

1 JAST (Journal of Animal Science and Technology) TITLE PAGE

2 Upload this completed form to website with submission

ARTICLE INFORMATION	Fill in information in each box below
Article Type	Research article
Article Title (within 20 words without abbreviations)	Evaluation of forage production, feed value, and ensilability of proso millet (<i>Panicum miliaceum</i> L.)
Running Title (within 10 words)	Proso millet as conserved forage
Author	Sheng Nan Wei ¹ , Eun Chan Jeong ¹ , Yan Fen Li ¹ , Hak Jin Kim ² , Farhad Ahmadi ² , Jong Geun Kim ^{1,2}
Affiliation	¹ Graduate School of International Agricultural Technology, Seoul National University, Pyeongchang 25354, Korea ² Research Institute of Eco-friendly Livestock Science, Institute of GreenBio Science Technology, Seoul National University, Pyeongchang 25354, Korea
ORCID (for more information, please visit https://orcid.org)	Sheng Nan Wei (https://orcid.org/0000-0001-5117-5140) Eun Chan Jeong (https://orcid.org/0000-0002-6559-2743) Yan Fen Li (https://orcid.org/0000-0002-7318-7318) Hak Jin Kim (https://orcid.org/0000-0002-7279-9021) Farhad Ahmadi (https://orcid.org/0000-0002-8760-053x) Jong Geun Kim (https://orcid.org/0000-0003-4720-1849)
Competing interests	No potential conflict of interest relevant to this article was reported.
Funding sources State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available.	Cooperative Research Program for Agriculture Science & Technology Development (Project No. PJ01401903)
Acknowledgements	Not applicable.
Availability of data and material	Upon reasonable request, the datasets of this study can be available from the corresponding author.
Authors' contributions Please specify the authors' role using this form.	Conceptualization: Wei, SN, Kim, JG Data curation: Wei, SN, Kim, JG, Farhad A. Formal analysis: Jeong, EC, Li, YF, Kim, HJ Methodology: Wei, SN, Jeong, EC, Kim, HJ Software: Li, YF, Wei, SN Validation: Kim, JG, Farhad, A. Investigation: Kim, HJ, Li, YF Writing - original draft: Wi, SN, Kim, JG Writing - review & editing: Kim, JG, Wei, SN, Farhad, A.
Ethics approval and consent to participate	This article does not require IRB/IACUC approval because there are no human and animal participants.

3 CORRESPONDING AUTHOR CONTACT INFORMATION

For the corresponding author (responsible for correspondence, proofreading, and reprints)	Fill in information in each box below
First name, middle initial, last name	Jong Geun Kim
Email address – this is where your proofs will be sent	forage@snu.ac.kr
Secondary Email address	silagekim@gmail.com
Address	Graduate School of International Agricultural Technology, Seoul National University, Pyeongchang 25354, Korea
Cell phone number	+82-10-3169-9377
Office phone number	+82-33-339-5728
Fax number	+82-33-339-5763

4

Running title: Proso millet as conserved forage

**Evaluation of forage production, feed value, and ensilability of
proso millet (*Panicum miliaceum* L.)**

Sheng Nan Wei¹, Eun Chan Jeong¹, Yan Fen Li¹, Han Jin Kim², Farhad Ahmadi², and Jong Geun
Kim^{1, 2*}

¹ Graduate School of International Agricultural Technology, Seoul National University, Pyeongchang,
25354, South Korea

² Research Institute of Eco-friendly Livestock Science, Institute of GreenBio Science Technology,
Seoul National University, Pyeongchang, 25354, South Korea

Correspondence

Jong Geun Kim, Graduate School of International Agricultural Technology and Institute of Eco-
friendly Livestock science, Institute of GreenBio Science Technology, Seoul National University, 1447
Pyeongchang-daero, Pyeongchang, Kangwon, 25354, South Korea

