

# JAST (Journal of Animal Science and Technology) TITLE PAGE

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<b>Type</b>	Research Article
<b>Title (English)</b>	Comparison between Berkshire and crossbreed on meat quality, and investigation of the relationship with fatty acid composition and meat quality
<b>Running Title (English)</b>	Comparison of meat quality and the relationship between quality parameters
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<b>Conflict of interest</b>	No potential conflict of interest relevant to this article was reported.
<b>Funding information</b>	This research was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science, and Technology (Project No. 2017R1D1A1B0403564414).
<b>Acknowledgements</b>	
<b>Availability of data and material</b>	Upon reasonable request, the datasets of this study can be made available by the corresponding author.
<b>Author Contribution</b>	Conceptualization: Seo JK Data curation: Seo JK Formal analysis: Seo JK, Eom JU Methodology: Seo JK, Yang HS Software: Seo JK, Eom JU Validation: Seo JK, Yang HS Investigation: Seo JK, Eom JU, Yang HS Writing - original draft: Seo JK Writing - review & editing: Seo JK, Eom JU, Yang HS
<b>IRB/IACUC approval</b>	All animals used in this research were approved by the Gyeongsang National University (GNU) Institutional Animal Care and Use Committee (GNU-IACUC; approval number: GNU-210614-P0058).
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6           **investigation of the relationship with fatty acid composition and meat quality**

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28 **Abstract**

29 This study aimed to compare meat quality traits between Berkshire and crossbreed  
30 (Landrace×Yorkshire×Duroc), and to investigate the relationship between meat quality traits and fatty acid  
31 composition. 20 Berkshire and 20 crossbreed pigs were used to compare pork loin quality and to determine  
32 the relationship between measured variables. 23 variables were measured including proximate composition,  
33 pH, drip loss and cooking loss, Warner–Bratzler shear force, and fatty acid composition. Berkshire had  
34 higher moisture content, pH, water-holding capacity, saturated fatty acids, and redness than the crossbreed  
35 pig ( $p<0.05$ ). The fat content and polyunsaturated fatty acid were low ( $p<0.05$ ) in Berkshire. Correlation  
36 analysis showed a negatively correlation between moisture and fat content, and a positively correlation  
37 between saturated fatty acid and fat content. Moreover, saturated fatty acid and polyunsaturated fatty acid  
38 were negatively correlated. As a result of factor analysis and partial least square regression, saturated fatty  
39 acid and polyunsaturated fatty acid were estimated to be the main factors affecting quality characteristics  
40 of pork. Pig breed is associated with differences in meat quality, and fatty acid composition can have an  
41 effect on meat quality parameters.

42

43 **Keywords:** Berkshire; LYD; Pig breed; Pork quality; Fatty acid composition; Relationship

44

## Introduction

45

46 It is known that there are more than 1,000 pig breeds throughout the world. However, since the late  
47 20th century, a relatively small number of breeds have been used for pig production due to intensive  
48 selective breeding and genetic improvement. The modern breeding environment and long-term selective  
49 breeding have resulted in improved breeding and growth rates, increased carcass yield, muscle growth  
50 efficiency, and improved intramuscular fat content [1]. Following this trend, the pork industry in Korea is  
51 dominated by the LYD, produced by crossing Landrace, Yorkshire, and Duroc, because of their improved  
52 productivity and meat quality characteristics. LYD are also widely used outside of Korea for their improved  
53 meat quality, excellent reproductive ability (high productivity), and increased muscle mass that results from  
54 crossing Landrace and Yorkshire pigs [2]. On the other hand, Berkshire have been reported to have lower  
55 productivity than LYD, but greater water holding capacity. Berkshire have deeper meat color, higher pH,  
56 lower drip and cooking loss than LYD [3]. Also, Berkshire have a high ratio of Type I muscle fiber  
57 compared to other breeds, and excellent protein solubility and water-holding capacity [4]. As a result, the  
58 meat quality characteristics are significantly different between pig breeds. With a more precise  
59 understanding of the meat quality characteristics of each breed, Korean consumers could be provided with  
60 additional purchasing opportunities for pork products along with LYD.

61 In pork, representative fatty acids in C16:0, C18:0 (in saturated fatty acid, SFA), C18:1, and C18:2 (in  
62 unsaturated fatty acid, UFA) constitute more than 80% of the total fatty acid composition. Also, long-chain  
63 fatty acids such as C18:3 and C20-22 are present in relatively high proportions. Many previous studies have  
64 argued differences in the fatty acid composition according to pig breed. Berkshire showed significantly  
65 higher saturated fatty acid (SFA) and lower mono-unsaturated fatty acid (MUFA) content than Duroc and  
66 Landrace [5]. Previous studies have suggested that differences in palmitoleic acid, oleic acid, linoleic acid,  
67 and linolenic acid between Pulawska (native species) and Polish Landrace (industrial breed) contribute to  
68 improved meat quality of native species [6]. Therefore, changes in meat quality characteristics and fatty

69 acid composition could be attributed to pig breed. Thus, the characteristics of meat that consumers can  
70 recognize will be affected by these changes.

