.00
Limestone	1.20	1.25
Salt	0.50	0.50
Vitamin premix ^a	0.08	0.08
Mineral premix ^b	0.05	0.05
Calculated composition		
ME (kcal/kg)	3276	3284
Crude protein (%)	18.00	15.50
Lysine (%)	1.01	0.97
Methionine (%)	0.33	0.29
Calcium (%)	0.78	0.76
Phosphorus (%)	0.62	0.58

Table 1. Ingredients and chemical composition of the basal experimental diets (as fed basis).

Note: ME, metabolizable energy.

^aProvided per kilogram of complete diet: 20 000 IU of vitamin A, 4000 IU of vitaminD₃, 80 IU of vitamin E, 16mg of vitamin K₃, 4 mg of thiamine, 20mg of riboflavin, 6 mg of pyridoxine, 0.08 mg of vitamin B₁₂, 120 mg of niacin, 50 mg of Ca-Pantothenate, 2 mg of folic acid, 0.08 mg of biotin.

^bProvided per kilogram of complete diet: 12.5 mg of manganese, 179 mg of zinc, 140 mg of copper, 0.5 mg of iodine, 0.4 mg of selenium.

Items	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value
BW, kg									
initial	28.00	27.53	27.64	28.97	28.62	28.48	27.88	0.309	0.868
4w	56.30 ^{ab}	53.23 ^b	53.77 ^b	57.43 ^{ab}	59.35ª	57.04 ^{ab}	56.15 ^{ab}	0.592	0.083
final	110.63ª	96.72°	97.23°	101.37 ^{bc}	109.43ª	106.41 ^{ab}	102.39 ^{bc}	0.767	< 0.001
0-4w									
ADG, kg	0.98 ^{ab}	0.89 ^b	0.90 ^b	0.98 ^{ab}	1.06 ^a	0.98 ^{ab}	0.98 ^{ab}	0.014	< 0.001
ADFI,kg	1.98 ^{bc}	2.03 ^{abc}	1.94°	2.01 ^{abc}	2.06 ^{ab}	2.08 ^a	2.03 ^{ab}	0.012	< 0.001
G:F	0.49 ^{ab}	0.44 ^b	0.46 ^{ab}	0.49 ^{ab}	0.51ª	0.47^{ab}	0.48 ^{ab}	0.006	< 0.001
4-10w							$, \bigvee$		
ADG, kg	1.26ª	1.01°	1.01°	1.02°	1.16 ^{ab}	1.15 ^b	1.08 ^{bc}	0.013	< 0.001
ADFI, kg	2.92ª	2.54°	2.82 ^{ab}	2.82 ^{ab}	2.88 ^{ab}	2.79 ^b	2.77 ^b	0.016	< 0.001
G:F	0.43ª	$0.40^{\rm abc}$	0.36 ^c	0.36°	0.41 ^{ab}	0.41 ^{ab}	0.39 ^{bc}	0.004	< 0.001
Overall period				\leq					
ADG, kg	1.16ª	0.97°	0.98 ^e	1.02 ^{de}	1.14 ^{ab}	1.1 ^{bc}	1.05 ^{cd}	0.009	< 0.001
ADFI, kg	2.58ª	2.29°	2.41 ^b	2.43 ^b	2.46 ^b	2.43 ^b	2.41 ^b	0.012	<0.001
G:F	0.45 ^{ab}	0.43 ^{cd}	0.41 ^d	0.42 ^{cd}	0.46 ^a	0.45 ^{ab}	0.44 ^{bc}	0.004	< 0.001

Table 2. Effects of different phytogenic feed additives on growth performance in growing-finishing pigs with stressed by stocking density

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; BW, body weight; ADG, average daily gain; ADFI, average daily feed intake; G:F, feed efficiency; SEM, standard error of means. Each value is the mean value of 4 replicates. ^{a-e}Means within column with different superscripts differ significantly (p < 0.05).