E-mail: forage@snu.ac.kr

Tel: +82-33-339-5728

Fax: +82-33-339-5727

25 **Abstract**

26 Whole-plant corn (*Zea mays* L.) and sorghum-sudangrass hybrid (*Sorghum bicolor* L.) are major
27 summer crops that can be fed as direct-cut or silage. Proso millet is a short-season growing crop with
28 distinct agronomic characteristics that can be productive in marginal lands. However, information is
29 limited about the potential production, feed value, and ensilability of proso millet forage. We evaluated
30 proso millet as a silage crop in comparison with conventional silage crops. Proso millet was sown on
31 June 8 and harvested on September 5 at soft-dough stage. Corn and sorghum-sudangrass hybrid were
32 planted on May 10 and harvested on September 10 at the half milk-line and soft-dough stages,
33 respectively. The fermentation was evaluated at 1, 2, 3, 5, 10, 15, 20, 30, and 45 days after ensiling.
34 Although forage yield of proso millet was lower than corn and sorghum-sudangrass hybrid, its relative
35 feed value was greater than sorghum-sudangrass hybrid. Concentrations of dry matter (DM), crude
36 protein, and water-soluble carbohydrate decreased commonly in the ensiling forage crops. The DM
37 loss was greater in proso millet than those in corn and sorghum-sudangrass hybrid. The *in vitro* dry
38 matter digestibility declined in the forage crops as fermentation progressed. In the early stages of
39 fermentation, pH dropped rapidly, which was stabilized in the later stages. Compared to corn and
40 sorghum-sudangrass hybrid, the concentration of ammonia-nitrogen was greater in proso millet. The
41 count of lactic acid bacteria reached the maximum level on day 10, with the values of 6.96, 7.77, and
42 6.95 log₁₀ cfu/g fresh weight for proso millet, corn, and sorghum-sudangrass hybrid, respectively. As
43 ensiling progressed, the concentrations of lactic acid and acetic acid of the three crops increased and
44 lactic acid proportion became higher in the order of sorghum-sudangrass hybrid, corn, and proso millet.
45 Overall, the shorter, fast-growing proso millet comparing with corn and sorghum-sudangrass hybrid
46 makes this forage crop an alternative option, particularly in areas where agricultural inputs are limited.
47 However, additional research is needed to evaluate the efficacy of viable strategies such as chemical
48 additives or microbial inoculants to minimize ammonia-nitrogen formation and DM loss during

ensuring a continuous supply of forage to animals [9, 10]. To our knowledge, few studies have investigated the fermentation dynamics of proso millet forage. The purpose of this research was to provide basic information about the ensiling feasibility of forage from proso millet in comparison to commonly cultivated summer crops (whole-plant corn and sorghum-sudangrass hybrid).

MATERIALS AND METHODS

Crop establishment and management

Establishment of experimental plots was made at the experimental site of Seoul National University, Pyeongchang Campus (located at 37° 32' 40" N, 128° 26' 33" E, average altitude is about 550 m above sea level) during the summer season of 2019. A detailed description of meteorological data including temperature and precipitation throughout the growing season (May to September, 2019) is illustrated in Figure 1. During the growing season, temperature ranged from 15.9 to 26.8°C (average = 21.5°C). Soil analysis on the 0–15-cm soil depth of the experimental site showed that it was slightly acidic (pH 6.55; soil:water suspension = 1:5), with 14.1% organic matter, 0.12% total nitrogen, and a cation exchange capacity of 16.5 cmol(+)/kg. Concentration of exchangeable cations including Ca, K, Mg, and Na averaged 1.75, 4.01, 0.92, and 0.10 mg/kg, respectively. For the three crops, nitrogen, phosphorus, and potassium fertilizers were applied at a rate of 200, 150, and 150 kg/ha, respectively. After preparation of seedbed, seeds were sown manually and grown on 3 replicate plots/each crop. Each plot was 3 m × 5 m in size. Proso millet (*Panicum miliaceum* L. var. Geumsilchal) was planted on June 8 at a seeding rate of 20 kg/ha, and harvested on September 5. Sorghum-sudangrass hybrid (*Sorghum bicolor* L. var. Turbo-gold) was sown at a seeding rate of 40 kg/ha on May 10 and harvested on September 10. Corn (*Zea mays* L. var. Gwangpyeongok) was sown on May 10 at a plant-to-plant distance of 20 cm and an inter-row spacing of 75 cm. Whole-crop corn was harvested on September 10. Sorghum-sudangrass hybrid and proso millet were harvested when they reached soft-dough stage

of the seedhead. Whole-crop corn was harvested at about the half milk-line stage, which is a reliable visionary criterion indicating the optimum time to harvest whole plant for silage making [11]. This was accomplished by splitting the corn ear in the center and visually inspecting the kernel milkline. Whole-crop corn was fractionated into cob (containing kernel and rachis) and stover component that was consisted of the remaining components of the plant after cob removal [12]. These fractions were separately weighed and approximately 1-kg representative subsamples were collected for dry matter (DM) determination. The proportion of these fractions in the whole plant was then calculated. Forage yield was determined by manually harvesting the forage material in the whole plot and calculating the fresh forage yield, which was then converted to units of fresh and DM/hectare.