71 The fatty acid composition could affect the firmness of adipose tissue, shelf-life, and flavor among  
72 other meat quality characteristics [7]. To summarize the arguments of authors for each factor: 1) firmness  
73 of adipose tissue: each fatty acid has a different melting point, so if the composition is different, the melting  
74 point of the whole fat is different; 2) shelf-life: the oxidation tendency of unsaturated fatty acids leads to an  
75 increase in oxidation color with an increase in lipid oxidation; 3) Flavor: changes in fatty acid composition  
76 can affect final sensory properties by causing changes in volatile compounds, which are Maillard reaction  
77 products. The previous study established the correlation between fatty acids and sensory properties [8].  
78 Among a total of nine fatty acids, only the n-6:n-3 ratio was negatively correlated with tenderness ( $r=-0.23$ ),  
79 softness ( $r=-0.26$ ), chewiness ( $r=-0.27$ ) and rate of breakdown ( $r=-0.30$ ) [8]. The proportion of unsaturated  
80 fatty acids and fat firmness were negatively correlated, and the correlation between fatty acid content and  
81 lean meat quality was insignificant [9]. Also, the correlations between fatty acid composition, protein, and  
82 fat content in Duroc, Landrace, Hampshire, and Pietrain [10]. The protein correlated positively with PUFA  
83 and correlated negatively with SFA, while fat concentration correlated negatively with PUFA [10].

84 Therefore, based on previous studies, fatty acid composition and meat quality seem to have a very  
85 high scientific relationship. Taken together, scientific evidence has demonstrated that the variation in meat  
86 quality characteristics and fatty acid composition between pig breeds is a fact. However, limited  
87 information is available regarding the relationship between pork fatty acid composition and meat quality  
88 characteristics. Therefore, the aim of this study is to not only compare meat quality between Berkshire and  
89 crossbreed, but also to characterize the relationship between pork fatty acid composition and meat quality.

90 The purpose of this study is to compare meat quality characteristics according to pig breed to identify  
91 meat quality characteristics by pig breed and investigate the relationship between fatty acid composition  
92 and meat quality properties (proximate components, pH, instrumental color, water holding capacity, and  
93 Warner-Bratzler shear force).

94

95

96

## Materials and Methods

### 97 **Sample preparation**

98 The pigs were in the same feeding condition according to the Korean Feeding Standard for Swine [11]

99 for 175-185 days. Twenty pigs for each breed were randomly selected from a local slaughterhouse in Korea.

100 A total of 40 pigs were used in the experiment. 20 Berkshire and 20 LYD pigs were slaughtered by

101 Livestock Products Sanitation Management Act. The average slaughter weight was 105-110 kg. The Korean

102 commercial procedures were applied during the slaughter, and the pork loin was removed from the carcass

103 after 24 h. The pork loins were transported to the laboratory from the slaughterhouse and analyzed after

104 refrigerating for 16 h at 4°C (2 days postmortem).

105

### 106 **Proximate composition**

107 The moisture (oven drying method, 950.46) and ash (dry ashing method, 942.05) content were

108 determined using AOAC [12], and fat content analysis was conducted using the method generated by Folch

109 [13]. The results were expressed in % of the sample. Protein content was analyzed using a nitrogen analyzer

110 (SpeedDigester K-425; Distillation Unit K-350, Büchi, Flawil, Switzerland), and % nitrogen was calculated

111 using 6.25 (conversion factor of total nitrogen to protein).

112

### 113 **Instrumental color and pH**

114 The instrumental color was measured using a colorimeter (CR-400, Konica Minolta, Tokyo, Japan)

115 and was taken 10 times per whole muscle sample. The color value was obtained from the average. The

116 measuring conditions were D65 illuminant and 2° standard observer, and Commission Internationale de

117 l'Eclairage (CIE) lightness ( $L^*$ ), redness ( $a^*$ ), and yellowness ( $b^*$ ) were determined. Before measuring, the

118 colorimeter was calibrated using a white calibration plate ( $Y = 81.2$ ;  $x = 0.3191$ ;  $y = 0.3263$ ).

119 The 3-g pork loin sample was homogenized with 27 mL of distilled water. The pH was measured using

120 a pH meter (S20 SevenEasy™, Geifensee, Switzerland) and calibrated to 7.00, 4.01, and 9.21 using a pH  
121 buffer. The measurement was repeated three times per sample, and the average value was utilized.

122

### 123 **Drip loss and Cooking loss**

124 A sample from which connective tissue and visible fat were removed was cut into 3-cm<sup>3</sup> pieces for  
125 drip loss. The experimental procedure was conducted using the method of Honikel [14], which was slightly  
126 modified. The surface of the prepared sample was lightly wiped off with a paper towel, and the initial  
127 weight was measured. The sample was hung in the middle of a plastic container, preventing contact with  
128 walls and outside air, and left to stand for 48 h at a constant temperature of 4°C then weighed. The  
129 calculation was expressed as a percentage of the difference in weight before and after standing.

130 The cooking loss samples were prepared in the form of steaks (2.5 cm height, 6 cm width, and 6 cm  
131 length), and after initially weighing, they were placed in a plastic bag and heated in a water bath at 72°C  
132 until the core temperature reached 70°C. At this time, the plastic bag was not sealed, and the prepared  
133 samples were observed with a thermocouple (HT-9815, Xintai Instrument, Guangdong, China). The weight  
134 was measured after heating was completed, and the calculation expressed the difference in weight before  
135 and after heating as a percentage. Drip loss and cooking loss were measured twice per sample, and the  
136 average value was used.

137

### 138 **Warner-Bratzler shear force (WBSF)**

139 For WBSF, 10 cores of 1.27 cm diameter were taken from each sample in the horizontal direction of  
140 the muscle fibers after measuring the cooking loss. The measurement was tested on a universal tensile  
141 testing machine (EZ-SX, Shimadzu Corp., Kyoto, Japan) with a 500-N load cell and a blade shear jig. The  
142 speed of the crosshead was set to 100 mm/min. The mean value of the cores was used for each sample.

