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| <b>Article Type</b>  | <a href="#">Research Article</a>   |
| <b>Article Title (within 20 words without abbreviations)</b>   | <a href="#">Applicability of non-invasive, digital palpation device to detection of woody breast conditions in chicken breast muscle</a>   |
| <b>Running Title (within 10 words)</b>   | <a href="#">Applicability of palpation device to detection of woody breast conditions</a>  |
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| <b>Competing interests</b>   | <a href="#">No potential conflict of interest relevant to this article was reported.</a>   |
| <b>Funding sources</b><br>State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available. | <a href="#">This work was supported by the research grant of the Gyeongsang National University in 2023.</a>   |
| <b>Acknowledgements</b>  |  |
| <b>Availability of data and material</b>   |  |
| <b>Authors' contributions</b><br>Please specify the authors' role using this form.   | <a href="#">Conceptualization: OH SH, Min B</a><br><a href="#">Data curation: OH SH, Noh E, Min B</a><br><a href="#">Formal analysis: OH SH, Noh E, Min B</a><br><a href="#">Methodology: OH SH, Noh E, Min B</a><br><a href="#">Software: OH SH, Noh E</a><br><a href="#">Validation: OH SH, Noh E, Min B</a><br><a href="#">Investigation: OH SH, Noh E, Min B</a><br><a href="#">Writing - original draft: OH SH</a><br><a href="#">Writing - review &amp; editing: OH SH, Noh E, Min B</a> |
| <b>Ethics approval and consent to participate</b>  | <a href="#">This article does not require IRB/IACUC approval because there are no human and animal participants.</a>   |

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6 **Applicability of non-invasive, digital palpation device to detection of woody breast**  
7 **conditions in chicken breast muscle**

8

9 **ABSTRACT**

10 Woody Breast (WB) is one of muscle myopathy found in chicken breast, characterized with  
11 enlarged size and extremely stiff texture. The WB condition is one of the most prevalent quality  
12 issues in the modern poultry industry. WB has been shown to be heritable, but no effective  
13 detection method of WB severity in live birds exists for the selection purpose. The objective  
14 of this study was to determine potential of a non-invasive, portable digital palpation device as  
15 WB detection method that can be used for the selection to estimate the heritability of WB. The  
16 physical and functional properties of WB was also investigated in comparison with normal  
17 breast (NB). Two hundred ten breast muscles were obtained from a local processing plant one  
18 day after harvest and sorted based on WB scoring (1 for NB and 2 and 3 for WB). The samples  
19 were subjected to physical and physicochemical analyses, determining biomechanical  
20 properties (muscle tone, stiffness, elasticity, relaxation, and creep), pH, color, cooking yield,  
21 and texture (firmness and compression energy were used for raw meat and shear force and  
22 energy for cooked meat). The least squares means of the following variables were significantly  
23 different between WB and NB ( $p < 0.01$ ): stiffness (603.4 vs 565.8; N/m), and elasticity (1.40  
24 vs 1.55). However, relaxation and creep were not significantly different ( $p > 0.05$ ). These results  
25 collectively showed that biomechanical properties of WB differ from NB. The degree of  
26 muscle stiffness in WB can be considered as a trait to be selected. The WB score showed strong  
27 negative correlations with cooking yield (-0.77) and cooked  $L^*$  (-0.74), which means that as the  
28 breast becomes harder, the cooking yield decreases, and the color becomes darker after cooking.  
29 The WB score showed high correlations with physical and functional characteristics and  
30 exhibited strong correlations with the biomechanical properties measured by the device.

31 Therefore, the results indicated that the digital palpation device has potential to detect the WB  
32 severity (degree of stiffness) of breast muscle.

33 **Keywords:** woody breast, Myoton, characteristics, broiler, breast meat

34

## 35 INTRODUCTION

36 Poultry producers' main goal is to raise chickens as fast and large as possible. This quest  
37 for growth efficiency is thought to have caused many muscle abnormalities, and a woody breast  
38 (WB) is one of them that is a significant concern for numerous producers and consumers in  
39 countries throughout the world, yet no practical solution has been proposed [1,2].

40 "Woody Breast" is the broiler breast muscle abnormality featured with enlarged size and  
41 extremely stiff textural properties often accompanies by hemorrhages on the surface, causing  
42 quality issues of broiler breast meat [2]. The WB condition is one of the most prevalent issues  
43 in the modern poultry industry. The frequency of the defect in Europe is over 30% and some  
44 have argued that similar levels may exist in the US, depending on the weight of the bird at  
45 harvest [3,4]. Given that chicken with WB is of poor economic value and are discriminated by  
46 the consumers, producers face significant economic hardship under the current production  
47 paradigm as the producers can find out occurrences of WB only after harvest [5-9].

48 One of the problems in identifying a solution is that the occurrence and severity of WB can  
49 be identified only after harvest. More specifically, WB is subjectively evaluated and separated  
50 by meat processors using visual cues and manual palpation after harvest [10]. Because more  
51 and more wholesale and retail vendors discriminate against WB chicken, WB is becoming a  
52 huge economic liability for the poultry industry [6,9]. As a result, defining and selecting against  
53 this quality defect is absolutely necessary for the future health of the poultry industry,  
54 specifically chicken (broiler) production.

55 Despite its largely unknown etiology, genetics, especially higher growth rate and breast  
56 muscle yield, are believed to be closely associated with the incidence and severity of WB [1,11-  
57 13]. Studies have indicated that WB shows moderate heritability ( $h^2 = 0.1 - 0.49$ ) [14,15].  
58 Therefore, if WB is heritable and measurable before harvest, it means that it can be controlled  
59 by genes to certain extent and therefore can be reduced by animal breeding – selection and  
60 mating. Genetic selection is a method to estimate the genetic (breeding) values of individual  
61 animals with pedigree information and environmental factors. Breeding value is the value of  
62 an individual as a (genetic) parent in a breeding program for a specific trait [16]. Bailey et al.  
63 [17] showed that the incidence of WB was reduced by 18.4% after 2 years of the genetic  
64 selection process.