Silage preparation

At harvest, four whole-crop plants from center rows in each plot were randomly selected and chopped into approximately 2–3 cm long pieces using a chopper (Richi Machinery Co., Ltd, Henan, China). The chopped crops were grouped into separate piles per each plot for silage experiment. The representative allotments were also collected for quality assessment of fresh biomass before ensiling. Ensiling was made by packing approximately 600 g chopped material into plastic film bags (28 cm × 36 cm). The bags were vacuum-sealed (Zhejiang Hongzhan Packing Machinery Co., Ltd, Wenzhou city, China) and stored in a dark and dry condition at room temperature (about 22°C). Bags were randomly opened on days 1, 2, 3, 5, 10, 15, 20, 30, and 45 of ensiling for quality assessment of silage fermentation. Silos were weighed at designated openings for DM loss determination [13]. Number of replicate silos for each crop at each opening was 3. Therefore, the design arrangement for the three forage types in the silage trial was as follows: 3 forage types × 9 silo openings × 3 replications, resulting in formation of a total of 81 silos. At each silo opening, the ensiled material inside each silo was emptied, mixed thoroughly and divided into 3 representative portions. The first portion was dried (65°C) to a constant weight and used for the chemical composition analysis. The second portion was

120 stored in a freezer at -80°C (TSE400D, Thermo Fisher Scientific, Waltham, MA, USA) for
121 quantification of organic acids and ammonia nitrogen. The third subsample was used for enumeration
122 of microbial population in ensiled biomass.

123 **Analytical analyses**

124 A 10-g fresh silage sample was placed into a 250 mL conical flask and covered with 100 mL distilled
125 water. The flasks were shaken for 1 h on a mechanical shaker (Green Sseriker, Vision Scientific,
126 Gyeonggi-Do, Korea) and stored in refrigerator for 24 h. The conical flasks were shaken by hand every
127 2 hours during refrigeration. The mixture was filtered through a filter paper (Whatman No. 6,
128 Advantech, Zurich, Switzerland). Silage pH was determined in the filtrate with a pH meter (AB 150,
129 Fisher Scientific International, Inc., Pittsburgh, PA, US). A 1.5 mL portion of the filtrate was used for
130 analysis of the organic acid concentration using high performance liquid chromatography (HPLC,
131 Agilent Technologies, Santa Clara, CA, US) equipped with a refractive index detector [8]. Ammonia
132 nitrogen ($\text{NH}_3\text{-N}$) was analyzed via the method described by Broderick and Kang [14]. The spread-
133 plate method [15] was used to enumerate the population of microorganisms. In brief, a 10-g sample
134 was diluted with 90 mL sterilized saline solution (8.50 g/L NaCl) and shaken for 1 h. Lactic acid
135 bacteria (LAB), molds, and total microorganisms were enumerated on Rogosa, and Sharpe
136 agar medium, potato dextrose agar, and plate count agar media, respectively. The limit of detection
137 was $2 \log_{10}$ CFU/g fresh mass.

138 Dry matter concentration in ensiled material was determined in triplicate at 65°C in a forced drying
139 oven for 72 h. The dried samples were ground to pass through a 1 mm screen (Thomas Scientific, Inc.,
140 Swedesboro, NJ, USA) for nutrient composition analysis. Total nitrogen was quantified via the Dumas
141 method [16], and crude protein (CP) was calculated as $\text{nitrogen} \times 6.25$. Acid detergent fiber (ADF)
142 and neutral detergent fiber (NDF) were measured following the method of Van Soest et al. [17]. Water-
143 soluble carbohydrate (WSC) was analyzed via a modification of the anthrone method proposed by

144 Yemm and Willis [18].

145 ***In vitro* dry matter digestibility**

146 *In vitro* DM digestibility (IVDMD) was performed in triplicate using an Ankom Daisy^{II} incubator
147 (ANKOM Technologies, Inc., Fairport, NY, USA) [19], as described by Goering and Van Soest [20].