143

### 144 **Fatty acid composition**

145 Lipid extraction was conducted by Folch [13], as well as saponification, methylation, and gas

146 chromatography under learning conditions employed by Seo [15]. The amount of fatty acids was expressed  
147 as a percent of total fatty acids.

148

## 149 **Statistical analysis**

150 Data analysis was conducted using SAS software (9.4 ver., SAS Institute Inc., NC, USA). Data were  
151 used for the analysis of a total of 40 pigs and were expressed as averages. Twenty-three dependent variables  
152 and one independent variable were used, and a general linear model was used to test the pig breed effect.  
153 One-way analysis of variance and Duncan's multiple range test were used to compare 23 dependent  
154 variables including fatty acid composition, proximate composition, color, pH, WBSF, drip loss, and  
155 cooking loss, which were tested with 95% significance. Factor analysis was conducted on all pork quality  
156 variables used in this study to find the main variables affecting pork quality. At this time, the pig breed  
157 effect was removed, and the proc factor was used for analysis. The principal component method was used  
158 for the initial factor extraction. The varimax was used as a rotated method to minimize the number of  
159 variables with high loadings for each factor and to simplify factor analysis. The results were expressed as  
160 a pattern plot of factors 1 and 2.

161

162

## 163 **Results and Discussion**

### 164 **Mean, maximum, minimum, standard deviation, and coefficient of variation of pork quality** 165 **traits from all pork loin**

166 Table 1 shows the mean, maximum, minimum, standard deviation, and coefficient of variation of the  
167 proximate composition, pH, instrumental color, drip loss, cooking loss, WBSF, and fatty acid composition  
168 of the pork loin. The mean of pH, L\*, and drip loss were 5.75, 52.25, and 2.11%, respectively. The L\* is  
169 approximately two points higher than the criteria for classification between normal and abnormal pork loin  
170 proposed by Warner [16] and Chmiel [17]. However, considering the pH and drip loss, it can be judged that

171 our pork loin was sufficiently within the normal meat range. In terms of proximate composition, fat content  
172 was approximately 3%. The fat percentage of most pork loins from pigs raised in Korea is 3% [18], which  
173 is comparable to the pork loin used in our study of common quality. C16:0, C18:0, C18:1, and C18:2 are  
174 the main fatty acids in pork loin.

175

## 176 **Effect of pig breed on meat quality traits and fatty acid composition**

177 Table 2 shows the effect of pig breeds on the physicochemical traits and fatty acid composition in pork  
178 loin. Moisture and fat content were significantly different according to the pig breed. The Berkshire had  
179 high moisture content while LYD had high-fat content ( $p<0.05$ ). According to a previous study, the fat  
180 content of Berkshire was lower than that of a Duroc used as a terminal sire among crossbred pigs [19, 20].  
181 In this study, the compared subject was LYD, and as a result, the findings were the same as in previous  
182 studies. Additionally, according to the authors, the moisture content was affected by the fat content. This is  
183 very helpful in understanding our results. Berkshire was higher in  $a^*$  and lower in  $b^*$  than LYD ( $p<0.05$ ),  
184 but  $L^*$  did not significantly differ in both pig breeds ( $p>0.05$ ). Lee [21] reported 8.47 as a result of measuring  
185 redness in 1,942 Berkshires, which was similar to our result. Also, Subramaniyan [3] compared meat quality  
186 characteristics between Berkshire and LYD, which was similar to our redness and  $b^*$  values except for  $L^*$ .  
187 Drip loss and cooking loss, which represented the water-holding capacity, were significantly higher in LYD  
188 than Berkshire ( $p<0.05$ ). Additionally, the WBSF of Berkshire was lower than that of LYD ( $p<0.05$ ).  
189 Subramaniyan [3] reported that there was no significant difference in the drip loss and WBSF between  
190 Berkshire and LYD but the cooking loss was significantly lower than that of LYD.

191 Collectively, based on the results obtained from the physicochemical properties, the main differences  
192 are fat content,  $a^*$ , pH, and water-holding capacity (drip loss and cooking loss). In addition, Barlocco [22]  
193 reported a positive correlation between intramuscular fat (IMF) and shear force ( $r=0.31$ ), as in our results,  
194 in experiments related to predictive models of IMF, moisture, and shear force in pork. On the other hand,  
195 Fortin [23] reported a negative correlation ( $r=-0.47$ ) between IMF and shear force but argued that it still  
196 needed to debate with pork tenderness and IMF levels. Therefore, considering our results, the high WBSF

197 despite the high fat content of LYD may be due to the higher cooking loss compared to Berkshire.