65 Based on the previous studies, we hypothesized in this study that WB is heritable and  
66 therefore it can be selected for, based on the severity of WB at certain point of growth of live  
67 birds. In order to successfully carry out the effective selection, we need to know the degrees to  
68 which the severity of WB could be adequately measured in live birds and quantified as a  
69 phenotype. Then the phenotype can be subjected to a selection study in the area of animal  
70 breeding using genetic (pedigree) information and quantitative (statistical) genetics.

71 However, the major obstacle against animal breeding research for the solutions of WB is an  
72 absence in an effective, reliable detection system of WB in the breast tissues of live birds along  
73 with an accurate, quick method to evaluate the severity of WB, which would be considered as  
74 the level of hardness. There have been advancements in this area. Tijare et al. [18] described  
75 the WB scoring method with regard to level of hardness and location (cranial vs. entire fillet)  
76 and Mudalal et al. [19] used texture analysis measuring the force required to compress fillets  
77 to determine the WB severity of chicken breasts. While those evaluations methods for textural  
78 characterization of WB after harvest are useful for separating WB from normal breast, they  
79 cannot be used for the detection of WB and its severity in affected muscle tissue of live birds

80 for animal breeding.

81 Myotonometry is a non-invasive technique that can measure biomechanical characteristics  
82 of muscle tone including stiffness by applying mechanical force perpendicularly to the muscle  
83 tissues [20]. Validity, reliability, and convenience of hand-held myotonometer measuring  
84 muscle stiffness in vivo have been demonstrated by previous clinical studies [20-22]. This  
85 demonstrates that the portable myotonometer can be used to measure WB in live birds. The  
86 objective of this study was to determine the applicability of myotonometer, a non-invasive,  
87 hand-held, digital palpation device, to the quantification of the severity of WB as a trait that  
88 can be used for the selection study to estimate the heritability of WB. This study also  
89 determines the physical and functional characteristics of WB in comparison with normal breast  
90 (NB).

## 92 **MATERIALS AND METHODS**

### 93 *Sample preparation*

94 Two hundred ten chicken breast fillets (*Pectoralis major*) with woody and normal conditions  
95 (35 WB and 35 NB per replication, 3 replications) were obtained from a local processing plant  
96 one day after slaughtering. The breast fillets were sorted into woody and normal conditions at  
97 the processing plant based on the plant processing procedure. Up on receipt, the severity of  
98 woody breast conditions of the samples was evaluated based on the WB scoring system  
99 described by Tijare et al. [18] with some modifications: 1 – normal (flexible throughout), 2 –  
100 moderate (rigid mainly in the cranial region but flexible in the caudal region), and 3 – severe  
101 (extremely rigid throughout the breast meat). They were divided into 2 groups based on the  
102 WB scoring: NB - 1 and WB - 2 and 3. Subsequently, the fillets were prepared for the  
103 evaluation of physical, physicochemical, and functional properties, including pH, color, water  
104 holding capacity, cooking yield, and textural properties. All the assays were completed within

105 2 days after the receipt.

106

107 *Biomechanical properties of raw breast muscle*

108 Biomechanical properties of the samples were determined using a myotonometer, non-invasive,  
109 portable digital palpation device (MyotonPRO, Myoton, Tallinn, Estonia). The probe of the  
110 device was placed perpendicular to the cranial portion of the breast muscle and then the  
111 mechanical impulse was briefly applied to the muscle at a force of 0.60 N for 15 ms [23]. The  
112 resulting oscillation curve of the muscle were recorded and used to calculate 5 biomechanical  
113 properties of muscle, including muscle tone, stiffness, elasticity, relaxation, and creep. Those  
114 properties were defined as follows: 1) muscle tone, intrinsic tension in the relaxed muscle  
115 measured as oscillation frequency (Hz), 2) stiffness, the resistance of muscle to deformation  
116 force measured as dynamic stiffness (N/m), 3) elasticity, the ability to recover its initial shape  
117 after removal of an external force of deformation measured as logarithmic decrement, 4)  
118 relaxation, the time for muscle to recover its shape from deformation after the removal of an  
119 external force measured as mechanical stress relaxation time (ms, MSRT), and 5) creep, the  
120 gradual elongation of a tissue over time when placed under a constant tensile stress measured  
121 as ratio of deformation and relaxation time (RDRT) [20,24,25].

122

123 *pH and color*

124 The pH of each breast fillet was measured using a portable pH meter (HI98163, Hanna  
125 Instrument, Smithfield, RI) equipped with an insertion pH electrode (FC2323). The electrode  
126 tip was inserted into the fillet approximately 1 cm below the surface and the reading was  
127 recorded after stabilized. Color on the skin-side surface of the fillet was determined using a  
128 colorimeter (CR-400, Konica Minolta Sensing America, Ramsey, NJ, USA) with 8 mm  
129 aperture, illuminant D65, and 2° standard observer. The color values taken from 3 random

130 locations in the cranial region of each fillet were averaged and reported. The color of the cooked  
131 breast fillet was also determined.

132

#### 133 *Water holding capacity*

134 The water holding capacity (WHC) of the raw fillet sample was determined using a  
135 centrifugation method [26]. A cell strainer containing a thin slice of breast fillet (approximately  
136 1 g and 1-2 mm in thickness) was placed onto a tube and centrifuged at  $400 \times g$  for 1 hr at 4 °C  
137 to remove free and loosely bound water from the meat. WHC (%) was calculated as the ratio  
138 of the weight of water removed to the initial weight of the slice and represented the amount of  
139 removable water (water loss) in 100 g of meat.