148 Ground samples (0.5–0.6 g) were weighed into F57 filter bags and sealed using a heat sealer. Samples

149 were evenly distributed on both sides of the digestion jars. Then, 1330 mL buffer solution A and 266

150 mL buffer solution B were added to each jar. Two ruminally cannulated Holstein steers were selected

151 and their rumen fluid was collected before the morning feed and passed through four layers of

152 cheesecloth. Then, 400 mL rumen fluid was added to the buffer solution and samples. The digestion

153 jar was purged with CO₂ gas for 30 s and then closed with a lid. The jars were incubated at 39°C for

154 48 h. Undigested NDF residues in original bags were extracted using an ANKOM²⁰⁰⁰ fiber analyzer.

155 **Statistical analysis**

156 Field experiment was arranged in a completely randomized block design with three replications. Data

157 were subjected to analysis of variance (ANOVA) using the general linear model (GLM) in SPSS (IBM

158 SPSS Statistics, Version 24.0 Armonk, NY, INM Corp). Individual plot was regarded as the

159 experimental unit in the model for analysis of data from the field experiment (Table 1). Individual silo

160 served as the experimental unit in the model for analysis of data from silage experiment. Prior to

161 statistical analysis, microbial data (Table 4) were logarithmically transformed. Mean treatment

162 differences were obtained by Duncan's multiple range tests, with a statistical significance level of 5%.

163

RESULTS AND DISCUSSION

Forage quality and yield

Yield and forage quality of experimental forage crops are presented in Table 1. Forage DM concentration was greatest in proso millet (303 g/kg), intermediate in corn (277 g/kg), and lowest in sorghum-sudangrass hybrid (193 g/kg). Whole-plant corn had the highest relative feed value (RFV) of 117, which was 20 and 40 units higher on average than proso millet and sorghum-sudangrass hybrid, respectively. A forage crop with an RFV between 103 and 124 is considered a high-quality forage [21], indicating the superiority of corn over sorghum-sudangrass hybrid and proso millet forage. Similar to our observations, Jahansouza et al. [22] also reported a similar trend in fresh forage yield. Concentration of total digestible nutrients was highest in corn (667 g/kg DM), intermediate with proso millet (631 g/kg DM), and lowest with sorghum-sudangrass hybrid (541 g/kg DM). In general, the forage nutritive value of proso millet is comparable to the value reported by Kim et al. [23] harvesting “Geumsilchal” variety in reclaimed lands located in Sihwa (Korea).

Forage yield was significantly different by forage types, with proso millet producing the least DM. The forage DM yield was greater in the order of sorghum-sudangrass hybrid (23.5 t/ha), corn (18.7 t/ha), and proso millet (7.68 t/ha). Forage yield of proso millet (fresh or DM basis) agrees with the values reported by Shin et al. [24]. Calamai et al. [4] also reported that total dry biomass in proso millet averaged 6.43 t/ha. Data of NDF and CP concentration of these forage crops is previously reported [8]. Neutral detergent fiber was highest in sorghum-sudangrass hybrid, intermediate in proso millet, and lowest in corn. No difference existed in CP concentration among crops, averaging 58 g/kg DM.

Chemical composition during ensiling

Changes in DM loss and chemical composition of the three forage crops during ensiling are reported in Table 2. As ensiling progressed, DM loss occurred in all crops, with proso millet losing the most

DM than corn or sorghum-sudangrass hybrid, most likely because a higher number of epiphytic molds existed on proso millet biomass. Loss of DM was faster in proso millet during the first day of fermentation, which may be justified by the significantly greater population of total microorganisms in fresh mass of proso millet than in corn or sorghum-sudangrass hybrid (Table 4). Microbial degradation of nutrients into carbon dioxide and water could possibly explain loss of DM with ensiling [25, 26]. Crude protein concentration displayed a downward trend during the ensiling process, which is suggestive of protein degradation with ensiling. A downward trend was also observed in NDF concentration of all forage crops with ensiling. From day 0 to 45, NDF concentration of proso millet decreased from 607 to 591 g/kg DM, which is less than the corresponding values in corn and sorghum-sudangrass hybrid. Chen et al. [26] suggested that hemicellulose degradation during the ensiling process is mainly responsible for NDF reduction with ensiling. This loss could be due to a combination of enzymatic and acid hydrolysis of the more digestible cell-wall fractions during the fermentation [10, 27]. After 45 days of ensiling, ADF concentration of proso millet silage declined by about 20 g/kg DM. Similar decreases also occurred for corn and sorghum-sudangrass hybrid.