198 The nine fatty acids were detected in our experiment of which C16:0, C17:1, C18:0, C18:2, C20:4,  
199 SFA, and polyunsaturated fatty acid (PUFA) significantly differed according to pig breed ( $p<0.05$ ). The  
200 C16:0 and C18:0 in SFA were higher in Berkshire than in LYD, and the C17:1, C18:2, and C20:4 in  
201 unsaturated fatty acid (UFA) were lower in Berkshire than in LYD. Thus, SFA was high in Berkshire  
202 whereas PUFA was low. Alonso [24] analyzed the fatty acid composition in three crossbred pigs using  
203 different sire lines. Similar to our results, PUFA decreased in crossbred pigs with increased SFA, and there  
204 was no difference in MUFA in all crossbreeds. Also, C16:0 and C18:0 showed the highest significance  
205 level for the effect of the pig breed. However, in our study, C18:2 and C20:4 had the highest significance  
206 level in the PUFA, whereas Alonso [24] reported that C18:3 had the highest significance level. Pigs are  
207 monogastric animals and are more affected by diet systems than ruminants; this means that feed consumed  
208 by pigs is absorbed in the small intestine, and stored in tissues without any chemical changes, such as  
209 hydrogenation in ruminants [25]. Therefore, the differences in our experiment may have been due to feeding  
210 systems and breeding, and several previous studies have shown that differences in PUFA could be attributed  
211 to animal type or breeding [24]. This is the effect of the specification, and because our study made the same  
212 specification, the results of previous studies do not apply to us. It has been suggested that the changes in  
213 muscle C17:0 and C17:1 in pigs are due to endogenous synthesis of ingested dietary fiber derived from  
214 propionic acid produced by fermentation in the posterior intestine [25]. Also, according to Álvarez-  
215 Rodríguez [26], there is a greater possibility that undigested starch will decrease than the proportion of  
216 structural carbohydrates reaching the posterior intestine. Therefore, further research should be conducted  
217 in this regard. Additionally, Cannata [27] reported that C20:4 was affected by the content of intramuscular  
218 fat and increased significantly with increasing intramuscular fat. The authors found that there was a negative  
219 correlation with fat content. Although C17:1 and C20:4 occupy a small proportion of pork loin, they may  
220 be significant fatty acids in relation to meat quality.

221 The fatty acids showing significant differences in our sample are known as the major fatty acids in  
222 pork loin [25]. The melting points of C16:0, C17:1, C18:0, C18:2, and C20:4 showing significance were

223 62.9°C, 61.3°C, 69.3°C, -12°C, and -4°C, respectively. The melting point of unsaturated fatty acids in the  
224 subzero temperature range was lower than the melting point of SFAs detected in the experiment. In terms  
225 of processed meat, it can act as a negative factor with an increase in unsaturated fatty acids with a low  
226 melting point. To explain, most of the unsaturated fatty acids detected in pork exists in oil form. Thus, this  
227 causes the texture characteristics of pork fat to soften. Increased softening of pork fat is greatly influenced  
228 by increasing amounts of unsaturated fatty acids, significantly impacting overall pork texture [28].

229 In our fatty acid results, PUFA was significantly higher in LYD than in Berkshire by approximately  
230 4%; this may affect shelf life in addition to the previously described aspects of pork quality characteristics.  
231 Inserra [29] investigated fat oxidation and fatty acid composition of pork produced through different  
232 feeding systems, and the authors mentioned that PUFA in intramuscular fat, which can be easily oxidized,  
233 can provide information on meat oxidation. The authors also reported that PUFAs increased with an  
234 increase in the TBARS (2-thiobarbituric acid reactive substances). Therefore, an increase in PUFA may  
235 have a negative effect on the oxidation of pork lipids, leading to a decrease in storage properties.

236

### 237 **Relationship between meat quality traits and fatty acid composition**

238 The correlation coefficients between significant physicochemical traits are shown in Table 3. Moisture  
239 content was negatively correlated with protein, fat, and cooking loss, and it was positively correlated with  
240 drip loss. Fat content showed a positive correlation between the cooking loss and WBSF, and a negative  
241 correlation with drip loss. Additionally, WBSF showed a strong negative correlation with protein content  
242 but a positive correlation with drip loss and cooking loss. Instrumental color ( $L^*$ ,  $a^*$ , and  $b^*$ ) showed a  
243 significant correlation with pH and negative correlations in all items. Additionally,  $b^*$  showed a negative  
244 correlation with moisture content but a positive correlation with fat content. Taken together, proximate  
245 composition causes percent change between them, and this can affect the water-holding capacity in meat  
246 quality. Also, the color is closely related to pH, and  $b^*$ , in particular, will be directly related to changes in  
247 moisture and fat content.

248 The correlation between fatty acids is shown in Table 4. The relationship between saturated versus  
249 unsaturated fatty acids in this study was confirmed. The correlation coefficients between SFA and MUFA  
250 or PUFA were  $-0.47$  and  $-0.80$ , respectively, showing a stronger negative correlation between SFA and  
251 PUFA, while MUFA and PUFA were not significant. Additionally, in terms of individual fatty acids, C16:0  
252 had a strong positive correlation with C18:0, and both of them showed a generally strong negative  
253 correlation with all unsaturated fatty acids ( $r = -0.47$  to  $-0.79$ ). The C17:1 had a negative correlation with  
254 C18:0 and C18:1 but showed a positive correlation with C18:2 and C20:4. Additionally, there was a positive  
255 correlation between C18:2 and C20:4. In summary, an increase in C16:0 led to an increase in C18:0, and  
256 an increase in SFAs leads to a decrease in unsaturated fatty acids. Additionally, C17:1 and C20:4 were  
257 found to be closely related to all fatty acids except for monounsaturated fatty acids.

258 Table 5 shows the correlation between physicochemical traits and fatty acid composition. C16:0 and  
259 C18:0 exhibited similar results; positive correlation with fat, cooking loss,  $L^*$ , and  $b^*$ ; and negative  
260 correlation with drip loss and pH. Also, C16:0 exhibited a negative correlation with moisture content and  
261  $a^*$ , whereas C18:0 exhibited no significant correlation. C17:1, C18:2, and C20:4 showed opposite results  
262 with C16:0 and C18:0 and exhibited a positive correlation with moisture content and drip loss in detail, and  
263 a negative correlation with fat content, cooking loss, and  $b^*$ . Additionally, C20:4 showed a correlation  
264 between pH and  $L^*$ . The C18:3 was significant with pH and  $a^*$ , which were negatively and positively  
265 correlated, respectively. Overall, SFA showed a positive correlation with fat content, cooking loss,  $L^*$ , and  
266 pH, whereas drip loss, pH, and  $a^*$  had a negative correlation with SFA. PUFA showed a positive correlation  
267 with moisture content, drip loss, and pH, whereas fat content, cooking loss, and  $L^*$  had a negative correlation  
268 with PUFA. Additionally, MUFA was not significantly correlated with meat quality. This is because C16:1  
269 and C18:1 (approximately 46%), which account for most of the detected MUFAs, did not show a correlation  
270 with meat quality.