140

#### 141 *Cooking yield*

142 A portion of breast fillet including the cranial region (approximately 200 g) was vacuum-  
143 packaged and cooked in a boiling water bath until the internal temperature reached 73.9 °C.  
144 After excessive moisture and fat on the surface, the cooked sample was stored at 4 °C in a  
145 refrigerator overnight prior to weighing. Cooking yield (%) was calculated as a ratio of the  
146 weight of the sample after and before cooking and expressed as a percentage. Subsequently,  
147 the cooked meat was used to determine the shear force and energy.

148

#### 149 *Firmness of raw breast fillet*

150 The firmness and compression energy of the raw breast fillet sample were determined in the  
151 cranial region using a Texture analyzer (model TA-XT2i, Texture Technologies, Hamilton,  
152 MA) equipped with a 50-kg loading cell and a cylinder probe (TA-10ss, 12.5 mm in diameter)  
153 attached to a converter (TA-71). The cranial region of the samples was compressed 10 mm in  
154 depth by the probe at the crosshead speed of 10 mm/sec. Peak force (N) and area under the

155 time-force curve (N\*sec) were reported as firmness and compression energy.

156

### 157 *Shear force of cooked breast meat*

158 Shear force and energy of the cooked breast meat were determined using the Meullenet-Owens  
159 Razor Shear (MORS) method [27], using a Texture analyzer equipped with a 5-kg loading cell  
160 and MORS blade (TA-46, 9 mm (blade) x 38 mm). The cooked breast meat was penetrated by  
161 the MORS blade perpendicular to the fiber direction. The penetration depth and crosshead  
162 speed were 20 mm and 10 mm/sec, respectively. The peak force (N) and area under the time-  
163 force curve (N\*mm) were calculated and reported as the shear force and energy, respectively.

164

## 165 **RESULTS**

166 Table 1 shows the significance of factors included in the statistical model. The effects of test  
167 and breast type were significant in almost all dependent variables except for water loss, b\* and  
168 frequency. The interaction effect was generally not significant, but it was significant in some  
169 cases and was not excluded from the statistical model.

170 Least square means of physical and physicochemical characteristics are shown in Table 2.  
171 The least squares means of the following variables were significantly different between WB  
172 and NB ( $p < 0.01$ ): DS (603.4 vs 565.8; N/m), and elasticity (1.40 vs 1.55). However, MSRT  
173 and RDRT were not significantly different ( $p > 0.05$ ). These results collectively show that the  
174 non-invasive measurements of WB differ from NB. The degree of muscle hardness in WB can  
175 be considered as a trait to be selected, and utilized for the genetic/genomic selection program  
176 collecting the measurements before harvest.

177 Pearson correlations are shown among parameters studied in Table 3. The WB score showed  
178 strong negative correlations with cooking yield (-0.77) and cooked L\*(-0.74), which means  
179 that as the breast becomes harder, the cooking yield decreases, and the color becomes darker



180 after cooking. The WB score showed high correlations with physicochemical characteristics  
181 and also exhibited strong correlations with the values measured by the device used in this study.  
182 Therefore, it was possible to measure the hardness of breast meat using the device and also  
183 observe the presence of variability.

184

## 185 **DISCUSSION**

186 There are large amounts of research going on in the industry to try and improve meat quality  
187 without decreasing the performance of the bird or negatively impacting the poultry industry  
188 and human health [6]. To date, the biological mechanism responsible for WB remains unknown  
189 [28]. About 90 studies were found in PubMed regarding WB, of which 34% were done on how  
190 it affected the different qualities of the meat; 23% on how the feeding regimen/diet affected  
191 the incidence or severity of the meat; 15.5% on how different genetic lines affected the  
192 incidence of the condition and which genes were expressed due to the condition; about 18% on  
193 the histology and morphology of the affected meat; 5.5% on the pathology; 2% on the incidence  
194 of the condition is affected by the age of the bird; and just 1% on how the time of hatch and  
195 incubation temperature can affect the morphology score [29].

196 Although WB poses trouble for poultry industries across the globe, there still hasn't been a  
197 practical solution or set of policies that are proposed or in use today because of limited research  
198 and information on it [30]. Woody breast is a phenomenon that affects the physical composition  
199 of broiler raw breast fillets. It has been reported that WB leads to multiple histological lesions  
200 such as myodegeneration and necrosis and regenerative changes [31,32].

201 Over the last 10 years, continuous selection for broilers resulted in about a 5% increase in  
202 breast meat yield [32]. This selection effort to increase bird size has led to a higher risk of  
203 disease incidences, economic loss, and welfare concerns, as well as a negative influence on  
204 meat quality traits [33,34]. Increased growth rate and continuous selection saw various

205 muscular defects [35,36] and increased muscle damage in chickens [37]. Furthermore, heavy  
206 broilers under intensive selection also had higher rates of myodegeneration and diminished  
207 thermoregulatory capacity, altered cation regulation in muscle cells and more resulting in  
208 various meat quality defects [32,38]. Why increased growth rates in modern broilers causes  
209 such myopathies is not known, but some researchers reported that heavier birds have higher  
210 incidence of severe WB [1,11-13]. Another consequence the WB phenomenon brings about is  
211 severe economic loss. As breast meat is a widely sought out source of lean meat across the  
212 nation, changes in the composition and anatomy of the breast translates to degradation of meat  
213 quality, texture, nutrition and taste, a devastating blow to the domestic poultry industry [9].

214 There have been some reports regarding hereditary muscular dystrophy in domestic fowl  
215 [39], in which affected birds exhibit a broad shallow body and short thick limb bones [40].  
216 Histopathological studies saw wide variations in fiber size, fast deposition, degeneration of the  
217 muscle fibers and more [41,42]. On top of changes in appearance, tenderness and fat content  
218 were also influenced [43,44]. Hereditary muscular dystrophy shares histological lesions with  
219 WB. Hete and Shung [45] observed that the tissue of WB chicken was stiff and had a rubbery  
220 texture compared to their control lines with flaccid muscles. There have also been approaches  
221 to studying gene expressions of WB meat.