Items	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value
4 week									
DM, %	85.55ª	84.02 ^b	85.72ª	85.71ª	86.14ª	85.43 ^{ab}	85.46 ^{ab}	0.143	0.03
СР, %	73.35ª	69.45 ^b	73.35ª	73.58ª	74.47ª	73.16 ^a	72.88ª	0.271	0.01
10 week									
DM, %	85.85ª	83.17 ^b	85.84ª	85.75ª	86.93ª	86.15ª	86.11ª	0.179	0.01
СР, %	67.98 ^{bc}	64.75°	70.58 ^{ab}	70.65 ^{ab}	72.87 ^a	71.37ª	71.22 ^{ab}	0.398	0.01

Table 3. Effects of different phytogenic feed additives on nutrient digestibility in growing-finishing pigs with stressed by stocking density

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; DM, dry matter; CP, crude protein; SEM, standard error of means. Each value is the mean value of 4 replicates. ^{a-c}Means within column with different superscripts differ significantly (p < 0.05).

Table 4. Effects of different phytogenic feed additives on fecal score in growing pigs with stressed by stocking
density

Items	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value
4 week									
Fecal score ¹	0.26 ^b	0.76 ^a	0.33 ^b	0.31 ^b	0.30 ^b	0.29 ^b	0.28 ^b	0.017	0.02

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; SEM, standard error of means. Each value is the mean value of 4 replicates. ¹Fecal score was determined as follow : 0, normal feces; 1, soft feces; 2, mild diarrhea; 3, severe diarrhea. ^{a-b}Means within column with different superscripts differ significantly (p < 0.05).



Items	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value
4 week									
WBC, 10 ³ /µl	23.03	23.47	23.18	23.21	22.72	22.99	23.04	0.155	0.93
Lymphocyte, %	45.31ª	36.9 ^b	46.77 ^b	47.47 ^{ab}	46.37 ^b	41.70 ^a	47.80ª	0.506	0.01
Neutrophil, %	39.43 ^{bc}	48.07ª	39.63 ^{bc}	38.53 ^{bc}	36.97°	39.83 ^b	40.03 ^b	0.436	0.03
Basophil, %	0.65	0.70	0.67	0.70	0.63	0.70	0.63	0.027	0.98
Cortisol, ug/dL	1.82°	3.47 ^a	2.60 ^b	2.19 ^{bc}	1.92°	2.74 ^b	2.78 ^b	0.069	0.01
TNF-α, pg/mL	61.90 ^b	73.13ª	62.63 ^b	62.77 ^b	61.93 ^b	62.67 ^b	62.40 ^b	0.506	0.01
IL-6, pg/mL	72.58	72.20	72.50	72.00	72.37	72.13	72.33	0.254	0.99
10 week									
WBC, 10 ³ /µl	17.74	17.76	17.85	17.64	17.76	17.63	17.72	0.148	0.99
Lymphocyte, %	43.40 ^a	35.90 ^b	45.40 ^a	46.57 ^a	44.37ª	45.63ª	46.87ª	0.492	0.01
Neutrophil, %	42.08 ^b	52.37ª	41.20 ^b	40.83 ^b	42.03 ^b	42.20 ^b	44.13 ^b	0.499	0.01
Basophil, %	0.68 ^{ab}	0.70^{a}	0.70 ^a	0.63 ^{abc}	0.50 ^{bc}	0.47°	0.47°	0.019	0.01
Cortisol, ug/dL	0.72 ^b	2.40 ^a	0.61 ^{bc}	0.64 ^{bc}	0.51°	0.67 ^b	0.61 ^{bc}	0.070	0.01
TNF-α, pg/mL	86.70 ^b	99.83ª	85.73 ^b	85.80 ^b	84.90 ^b	85.70 ^b	85.83 ^b	0.596	0.01
IL-6, pg/mL	80.80 ^{ab}	81.77ª	80.37 ^{ab}	80.70 ^{ab}	80.30 ^b	80.43 ^{ab}	80.53 ^{ab}	0.132	0.04

Table 5. Effects of different phytogenic feed additives on blood profile in growing-finishing pigs with stressed by stocking density

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; WBC, white blood cell; TNF- α , tumor necrosis factor- α ; IL-6, InterLeukin-6; SEM, standard error of means. Each value is the mean value of 4 replicates. ^{a-c}Means within column with different superscripts differ significantly (p < 0.05).