Fermentation quality during ensiling

Changes in silage pH as a function of fermentation time are illustrated in Figure 2. The day-0 pH of corn crop (5.80) was generally lower than proso millet or sorghum-sudangrass hybrid (mean 6.05), which is in agreement with the mean values (5.50 to 6.0) reported for the different forages after chopping [28, 29]. Silage pH of corn and sorghum-sudangrass hybrid fell rapidly to below 5 within 24 hrs of ensiling, but it took 3 days for proso millet pH to decline below this value. During the late phase of ensiling, silage pH remained stable and was significantly lower in corn than in proso millet or sorghum-sudangrass hybrid ($p < 0.05$), possibly due to the higher population of LAB in corn silage biomass (Table 4). During the 45-day ensiling period, silage pH of corn, proso millet, and sorghum-sudangrass hybrid decreased by 1.94, 1.65, and 2.04 units, respectively. Buffering capacity, WSC

211 concentration, and moisture level have been identified as critical parameters influencing the
212 ensilability of forages if epiphytic LAB exist in sufficient numbers [30]. Buffering capacity was lowest
213 in corn (24.2 mEq/kg DM), intermediate in proso millet (32 mEq/kg DM), and highest in sorghum-
214 sudangrass hybrid (55.5 mEq/kg DM) [8]. Forages with higher buffering capacity require more acids
215 for pH reduction. This supports the faster pH reduction in corn plant at the initial phase of ensiling
216 than proso millet or sorghum-sudangrass hybrid.

217 Time-course of silage ammonia-nitrogen development, expressed as a proportion of total N is
218 illustrated in Figure 3. Initial $\text{NH}_3\text{-N}$ (g/kg total N) level before ensiling was highest in corn (35),
219 intermediate in proso millet (30), and lowest in sorghum-sudangrass hybrid (14.4). Ammonia-N
220 concentration increased in three forage crops as ensiling progressed, with proso millet exhibiting the
221 highest rise. This indicates that protein fractions in proso millet were degraded to a greater extent
222 during ensiling, perhaps because of accelerated rate of proteolysis and deamination [31]. The $\text{NH}_3\text{-N}$
223 concentration of less than 70 g/kg total N indicates successful silage fermentation, whereas amounts
224 greater than 100 g/kg total N have been linked to poor silage fermentation [32]. This criterion indicates
225 more degradation of protein in proso millet than corn and sorghum-sudangrass hybrid. The rapid
226 acidification of silage mass is known to inhibit growth and activity of undesirable microorganisms as
227 well as proteolytic activity [10, 33]. The higher $\text{NH}_3\text{-N}$ concentration in proso millet silage could be
228 attributed to its higher pH during ensiling, which was likely insufficient to effectively suppress
229 enzymes and microorganisms involved in protein degradation during fermentation.

230 Concentration of WSC in silage mass over the course of the 45-d fermentation is presented in Figure
231 4. Initial WSC concentration (before ensiling) was higher in proso millet than in corn or sorghum-
232 sudangrass hybrid (170 vs. mean 141 g/kg DM). An initial WSC concentration between 60 and 80 g/kg
233 DM has been suggested as an adequate amount to promote an efficient silage fermentation [34]. This
234 indicates that the forage crops evaluated in this study contained sufficient WSC to promote a good-
235 quality silage fermentation. The exhaustion of WSC was faster in corn plant as ensiling progressed,

236 reaching a minimum of 6.70 g/kg DM after 3 days of ensiling, after which WSC concentration
237 decreased slightly until day 45 of ensiling (5.20 g/kg DM). Proso millet experienced a comparatively
238 slower rate of decline in WSC during ensiling, decreasing to 18.2 g/kg DM on day 15 of ensiling and
239 reaching a mean value of 5.9 g/kg DM after 45 days of ensiling.