222 Velleman [46] studied gene expressions of WB affected meat and reported that different  
223 broiler lines in the study possessed different cellular mechanisms. Lack of these nutrients could  
224 damage cells and retard integral cellular reactions and processes, raising the influence and  
225 occurrence of harmful pathological conditions. Moreover, increased selection process and  
226 higher breast yield could be the source behind the various myopathies, according to Petracci et  
227 al. [32]. This statement was highlighted in Bailey et al. [14] when they observed two different  
228 lines of broiler chicken and the chicken with higher breast yield showed a greater incidence in  
229 myopathies than the chicken with lower breast yield.

230 Challenge regarding WB research is an absence of an effective standardized scoring scale  
231 since recording and judging WB relies on a subjective scoring system. When scoring, having  
232 multiple people as opposed to one person reaching a consensus on the severity of WS and WB  
233 may be more effective until scorers are familiar with the existing or upcoming scoring systems.  
234 Fast methods to evaluate hardness would also be helpful. There have been some advancements  
235 in this area, where Tijare et al. [18] described scoring methods with regard to level of hardness  
236 and location (cranial vs. entire fillet) and Mudalal et al. [19] used texture analysis to determine  
237 the force required to compress fillets, thus determining the severity of the physical state of  
238 chicken breasts with WB.

239

## 240 **CONCLUSION**

241 In this study, WB was found to be measurable with a device and it has a variation, which  
242 means that chickens would be selected based on the severity of WB. We knew that the severity  
243 in the symptom of WB (meat) could be scored (quantified) as a phenotype. The next stage of  
244 the study will score the parameters of breast muscle during the growth of broilers (live birds)  
245 to assess the exact time or time interval of when WB can be detected and indexed to pinpoint  
246 the onset of WB in growing broilers. Each individual can be scored based on its severity, and  
247 the scores will be used to select the chickens to be mated. In sum, a selection study will be  
248 possible with this proposed study to establish a genetic line to minimize the severity of WB  
249 while maximizing the growth rate and all other economically important traits using genetic  
250 (pedigree) information and quantitative genetics (statistical animal breeding) [47].

251

## 252 **ACKNOWLEDGMENTS**

253 This work was supported by the research grant of the Gyeongsang National University in  
254 2023.

255 **REFERENCES**

- 256 1. Kuttappan VA, Brewer VB, Waldroup PW, Owens CM. Influence of growth rate on the  
257 occurrence of WS in broiler breast fillets. *Poult Sci.* 2012a;91:2677-85.  
258 <https://doi.org/10.3382/ps.2012-02259>
- 259 2. Sihvo HK, Immonen K, Puolanne E. Myodegeneration with fibrosis and regeneration in  
260 the pectoralis major muscle of broilers. *Vet Pathol.* 2014;51:619-23.  
261 <https://doi.org/10.1177/0300985813497488>
- 262 3. Kuttappan VA, Hargis BM, Owens CM. White striping and woody breast myopathies in  
263 the modern poultry industry: a review. *Poult Sci.* 2016;95:2724-33.  
264 <https://doi.org/10.3382/ps/pew216>
- 265 4. Radaelli G, Piccirillo A, Birolo M, Bertotto D, Gratta F, Ballarin C, et al. Effect of age on  
266 the occurrence of muscle fiber degeneration associated with myopathies in broiler  
267 chickens submitted to feed restriction. *Poult Sci.* 2017;96:309-19.  
268 <https://doi.org/10.3382/ps/pew270>
- 269 5. Abasht B, Mutryn MF, Michalek RD, Lee WR. Oxidative stress and metabolic  
270 perturbations in wooden breast disorder in chickens. *PLoS One.* 2016;11(4):e0153750.  
271 <https://doi.org/10.1371/journal.pone.0153750>
- 272 6. Babak MP. Morphological and molecular characterization of wooden breast myopathy in  
273 commercial broiler chickens. [Ph.D. dissertation]. Newark, DE: University of Delaware;  
274 2019.
- 275 7. Bunge J. Fast-Growth Chickens Produce New Industry Woe: ‘Spaghetti Meat’ [Internet].  
276 *Wall Street Journal.* 2019. [cited 2023 Jan 14]. <https://www.wsj.com/articles/fast-growth-chickens-produce-new-industry-woe-spaghetti-meat-11552226401>
- 278 8. Gee K. Bigger Chickens Bring a Tough New Problem: ‘Woody Breast’ [Internet]. *Wall*  
279 *Street Journal.* 2016. [cited 2023 Jan 14]. <https://www.wsj.com/articles/bigger-chickens-bring-a-tough-new-problem-woody-breast-1459207291>
- 281 9. Soglia F, Mudalal S, Babini E, Di Nunzio M, Mazzoni M, Sirri F, et al. Histology,  
282 composition, and quality traits of chicken Pectoralis major muscle affected by wooden  
283 breast abnormality. *Poult Sci.* 2016;95:651-9. <https://doi.org/10.3382/ps/pev353>
- 284 10. Lorenzi M, Mudalal S, Cavani C, Petracchi M. Incidence of white striping under  
285 commercial conditions in medium and heavy broiler chickens in Italy. *J Appl Poult Res.*  
286 2014;23:754-8. <https://doi.org/10.3382/japr.2014-00968>