271 Factor analysis was performed on all variables, and the factor loading is shown in Table 6. As a result  
272 of the rotated factor analysis, all variables of the study were classified into 5 factors, and the cumulative  
273 variance was about 97%. Factor 1 was assigned the most variables and showed an explained variance of

274 34.43%. SFA, C16:0, C18:0, lightness, cooking loss, C20:4 pH, PUFA, and C18:2 were classified as factor  
275 1. In addition, SFA, C16:0, C18:0, lightness, and cooking loss showed positive factor loading, and the rest  
276 showed negative factor loading. Factor 2 was assigned MUFA and C18:1 and showed explained variance  
277 of 16.96%. Therefore, SFA, PUFA, and MUFA in factors 1 and 2 were selected as the main variables in  
278 this study considering our correlation analysis.

279 Partial least squares regression analysis was performed to figure out the causation of variables and the  
280 results are shown in Figure 1. Based on the results of factor analysis, SFA, MUFA, and PUFA were used  
281 as explanatory variables used in the PLS model, and the other variables were assigned as response variables,  
282 and the results were expressed as PLS factors 1 and 2. Figure 1. (A) and (B) are correlation loading plots,  
283 and the explanatory power of PLS factors 1 and 2 changed according to the role of MUFA. In view of the  
284 results of Figure 1. (C), it was decided that it is appropriate to use MUFA as a response variable rather than  
285 an explanatory variable because the variable importance value of MUFA is 0.8 or less. Therefore, the PLS  
286 model is shown in Figure 1. (A) which uses SFA and PUFA as explanatory variables. Figure 1. (A)  
287 explained the results of this study well. PLS factor 1 showed an R<sup>2</sup> value of 89.8% on the X-axis and 15.7%  
288 on the Y-axis, and PLS factor 2 showed an R<sup>2</sup> value of 10.2% and 10.4% on the X-axis and Y-axis,  
289 respectively. Therefore, in PLS factor 1, 89.8% of the total variables can be explained by SFA and PUFA,  
290 so it is considered to be the most important factor in this study.

291 Consequently, SFA affects fat content, water-holding capacity, and lightness, and while PUFA affects  
292 moisture and pH. These results are thought to be affected by the composition of SFA and PUFA, and  
293 changes in SFA lead to changes in fat content. Changes in fat content would lead to changes in the proximate  
294 composition result and ultimately affect water-holding capacity. Also, changes in fat directly affect  
295 lightness by changing reflectance for light. In the PLS correlation loading plot (Figure 1. (A)), SFA and  
296 PUFA are in opposite positions, and when the effect of SFA is considered, PUFA have the opposite result  
297 of SFA. Thus, only the relationship between PUFA and pH should be considered. Leite [30] reported that  
298 the effect of fat content was significant in pH, which was increased with increasing fat content. One possible  
299 logical explanation is that an increase in SFA will lead to an increase in fat content and ultimately an

300 increase in pH. Therefore, PUFA shows the opposite result from SFA, so it can be considered a very  
301 appropriate interpretation.

302

303

304

## Conclusion

305 This study aimed to investigate the effect of pig breeds on meat quality and fatty acid characteristics  
306 in pork loin. This study also aimed to figure out the relationship between the meat quality traits and fatty  
307 acid composition that determine the quality characteristics of pork using a partial least square regression.  
308 The study was conducted on the pork loin of Berkshire and LYD, which is considered normal quality. High  
309 moisture content, pH, and water-holding capacity in Berkshire were confirmed. Additionally, it was  
310 confirmed that Berkshire have more intense redness and higher SFA than LYD. As a result of conducting  
311 a correlation analysis between variables that determine pork quality characteristics, SFA was closely related  
312 to fat content, and PUFA was closely related to moisture content, and two hypotheses could be derived.  
313 First, an increase in SFA leads to an increase in fat content, which leads to an increase in cooking loss, and  
314 it can be hypothesized that  $L^*$  and  $b^*$  may increase because of an increase in fat content. Second, an increase  
315 in PUFA leads to an increase in moisture content and an increase in drip loss. As a result of the factor  
316 analysis, the main factors were SFA, MUFA, and PUFA, and a PLS model was generated using SFA and  
317 PUFA except for MUFA. As a result, SFA and PUFA have high variable explanatory power. In conclusion,  
318 the effect of pig breeds caused differences in several measurement parameters, and these differences were  
319 closely related to fatty acid composition, especially SFA and PUFA.

320

## 321 Ethics approval and consent to participate

322 All animals used in this research were approved by the Gyeongsang National University (GNU)  
323 Institutional Animal Care and Use Committee (GNU-IACUC; approval number: GNU-210614-P0058).

324

325 **Competing Interests**

326 No potential conflict of interest relevant to this article was reported.

327

328 **Acknowledgments**

329 This research was supported by Basic Science Research Program through the National Research  
330 Foundation of Korea (NRF) funded by the Ministry of Education, Science, and Technology (Project No.  
331 2017R1D1A1B0403564414).