240 During the ensiling fermentation, LAB consume WSC as a readily available source of energy and
241 primarily convert it to lactic acid, which is associated with silage mass acidification and inhibition of
242 the activities of undesirable microorganisms [26]. Variations in WSC consumption rates amongst
243 forage crops during the early phase of ensiling might be ascribed to differences in microbial activity
244 and plant enzymes in the crops prior to ensiling. In general, WSC supplies the energy required to drive
245 silage fermentation [35]. A sufficient quantity of WSC has been identified as an important factor in
246 fast acidification during the initial phase of ensiling, which is associated with DM loss reduction and
247 improvement of silage quality [10]. In our experiment, the faster reduction of WSC in corn compared
248 to proso millet forage represented a faster decline in silage pH, which was associated with less DM
249 loss and $\text{NH}_3\text{-N}$ production during ensiling.

250 The IVDMD of the experimental forage crops as a function of ensiling duration are illustrated in
251 Figure 5. Before ensiling, IVDMD of proso millet and sorghum-sudangrass hybrid was not different,
252 averaging 643 g/kg DM, which was approximately 16% less than corn (746 g/kg DM). All crops
253 experienced a decline in IVDMD with ensiling. Previous studies have identified that ADF and NDF
254 concentrations correlate negatively with IVDMD [36]. This supports findings of the current study
255 because corn had less NDF and ADF fractions than proso millet or sorghum-sudangrass hybrid,
256 resulting in the higher digestibility of corn than the other two crops.

257 **Organic acids formation during ensiling**

258 Formation of lactic acid and acetic acid as a function of ensiling duration is illustrated in Table 3.
259 Butyric acid was undetectable during the 45-day ensiling period, which indicates a well-fermented

260 silage and a lack of clostridial activity during ensiling process [10, 26, 29]. High silage pH, typically
261 greater than 4.5, low DM concentration, and high buffering capacity have been identified as probable
262 factors which contribute to clostridia growth and proliferation during ensiling [32, 37]. This suggests
263 that among the forage types evaluated in this experiment, proso millet had a greater susceptibility to
264 clostridial activity and, thus butyric acid production. However, such an effect was not observed in this
265 experiment and the absence of butyric acid detection during silage fermentation of proso millet
266 indicates its low susceptibility to putrefaction by clostridial fermentation.

267 Lactic acid formation increased as silage fermentation progressed, and the magnitude of this
268 increase was generally greater in sorghum-sudangrass hybrid, intermediate in corn, and lowest in proso
269 millet. During the 45-day ensiling period, lactic acid concentration displayed an upward trend and
270 reached a maximum on day 45, with values of 42.5 g/kg DM for proso millet, 67.7 g/kg DM for corn,
271 and 127 g/kg DM for sorghum-sudangrass hybrid. Lactic acid is typically found in concentrations
272 ranging from 20 to 40 g/kg DM in commonly used silages [29], which indicates that all forages in the
273 present experiment underwent an adequate lactic acid fermentation. Similar to lactic acid production,
274 acetic acid also increased with ensiling, the rate of its production was generally larger in the earlier
275 phase of silage fermentation. During ensiling process, acetic acid was usually lower in proso millet
276 than in corn and sorghum-sudangrass hybrid. Acetic acid concentration in sorghum-sudangrass hybrid
277 reached a maximum concentration of 100 g/kg DM on day 45 of silage fermentation. The higher lactic
278 acid and acetic acid production in sorghum-sudangrass hybrid during silage fermentation could be
279 explained by its higher moisture concentration than the other two crops, which accelerates microbial
280 activity and acid production during the ensiling process. This explanation is supported by the findings
281 of a previous study identifying that a lower moisture level limits silage fermentation [38]. Although no
282 consistent trend was seen in lactic acid: acetic acid ratio, there was a general downward trend for each
283 crop, which is likely indicative of a shift from homo- to hetero-fermentative pattern. This observation
284 is consistent with results reported by Shao et al. [39, 40]. The higher ratio of lactic acid: acetic acid in

285 corn is most likely suggestive of the dominance of homofermentative LAB during the ensiling process.