- 287 11. Velleman SG, Clark DL. Histopathologic and myogenic gene expression changes  
288 associated with wooden breast in broiler breast muscles. *Avian Dis.* 2015;59:410-8.  
289 <https://doi.org/10.1637/11097-042015-Reg.1>
- 290 12. Russo E, Drigo M, Longoni C, Pezzotti R, Fasoli P, Recordati C. Evaluation of white  
291 striping prevalence and predisposing factors in broilers at slaughter. *Poult Sci.*  
292 2015;94:1843-8. <https://doi.org/10.3382/ps/pev172>
- 293 13. Trocino A, Piccirillo A, Birolo M, Radaelli G, Bertotto D, Filiou E, et al. Effect of  
294 genotype, gender and feed restriction on growth, meat quality and the occurrence of white  
295 striping and wooden breast in broiler chickens. *Poult Sci.* 2015;94:2996-3004.  
296 <https://doi.org/10.3382/ps/pev296>
- 297 14. Bailey RA, Watson KA, Bilgili SF, Avendano S. The genetic basis of pectoralis major  
298 myopathies in modern broiler chicken lines. *Poult Sci.* 2015;94:2870-9.  
299 <https://doi.org/10.3382/ps/pev304>
- 300 15. Lake JA, Dekkers JCM, Abasht B. Genetic basis and identification of candidate genes for  
301 wooden breast and white striping in commercial broiler chickens. *Sci Rep.* 2021;11:6785.  
302 <https://doi.org/10.1038/s41598-021-86176-4>
- 303 16. Weigel KA, VanRaden PM, Norman HD, Grosu H. A 100-Year Review: Methods and  
304 impact of genetic selection in dairy cattle - From daughter-dam comparisons to deep  
305 learning algorithms. *J Dairy Sci.* 2017;100(12):10234-50.  
306 <https://doi.org/10.3168/jds.2017-12954>
- 307 17. Bailey RA, Souza E, Avendano S. Characterising the influence of genetics on breast  
308 muscle myopathies in broiler chickens. *Front Physiol.* 2020;11:1-12.  
309 <https://doi.org/10.3389/fphys.2020.01041>
- 310 18. Tijare VV, Yang FL, Kuttappan VA, Alvarado CZ, Coon CN, Owens CM. Meat quality  
311 of broiler breast fillets with white striping and woody breast muscle myopathies. *Poult*  
312 *Sci.* 2016;95:2167-73. <https://doi.org/10.3382/ps/pew129>
- 313 19. Mudalal S, Lorenzi M, Soglia F, Cavani C, Petracci M. Implications of white striping and  
314 wooden breast abnormalities on quality traits of raw and marinated chicken meat. *Animal.*  
315 2015;9:728-34. <https://doi.org/10.1017/S175173111400295X>
- 316 20. Davidson MJ, Bryant AL, Bower WF, Frawley HC. Myotonometry reliably measures  
317 muscle stiffness in the thenar and perineal muscles. *Physiother Can.* 2017;69(2):104-12.  
318 <https://doi.org/10.3138/ptc.2015-85>

- 319 21. Garcia-Bernal MI, AM Herdia-Rizo, P Gonzalez-Garcia, MD Cortes-Vega, MJ Casuso-  
320 Holgado. Validity and reliability of myotonometry for assessing muscle viscoelastic  
321 properties in patients with stroke: A systematic review and meta-analysis. *Sci Rep.*  
322 2021;11(1):1-12. <https://doi.org/10.1038/s41598-021-84656-1>
- 323 22. McGowen JM, Hoppes CW, Forsse JS, Albin SR, Abt J, Koppenhaver SL. Myotonometry  
324 is capable of reliably obtaining trunk and thigh muscle stiffness measures in military  
325 cadets during standing and squatting postures. *Mil Med.* 2023.  
326 <https://doi.org/10.1093/milmed/usad179>
- 327 23. Myoton AS. MyotonPro digital palpation device user manual. Tallinn, Estonia. 2023.
- 328 24. Schneider S, Peipsi A, Stokes M, Knicker A, Abeln V. Feasibility of monitoring muscle  
329 health in microgravity environments using Myoton technology. *Med Biol Eng Comput.*  
330 2015;53:57-66. <https://doi.org/10.1007/s11517-014-1211-5>
- 331 25. Dellalana LE, Chen F, Vain A, Gandelman JS, Pöldemaa M, Chen H, et al.  
332 Reproducibility of the durometer and myoton devices for skin stiffness measurement in  
333 healthy subjects. *Skin Res Technol.* 2019;25(3):289-93. <https://doi.org/10.1111/srt.12646>
- 334 26. Bouton PE, Harris PV, Shorthose WR. The effects of ultimate pH on ovine muscle:  
335 Mechanical properties. *J Food Sci.* 1972;37(3):356-60. <https://doi.org/10.1111/j.1365-2621.1972.tb02636.x>
- 337 27. Cavitt LC, Youm GW, Meullenet JF, Owens CM, Xiong R. Prediction of poultry meat  
338 tenderness using razor blade shear, Allo-Kramer shear, and sarcomere length. *J Food Sci.*  
339 2004;69:11-5. <https://doi.org/10.1111/j.1365-2621.2004.tb17879.x>
- 340 28. Abasht B. Metabolic risk factors of wooden breast disease in commercial broiler chickens.  
341 Plant and Animal Genome XXVII Conference (January 12-16, 2019). 2019.
- 342 29. Oh SH, Oh M, Brown D. Systematic review of the different causes and effects of the  
343 wooden breast condition in poultry production. 2020 International Poultry Scientific  
344 Forum, p66.
- 345 30. Kuttappan VA, Lee YS, Erf GF, Meullenet JFC, McKee SR, Owens CM. Consumer  
346 acceptance of visual appearance of broiler breast meat with varying degrees of WS. *Poult*  
347 *Sci.* 2012b;91:1240-7. <https://doi.org/10.3382/ps.2011-01947>
- 348 31. Kuttappan VA, Shivaprasad HL, Shaw DP, Valentine BA, Hargis BM, Clark FD, et al.  
349 Pathological changes associated with WS in broiler breast muscles. *Poult Sci.*  
350 2013;92:331-8. <https://doi.org/10.3382/ps.2012-02646>