	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value
Basic behavior (mi	n/hour)								
Feed intake	4.03	4.01	4.03	4.02	4.10	4.10	4.05	0.009	0.02
Standing	7.05	7.11	6.99	7.12	7.01	6.98	7.13	0.030	0.74
Lying	44.58	44.16	44.39	44.51	44.09	44.66	44.29	0.106	0.78
Sitting	4.34	4.72	4.59	4.35	4.80	4.26	4.53	0.087	0.64
Singularity behavio	or (count/	hour)							
Drink water	5.04	5.19	5.15	5.10	5.11	5.14	5.12	0.015	0.23
Rooting	1.08	1.10	1.11	1.04	1.12	1.06	1.03	0.014	0.46
Posture transition (lying-sitting)	3.54	3.49	3.50	3.48	3.44	3.44	3.51	0.020	0.86
Posture transition (sitting-lying)	3.53	3.48	3.50	3.48	3.43	3.45	3.50	0.015	0.64
Biting	0.18 ^b	0.23ª	0.21 ^{ab}	0.18 ^b	0.15°	0.17 ^b	0.18 ^b	0.05	< 0.001

Table 6. Effects of different phytogenic feed additives on behavior changes in growing pigs with stressed by stocking density

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; SEM, standard error of means. Each value is the mean value of 4 replicates. ^{a-b}Means within column with different superscripts differ significantly (p < 0.05).

	PC	NC	PT1	PT2	PT3	PT4	PT5	SEM	P-value
Basic behavior (_		
Dasic Denavior ((mm/nour)								
Feed intake	4.63 ^a	4.37 ^b	4.67 ^a	4.59 ^a	4.61 ^a	4.67 ^a	4.61 ^a	0.020	< 0.001
Standing	6.77 ^a	6.32°	6.38°	6.44 ^{bc}	6.71ª	6.56 ^{ab}	6.61 ^{ab}	0.031	< 0.001
Lying	44.88°	45.51ª	45.21 ^b	45.18 ^b	44.98°	44.99°	44.96 ^{bc}	0.041	< 0.001
Sitting	3.72	3.80	3.74	3.79	3.70	3.78	3.82	0.012	0.75
Singularity beha	avior (cour	nt/hour)							
Drink water	5.34	5.28	5.27	5.30	5.44	5.38	5.29	0.026	0.59
Rooting	1.12	1.08	1.11	1.09	1.21	1.19	1.15	0.021	0.58
Posture transition (lying-sitting)	3.78	3.43	3.49	3.58	3.71	3.68	3.69	0.035	0.60
Posture transition (sitting-lying)	3.77	3.41	3.50	3.60	3.70	3.67	3.68	0.045	0.37
Biting	0.16	0.23	0.18	0.15	0.17	0.16	0.18	0.008	0.20

Table 7. Effects of different phytogenic feed additives on behavior changes in finishing pigs with stressed by stocking density

Abbreviation: PC, basal diet in animal welfare density; NC, basal diet in high stocking density; PT1, basal diet with PFA1 in high stocking density; PT2, basal diet with PFA2 in high stocking density; PT3, basal diet with PFA3 in high stocking density; PT4, basal diet with PFA4 in high stocking density; PT5, basal diet with PFA5 in high stocking density; SEM, standard error of means. Each value is the mean value of 4 replicates. ^{a-} ^eMeans within column with different superscripts differ significantly (p < 0.05).

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Abbreviation: A, Feed intake; B, Standing; C, Lying; D, Sitting; E, Drink water; F, Posture transition (lying \rightarrow standing); G, Posture transition (standing \rightarrow lying); H, Rooting; I, Biting.