332

333 **Author's Contribution**

334 Conceptualization: Seo JK.

335 Data curation: Seo JK.

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337 Methodology: Seo JK, Yang HS.

338 Software: Seo JK, Eom JU.

339 Validation: Seo JK, Yang HS.

340 Investigation: Seo JK, Eom JU, Yang HS.

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343

344 **Ethic Approval and Consent to Participate**

345 Not applicable.

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440 **Table 1.** Descriptive statistics for meat quality traits and fatty acid composition of pork loin

Traits	Mean	Max	Min	SD <sup>1)</sup>	CV <sup>2)</sup>
Moisture (%)	73.82	75.42	68.79	1.45	1.97
Protein (%)	22.03	25.74	19.87	1.15	5.23
Fat (%)	2.84	6.34	1.29	1.53	49.53
Ash (%)	1.32	1.56	0.85	0.19	16.23
pH	5.75	6.18	5.48	0.16	2.81
L*	52.25	57.72	45.22	2.77	5.31
a*	7.80	13.38	4.58	2.46	31.59
b*	4.36	7.64	2.41	1.10	25.29
Drip loss (%)	2.11	3.20	0.62	0.52	24.82
Cooking loss (%)	26.93	32.66	21.14	3.11	11.53
WBSF <sup>3)</sup> (N)	23.14	33.41	15.26	4.56	19.71
C14:0	1.62	2.97	1.09	0.35	21.90
C16:0	24.29	29.15	21.66	2.04	8.39
C16:1	3.79	4.46	2.55	0.37	9.73
C17:1	0.61	1.25	0.19	0.30	48.99
C18:0	11.68	15.92	9.02	1.32	11.28
C18:1	42.66	45.52	37.73	1.75	4.11
C18:2	12.17	16.05	7.34	2.14	17.58
C18:3	0.79	1.22	0.35	0.30	37.34
C20:4	1.77	3.82	0.54	0.79	44.75
SFA <sup>4)</sup>	38.02	46.74	32.70	3.02	7.95
MUFA <sup>5)</sup>	47.06	50.19	41.03	1.76	3.74
PUFA <sup>6)</sup>	14.74	19.63	8.43	2.77	18.81

441 \*, p &lt; 0.05; \*\*, p &lt; 0.01; \*\*\*, p &lt; 0.001

442 <sup>1)</sup>SD, standard deviation; <sup>2)</sup>CV, coefficient of variation; <sup>3)</sup>WBSF, Warner-Bratzler shear force; <sup>4)</sup>SFA,443 saturated fatty acid; <sup>5)</sup>MUFA, monounsaturated fatty acid; <sup>6)</sup>PUFA, polyunsaturated fatty acid.

445 **Table 2.** Effect of pig breed on meat quality properties and fatty acid composition in pork loin

Traits	Berkshire	LYD <sup>1)</sup>	SE <sup>2)</sup>
Moisture (%)	74.81**	72.83	0.28
Protein (%)	22.12	21.94	0.23
Fat (%)	1.79	3.88***	0.16
Ash (%)	1.28	1.35	0.02
pH	5.86*	5.64	0.03
L*	51.63	52.87	0.61
a*	8.35*	7.24	0.54
b*	3.92	4.80*	0.22
Drip loss (%)	1.80	2.42*	0.08
Cooking loss (%)	24.96	28.90**	0.49
WBSF <sup>3)</sup> (N)	20.44	25.83*	0.72
C14:0	1.64	1.59	0.07
C16:0	25.59*	23.00	0.35
C16:1	3.61	3.97	0.07
C17:1	0.42	0.80*	0.05
C18:0	12.49**	10.86	0.22
C18:1	42.86	42.46	0.39
C18:2	10.71	13.64***	0.34
C18:3	0.85	0.73	0.06
C20:4	1.36	2.18***	0.14
SFA <sup>4)</sup>	39.81**	36.22	0.52
MUFA <sup>5)</sup>	46.89	47.23	0.39
PUFA <sup>6)</sup>	12.92	16.55**	0.47

446 \*, p < 0.05; \*\*, p < 0.01; \*\*\*, p < 0.001

447 <sup>1)</sup>LYD, Landrace×Yorkshire×Duroc; <sup>2)</sup>SE, standard error of the means; <sup>3)</sup>WBSF, Warner-Bratzler shear

448 force; <sup>4)</sup>SFA, saturated fatty acid; <sup>5)</sup>MUFA, monounsaturated fatty acid; <sup>6)</sup>PUFA, polyunsaturated fatty acid.

**Table 3.** The correlation coefficient between meat quality traits

	Moisture	Protein	Fat	Ash	Drip loss	Cooking loss	WBSF <sup>1)</sup>	pH	L*	a*
Protein	<b>-0.35</b>									
Fat	<b>-0.55</b>	-0.13								
Ash	-0.22	-0.29	0.56							
Drip loss	<b>0.37</b>	-0.12	<b>-0.45</b>	-0.62						
Cooking loss	<b>-0.37</b>	-0.20	<b>0.60</b>	0.35	-0.25					
WBSF	-0.02	<b>-0.80</b>	<b>0.58</b>	0.64	<b>0.33</b>	<b>0.51</b>				
pH	0.13	0.00	<b>-0.53</b>	-0.49	<b>0.32</b>	<b>-0.45</b>	<b>-0.37</b>			
L*	-0.27	<b>0.35</b>	0.13	0.04	0.01	0.16	-0.17	<b>-0.55</b>		
a*	-0.19	0.09	0.26	0.07	-0.23	0.22	0.11	<b>-0.31</b>	0.00	
b*	<b>-0.60</b>	<b>0.34</b>	<b>0.48</b>	0.10	-0.09	0.29	0.01	<b>-0.38</b>	<b>0.60</b>	0.16

Bold values represent significant correlations ( $p < 0.05$ ).