- 351 32. Petracci M, Mudalal S, Soglia F, Cavani C. Meat quality in fast-growing broiler chickens.  
352 *Worlds Poult Sci J.* 2015;71:363-74. <https://doi.org/10.1017/S0043933915000367>
- 353 33. Anthony NB. A review of genetic practices in poultry: Efforts to improve meat quality. *J*  
354 *Muscle Foods.* 1998;9:25-33. <https://doi.org/10.1111/j.1745-4573.1998.tb00641.x>
- 355 34. Julian RJ. Production and growth related disorders and other metabolic diseases of  
356 poultry-A review. *Vet J.* 2005;169:350-69. <https://doi.org/10.1016/j.tvjl.2004.04.015>
- 357 35. Hoving-Bolink AH, Kranen RW, Klont RE, Gerritsen CLM, de Greef KH. Fibre area and  
358 capillary supply in broiler breast muscle in relation to productivity and ascites. *Meat Sci.*  
359 2000;56:397-402. [https://doi.org/10.1016/S0309-1740\(00\)00071-1](https://doi.org/10.1016/S0309-1740(00)00071-1)
- 360 36. Macrae VE, Mahon M, Gilpin S, Sandercock DA, Mitchell MA. Skeletal muscle fibre  
361 growth and growth associated myopathy in the domestic chicken (*Gallus domesticus*). *Br*  
362 *Poult Sci.* 2006;47:264-72. <https://doi.org/10.1080/00071660600753615>
- 363 37. Velleman SG, Nestor KE. Effect of selection for growth rate on myosin heavy chain  
364 temporal and spatial localization during turkey breast muscle development. *Poult Sci.*  
365 2003;82:1373-7. <https://doi.org/10.1093/ps/82.9.1373>
- 366 38. Mazzoni M, Petracci M, Meluzzi A, Cavani C, Clavenzani P, Sirri F. Relationship  
367 between pectoralis major muscle histology and quality traits of chicken meat. *Poult Sci.*  
368 2015;94:123-30. <https://doi.org/10.3382/ps/peu043>
- 369 39. Julian L, Asmundson V. Muscular dystrophy of the chicken. *Muscular Dystrophy in Man*  
370 *and Animals.* 1963;458-98.
- 371 40. Wilson BW, Randall WR, Patterson GT, Entrikin RK. Major physiologic and  
372 histochemical characteristics of inherited dystrophy of the chicken. *Ann N Y Acad Sci.*  
373 1979;317:224-46. <https://doi.org/10.1111/j.1749-6632.1979.tb56531.x>
- 374 41. Jordan JP, Kratzer FH, Johnson AM, Freeman RL. Fat deposition in musculature of the  
375 genetically dystrophic chicken. *Exp Biol Med.* 1959;101:449-51.  
376 <https://doi.org/10.3181/00379727-101-24975>
- 377 42. McMurtry SL, Julian LM, Asmundson VS. Hereditary muscular dystrophy of the chicken.  
378 Quantitative histopathological findings of the Pectoralis at 6 weeks of age. *Arch Pathol.*  
379 1972;94:217-224.
- 380 43. Scholtyssek S, Bele LM, Sayre RN, Klose AA, Peterson DW. Tenderness of dystrophic

- 381 chicken meat. *Poult Sci.* 1967;46:1050-3.
- 382 44. Hete B, Shung KK. Using RF ultrasound to monitor the progression of muscular  
383 dystrophy through passive stretching. *Ultrasonics Symposium, Proceedings. Institute of*  
384 *Electrical and Electronics Engineers.* 1991;2:1123-6.
- 385 45. Peterson DW, Lilyblade AL. Relative differences in tenderness of breast muscle in normal  
386 and two dystrophic mutant strains of chickens. *J Food Sci.* 1969;34:142-5.  
387 <https://doi.org/10.1111/j.1365-2621.1969.tb00905.x>
- 388 46. Velleman SG. Relationship of skeletal muscle development and growth to breast muscle  
389 myopathies: A review. *Avian Dis.* 2015;59:525-31. [https://doi.org/10.1637/11223-](https://doi.org/10.1637/11223-063015-Review.1)  
390 [063015-Review.1](https://doi.org/10.1637/11223-063015-Review.1)
- 391 47. Cho E, Kim M, Kim JH, Roh HJ, Kim SC, Jin DH, et al. Application of genomic big data  
392 to analyze the genetic diversity and population structure of Korean domestic chickens. *J*  
393 *Anim Sci Technol.* 2023;65(5):912-21. <https://doi.org/10.5187/jast.2023.e8>



394 **Table 1.** Significance of factors included in the statistical model.

|                  | Test | Breast Type | Interaction |
|------------------|------|-------------|-------------|
| pH               | **   | **          | NS          |
| Cooking Yield    | **   | **          | **          |
| Water Loss       | NS   | **          | NS          |
| Raw L*           | **   | **          | NS          |
| Raw a*           | NS   | **          | NS          |
| Raw b*           | **   | **          | **          |
| Cooked L         | **   | **          | **          |
| Cooked a         | *    | **          | NS          |
| Cooked b         | **   | **          | NS          |
| Firmness         | **   | **          | **          |
| Work Penetration | **   | **          | **          |
| Shear Force      | **   | **          | **          |
| Shear Energy     | **   | **          | **          |
| Muscle tone      | **   | NS          | NS          |
| Stiffness        | **   | **          | NS          |
| Elasticity       | **   | **          | NS          |
| Relaxation       | **   | **          | NS          |
| Creep            | **   | **          | NS          |

395 \*\* p < 0.01; \* p < 0.05; NS: Not Significant

396 **Table 2.** Least square means of physical, physicochemical, and functional characteristics.

|                          | Test 1                      |                             | Test 2                      |                             | Test 3                      |                             |
|--------------------------|-----------------------------|-----------------------------|-----------------------------|-----------------------------|-----------------------------|-----------------------------|
|                          | Normal                      | Wooden                      | Normal                      | Wooden                      | Normal                      | Wooden                      |
| pH                       | 6.00±0.02 <sup>a</sup> (30) | 6.12±0.01 <sup>b</sup> (33) | 5.79±0.01 <sup>a</sup> (35) | 5.96±0.01 <sup>b</sup> (34) | 5.76±0.01 <sup>a</sup> (33) | 5.92±0.01 <sup>b</sup> (31) |
| Cooking Yield (%)        | 73.5±0.46 <sup>a</sup> (31) | 60.2±0.43 <sup>b</sup> (35) | 72.3±0.45 <sup>a</sup> (33) | 66.7±0.44 <sup>b</sup> (34) | 69.9±0.44 <sup>a</sup> (34) | 59.5±0.43 <sup>b</sup> (35) |
| Water Loss (%)           | 9.22±0.46 <sup>a</sup> (35) | 11.8±0.46 <sup>b</sup> (35) | 9.95±0.46 <sup>a</sup> (35) | 11.7±0.48 <sup>b</sup> (32) | 9.57±0.46 <sup>a</sup> (35) | 11.9±0.47 <sup>b</sup> (33) |
| Raw L*                   | 63.3±0.38 <sup>a</sup> (35) | 66.2±0.38 <sup>b</sup> (34) | 61.9±0.38 <sup>a</sup> (35) | 65.9±0.38 <sup>b</sup> (34) | 61.4±0.39 <sup>a</sup> (33) | 64.6±0.38 <sup>b</sup> (35) |
| Raw a*                   | 0.73±0.14 <sup>a</sup> (34) | 1.56±0.14 <sup>b</sup> (33) | 0.89±0.14 <sup>a</sup> (34) | 1.35±0.13 <sup>b</sup> (35) | 0.55±0.14 <sup>a</sup> (31) | 1.67±0.13 <sup>b</sup> (35) |
| Raw b*                   | 6.96±0.32 <sup>a</sup> (35) | 9.74±0.33 <sup>b</sup> (33) | 8.90±0.32 <sup>a</sup> (35) | 9.65±0.36 <sup>a</sup> (27) | 9.05±0.33 <sup>a</sup> (33) | 10.4±0.32 <sup>b</sup> (34) |
| Cooked L*                | 83.1±0.34 <sup>a</sup> (33) | 76.4±0.34 <sup>b</sup> (34) | 83.1±0.34 <sup>a</sup> (33) | 78.4±0.33 <sup>b</sup> (35) | 82.7±0.34 <sup>a</sup> (33) | 75.5±0.34 <sup>b</sup> (34) |
| Cooked a*                | 0.75±0.09 <sup>a</sup> (30) | 1.98±0.09 <sup>b</sup> (33) | 0.71±0.09 <sup>a</sup> (34) | 1.93±0.09 <sup>b</sup> (34) | 1.09±0.09 <sup>a</sup> (35) | 1.97±0.09 <sup>b</sup> (34) |
| Cooked b*                | 14.9±0.18 <sup>a</sup> (31) | 15.8±0.19 <sup>b</sup> (30) | 15.5±0.18 <sup>a</sup> (33) | 16.2±0.18 <sup>b</sup> (33) | 15.6±0.17 <sup>a</sup> (35) | 16.0±0.18 <sup>a</sup> (32) |
| Firmness (N)             | 14.9±1.27 <sup>a</sup> (35) | 33.6±1.29 <sup>b</sup> (34) | 14.3±1.27 <sup>a</sup> (35) | 37.5±1.27 <sup>b</sup> (35) | 14.7±1.27 <sup>a</sup> (35) | 46.2±1.27 <sup>b</sup> (35) |
| Work Penetration (N*sec) | 47.0±3.58 <sup>a</sup> (34) | 103±3.69 <sup>b</sup> (32)  | 46.5±3.69 <sup>a</sup> (32) | 118±3.69 <sup>b</sup> (32)  | 50.0±3.53 <sup>a</sup> (35) | 151±3.53 <sup>b</sup> (35)  |
| Shear Force (N)          | 10.9±0.39 <sup>a</sup> (29) | 12.6±0.37 <sup>b</sup> (33) | 10.8±0.37 <sup>a</sup> (33) | 12.3±0.36 <sup>b</sup> (35) | 10.8±0.37 <sup>a</sup> (33) | 15.8±0.37 <sup>b</sup> (33) |
| Shear Energy (N*mm)      | 15.1±0.56 <sup>a</sup> (31) | 18.7±0.53 <sup>b</sup> (34) | 14.5±0.53 <sup>a</sup> (35) | 17.1±0.53 <sup>b</sup> (35) | 14.3±0.57 <sup>a</sup> (30) | 23.8±0.54 <sup>b</sup> (33) |
| Muscle tone (Hz)         | 25.1±0.21 <sup>a</sup> (33) | 24.9±0.22 <sup>a</sup> (31) | 26.1±0.21 <sup>a</sup> (34) | 25.5±0.21 <sup>a</sup> (34) | 25.8±0.21 <sup>a</sup> (34) | 26.0±0.22 <sup>a</sup> (30) |
| Stiffness (N/m)          | 508±7.03 <sup>a</sup> (33)  | 575±6.92 <sup>b</sup> (34)  | 546±6.92 <sup>a</sup> (34)  | 602±6.92 <sup>b</sup> (34)  | 546±6.83 <sup>a</sup> (35)  | 612±6.92 <sup>b</sup> (34)  |
| Elasticity               | 1.66±0.02 <sup>a</sup> (35) | 1.45±0.02 <sup>b</sup> (31) | 1.56±0.02 <sup>a</sup> (31) | 1.32±0.02 <sup>b</sup> (35) | 1.52±0.02 <sup>a</sup> (34) | 1.33±0.02 <sup>b</sup> (34) |
| Relaxation (ms)          | 9.87±0.12 <sup>a</sup> (32) | 9.13±0.11 <sup>b</sup> (34) | 9.12±0.11 <sup>a</sup> (34) | 8.65±0.11 <sup>b</sup> (33) | 9.26±0.11 <sup>a</sup> (35) | 8.50±0.12 <sup>b</sup> (32) |
| Creep                    | 0.64±0.01 <sup>a</sup> (33) | 0.60±0.01 <sup>b</sup> (34) | 0.59±0.01 <sup>a</sup> (34) | 0.58±0.01 <sup>a</sup> (35) | 0.60±0.01 <sup>a</sup> (35) | 0.57±0.01 <sup>b</sup> (31) |