<sup>1)</sup>WBSF, Warner-Bratzler shear force.

**Table 4.** The correlation coefficient between fatty acid composition

	C14:0	C16:0	C16:1	C17:1	C18:0	C18:1	C18:2	C18:3	C20:4	SFA <sup>1)</sup>	MUFA <sup>2)</sup>
C16:0	<b>0.38</b>										
C16:1	0.14	-0.05									
C17:1	<b>-0.42</b>	<b>-0.63</b>	0.17								
C18:0	0.00	<b>0.85</b>	-0.27	<b>-0.32</b>							
C18:1	-0.13	-0.19	0.19	<b>-0.47</b>	<b>-0.40</b>						
C18:2	-0.05	<b>-0.79</b>	-0.10	<b>0.56</b>	<b>-0.73</b>	-0.20					
C18:3	<b>-0.57</b>	<b>-0.47</b>	<b>-0.36</b>	0.17	-0.21	<b>0.33</b>	0.04				
C20:4	-0.16	<b>-0.71</b>	-0.02	<b>0.84</b>	<b>-0.47</b>	<b>-0.43</b>	<b>0.66</b>	0.21			
SFA	0.24	<b>0.96</b>	-0.10	<b>-0.44</b>	<b>0.95</b>	<b>-0.37</b>	<b>-0.77</b>	<b>-0.41</b>	<b>-0.56</b>		
MUFA	-0.17	-0.31	<b>0.43</b>	-0.26	<b>-0.51</b>	<b>0.96</b>	-0.13	0.28	-0.29	<b>-0.47</b>	
PUFA <sup>3)</sup>	-0.15	<b>-0.86</b>	-0.12	<b>0.69</b>	<b>-0.72</b>	-0.24	<b>0.97</b>	0.20	<b>0.82</b>	<b>-0.80</b>	-0.15

Bold values represent significant correlations ( $p < 0.05$ ).

<sup>1)</sup>SFA, saturated fatty acid; <sup>2)</sup>MUFA, monounsaturated fatty acid; <sup>3)</sup>PUFA, polyunsaturated fatty acid.

**Table 5.** Correlation coefficient between physicochemical traits and fatty acid composition in pork loin

	C14:0	C16:0	C16:1	C17:1	C18:0	C18:1	C18:2	C18:3	C20:4	SFA <sup>1)</sup>	MUFA <sup>2)</sup>	PUFA <sup>3)</sup>
Moisture	-0.17	<b>-0.31</b>	0.16	<b>0.69</b>	-0.08	<b>-0.51</b>	<b>0.38</b>	-0.06	<b>0.53</b>	-0.17	<b>-0.36</b>	<b>0.44</b>
Protein	-0.21	0.08	0.04	-0.05	0.21	0.05	-0.19	-0.07	-0.08	0.17	0.04	-0.17
Fat	0.08	<b>0.47</b>	-0.30	<b>-0.58</b>	<b>0.40</b>	0.27	<b>-0.58</b>	0.16	<b>-0.45</b>	<b>0.40</b>	0.11	<b>-0.56</b>
Ash	0.21	<b>0.50</b>	<b>-0.54</b>	<b>-0.50</b>	<b>0.50</b>	0.04	<b>-0.50</b>	0.21	<b>-0.38</b>	<b>0.47</b>	-0.16	<b>-0.47</b>
Drip loss	0.02	<b>-0.33</b>	0.21	<b>0.43</b>	<b>-0.36</b>	-0.24	<b>0.51</b>	-0.20	<b>0.35</b>	<b>-0.32</b>	-0.12	<b>0.47</b>
Cooking loss	-0.18	<b>0.46</b>	-0.24	<b>-0.35</b>	<b>0.46</b>	0.07	<b>-0.51</b>	0.06	<b>-0.37</b>	<b>0.43</b>	-0.04	<b>-0.49</b>
WBSF <sup>4)</sup>	0.15	0.25	-0.31	<b>-0.35</b>	0.15	0.07	-0.18	0.17	-0.26	0.15	-0.05	-0.19
pH	0.25	<b>-0.49</b>	0.19	0.27	<b>-0.60</b>	0.04	<b>0.61</b>	<b>-0.37</b>	<b>0.42</b>	<b>-0.54</b>	0.13	<b>0.55</b>
L*	-0.19	<b>0.35</b>	0.05	-0.21	<b>0.42</b>	0.04	<b>-0.45</b>	0.21	<b>-0.43</b>	<b>0.39</b>	0.02	<b>-0.45</b>
a*	<b>-0.57</b>	<b>-0.43</b>	<b>-0.38</b>	0.15	-0.15	0.17	0.11	<b>0.80</b>	0.30	<b>-0.34</b>	0.11	0.26
b*	0.05	<b>0.35</b>	-0.15	<b>-0.44</b>	<b>0.32</b>	0.05	-0.23	-0.04	<b>-0.44</b>	<b>0.34</b>	-0.06	-0.31

Bold values represent significant correlations ( $p < 0.05$ ).