397 \* Subscripts represent the comparative results of the breast condition within each test. Different letters indicate significant differences  
 398 (p<0.05).

399 \*\* The numbers in parentheses represent the sample size.

400

401 **Table 3.** Pearson correlations among parameters studied.

|                       | (2)  | (3)   | (4)   | (5)   | (6)   | (7)    | (8)   | (9)   | (10)   | (11)  | (12)  | (13)  | (14)  | (15)   | (16)  | (17)   | (18)   | (19) <sup>1</sup> |
|-----------------------|------|-------|-------|-------|-------|--------|-------|-------|--------|-------|-------|-------|-------|--------|-------|--------|--------|-------------------|
| (1) WB Score          | 0.48 | -0.77 | 0.23  | 0.52  | 0.39  | 0.36   | -0.74 | 0.68  | 0.28   | 0.77  | 0.77  | 0.51  | 0.59  | -0.07* | 0.56  | -0.61  | -0.38  | -0.30             |
| (2) pH                |      | -0.44 | 0.08* | 0.48  | 0.27  | -0.06* | -0.48 | 0.38  | 0.07*  | 0.37  | 0.34  | 0.17  | 0.24  | -0.29  | 0.13  | -0.15  | 0.00*  | 0.05*             |
| (3) Cooking Yield     |      |       | -0.34 | -0.45 | -0.45 | -0.40  | 0.87  | -0.70 | -0.24  | -0.78 | -0.76 | -0.60 | -0.69 | 0.02*  | -0.56 | 0.58   | 0.40   | 0.34              |
| (4) Water Loss        |      |       |       | 0.27  | 0.17  | 0.25   | -0.33 | 0.26  | 0.10*  | 0.32  | 0.31  | 0.08* | 0.14  | -0.13* | 0.16  | -0.33  | -0.09* | -0.09*            |
| (5) Raw L*            |      |       |       |       | 0.08* | 0.40   | -0.43 | 0.32  | 0.21   | 0.42  | 0.44  | 0.10* | 0.14  | -0.39  | 0.15  | -0.41  | 0.04*  | 0.09*             |
| (6) Raw a*            |      |       |       |       |       | 0.20   | -0.59 | 0.53  | -0.04* | 0.41  | 0.39  | 0.35  | 0.41  | 0.05*  | 0.28  | -0.25  | -0.23  | -0.22             |
| (7) Raw b*            |      |       |       |       |       |        | -0.38 | 0.30  | 0.47   | 0.33  | 0.35  | 0.15  | 0.17  | -0.12* | 0.26  | -0.42  | -0.15  | -0.10*            |
| (8) Cooked L*         |      |       |       |       |       |        |       | -0.81 | -0.37  | -0.76 | -0.75 | -0.60 | -0.68 | -0.02* | -0.59 | 0.56   | 0.42   | 0.35              |
| (9) Cooked a*         |      |       |       |       |       |        |       |       | 0.27   | 0.67  | 0.63  | 0.50  | 0.54  | 0.02*  | 0.53  | -0.50  | -0.37  | -0.31             |
| (10) Cooked b*        |      |       |       |       |       |        |       |       |        | 0.25  | 0.29  | 0.08* | 0.10* | -0.00* | 0.25  | -0.27  | -0.15  | -0.12*            |
| (11) Firmness         |      |       |       |       |       |        |       |       |        |       | 0.99  | 0.61  | 0.67  | 0.11*  | 0.66  | -0.67  | -0.51  | -0.45             |
| (12) Work Penetration |      |       |       |       |       |        |       |       |        |       |       | 0.59  | 0.65  | 0.10*  | 0.67  | -0.70  | -0.51  | -0.44             |
| (13) Shear Force      |      |       |       |       |       |        |       |       |        |       |       |       | 0.96  | 0.23   | 0.49  | -0.30  | -0.40  | -0.36             |
| (14) Shear Energy     |      |       |       |       |       |        |       |       |        |       |       |       |       | 0.24   | 0.52  | -0.32  | -0.44  | -0.39             |
| (15) Muscle tone      |      |       |       |       |       |        |       |       |        |       |       |       |       |        | 0.65  | -0.12* | -0.79  | -0.81             |
| (16) Stiffness        |      |       |       |       |       |        |       |       |        |       |       |       |       |        |       | -0.69  | -0.97  | -0.95             |
| (17) Elasticity       |      |       |       |       |       |        |       |       |        |       |       |       |       |        |       |        | 0.56   | 0.52              |
| (18) Relaxation       |      |       |       |       |       |        |       |       |        |       |       |       |       |        |       |        |        | 0.99              |

402 <sup>1</sup>(19) Creep

403 \* The correlation coefficients are not significantly different from zero ( $p > 0.05$ ).

404 WB: Woody Breast

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