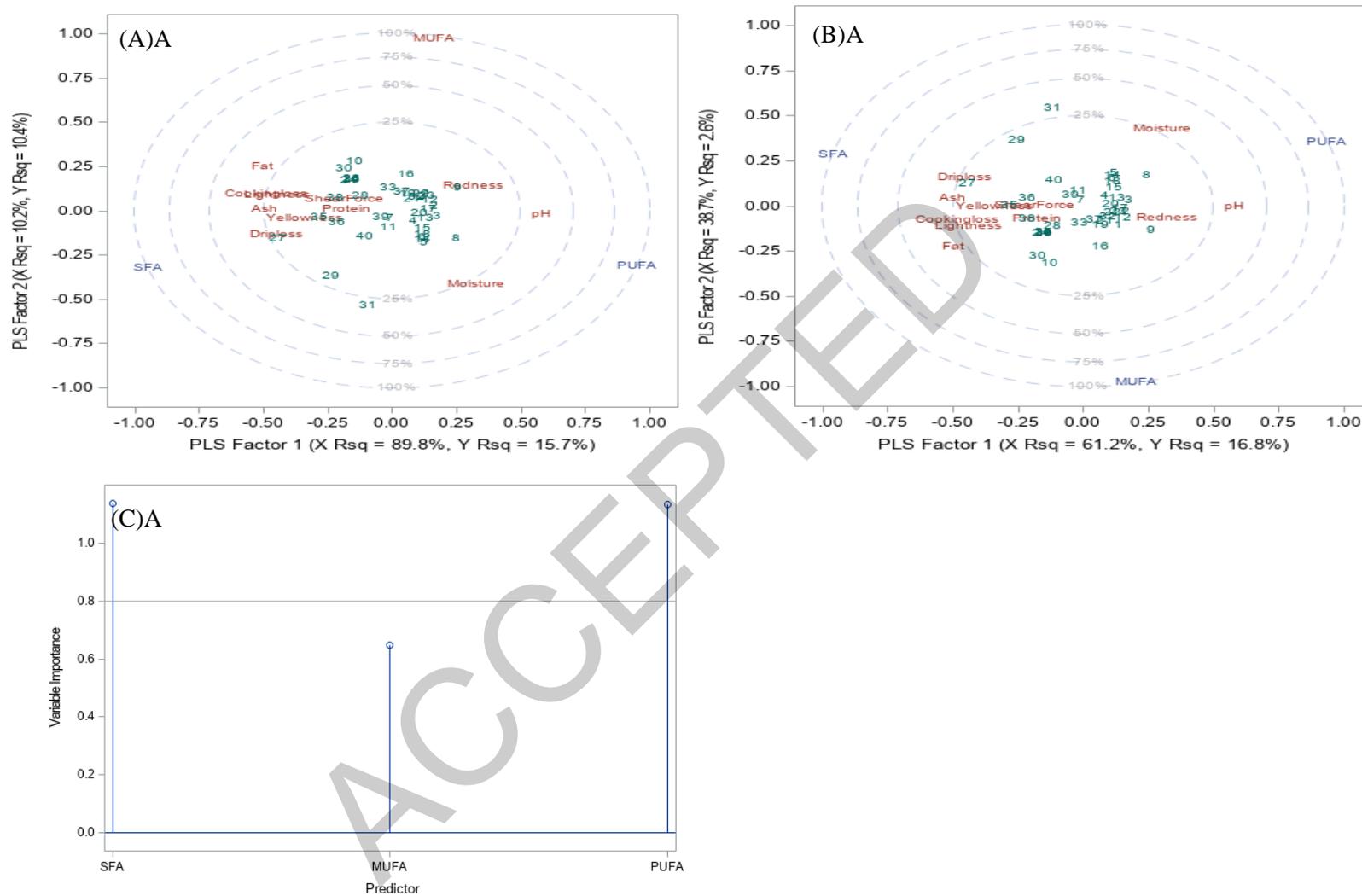
<sup>1)</sup>SFA, saturated fatty acid; <sup>2)</sup>MUFA, monounsaturated fatty acid; <sup>3)</sup>PUFA, polyunsaturated fatty acid; <sup>4)</sup>WBSF, Warner-Bratzler shear force.

**Table 6.** Determination of factors with rotated factor pattern from total measured variables

	Factor1 <sup>5)</sup>	Factor2	Factor3	Factor4	Factor5	Communality
SFA <sup>1)</sup>	0.87					0.98
C16:0	0.86					0.98
C18:0	0.85					0.94
Lightness	0.56					0.62
Cooking loss	0.53					0.54
C20:4	-0.67					0.83
pH	-0.72					0.78
PUFA <sup>2)</sup>	-0.93					0.98
C18:2	-0.93					0.91
MUFA <sup>3)</sup>		0.97				0.97
C18:1		0.96				0.96
Moisture			0.72			0.85
C16:1			0.69			0.73
C17:1			0.55			0.91
Fat			-0.55			0.75
Drip loss			-0.57			0.56
Yellowness			-0.72			0.72
C18:3				0.91		0.87
Redness				0.84		0.84
C14:0				-0.79		0.70
WBSF <sup>4)</sup>					0.92	0.91
Ash					0.61	0.72
Protein					-0.87	0.82
Eigenvalue	6.54	3.22	3.19	3.18	2.87	

<sup>1)</sup>SFA, saturated fatty acid; <sup>2)</sup>PUFA, polyunsaturated fatty acid; <sup>3)</sup>MUFA, monounsaturated fatty acid;

<sup>4)</sup>WBSF, Warner-Bratzler shear force. <sup>5)</sup>The pattern was extracted by principal components analysis and rotated with varimax method.



**Figure 1.** Rotated factor pattern plot of physicochemical properties and fatty acid composition. 42.5% and 18.93% explained variance in factor 1 and factor 2, respectively. A: C14:0, B: C16:0, C: C16:1, D: C17:1, E: C18:0, F: C18:1, G: C18:2, H: C18:3 and I: C20:4.