286 **Microbial composition during ensiling**

287 Changes in microbial population as a function of ensiling duration are shown in Table 4. The pre-
288 ensiling population of LAB, mold, and total microorganisms is presented in our companion paper [8].
289 Briefly, the highest LAB count was detected in corn ($6.15 \log_{10}$ cfu/g), followed by proso millet (5.91
290 \log_{10} cfu/g), and sorghum-sudangrass hybrid ($5.88 \log_{10}$ cfu/g). An LAB count of $5.0 \log_{10}$ cfu/g
291 biomass has been suggested as a minimum number to enable the dominance of the epiphytic LAB
292 during ensiling [41, 42]. This suggests that the forage crops had sufficient epiphytic LAB population
293 to initiate an efficient silage fermentation. Number of mold was highest on proso millet biomass (4.53
294 \log_{10} cfu/g fresh mass), which was 0.23 and $1.23 \log_{10}$ cfu/g fresh mass greater than corn and sorghum-
295 sudangrass hybrid, respectively. Forage species, maturity stage, weather, and field wilting have all been
296 identified as factors causing differences in the population of epiphytic microorganisms in forage crops
297 [43]. During the 45-day of fermentation, LAB count was generally lower in proso millet than corn or
298 sorghum-sudangrass hybrid. Number of LAB increased during the early ensiling period and peaked on
299 day 10 of ensiling. Low pH and the exhaustion of fermentable substrates have been identified as the
300 primary factors contributing to the decline of LAB population as ensiling proceeds [44].

301 Mold was always present in each crop during fermentation, with a lower number existing on corn
302 than proso millet or sorghum-sudangrass hybrid. The lower mold population in corn biomass is likely
303 related to the rapid acidification (lower pH) of corn silage, inhibiting the growth of undesirable
304 microorganisms [26, 42]. Another factor inhibiting mold growth during ensiling is a high acetic acid
305 concentration [45]. Less formation of acetic acid and lactic acid (higher pH) during ensiling
306 fermentation of proso millet could possibly explain the higher mold number in proso millet biomass
307 during ensiling. The count of total microorganisms was generally higher in corn than in sorghum-
308 sudangrass hybrid or proso millet. Total microorganisms reached the maximum number on day 10 of

ensiling, and then followed a downward trend, which could be explained by pH reduction at this time point, limiting the growth of microorganisms.

CONCLUSION

Silage fermentation of proso millet forage resulted in a significant increase in ammonia nitrogen generation and a larger loss of dry matter when compared to corn or sorghum-sudangrass hybrid, perhaps because of its higher buffering capacity and silage pH. However, butyrate was undetectable during its ensiling fermentation. Further research is needed to optimize the fermentation quality of proso millet forage, possibly by using the appropriate silage additives to minimize ammonia-nitrogen formation during fermentation, as well as to promote greater lactate production, which is associated with a further decline in silage pH and mold growth inhibition, and thus with a reduction in dry matter loss. Despite the lower productivity (less forage production per unit of cultivated land) than corn and sorghum-sudangrass hybrid, nutrient value of proso millet was comparable to sorghum-sudangrass hybrid. Proso millet could be harvested in a shorter period of time, making it a potential summer crop in situations where cultivation of other major summer crops is limited.

CONFLICT OF INTEREST

We certify that there is no conflict of interest with any financial organization regarding the material discussed in the manuscript.

ACKNOWLEDGEMENTS

This research was supported by Cooperative Research Program for Agriculture Science & Technology Development (Project No. PJ01401903), Rural Development Administration, Republic of Korea.

330 **ORCID number**

- 331 Sheng Nan Wei (<https://orcid.org/0000-0001-5117-5140>)
332 Eun Chan Jeong (<https://orcid.org/0000-0002-6559-2743>)
333 Yan Fen Li (<https://orcid.org/0000-0002-7318-7318>)
334 Hak Jin Kim (<https://orcid.org/0000-0002-7279-9021>)
335 Farhad Ahmadi (<https://orcid.org/0000-0002-8760-053x>)
336 Jong Geun Kim (<https://orcid.org/0000-0003-4720-1849>)

337 **English check**

- 338 The English in this document has been checked by at least two professional editors, both native
339 speakers of English. For a certificate, please refer to:
340 <http://www.textcheck.com/certificate/vjIv2C>

341 **REFERENCES**

- 342 1. Korean Statistical Information Service. Farm households by size of raising Korean beef cattle/total
343 head [Internet]. 2017. Daejeon, Korea: Korean Statistical Information Service.
- 344 2. McCartney D, Fraser J, Ohama A. Potential of warm-season annual forages and Brassica crops for
345 grazing: A Canadian review. Canadian Journal of Animal Science. 2009;89(4):431-40.
346 <https://doi.org/10.4141/CJAS09002>
- 347 3. Habiyaemye C, Matanguihan JB, D'Alpoim Guedes J, Ganjyal GM, Whiteman MR, Kidwell KK,
348 et al. Proso millet (*Panicum miliaceum* L.) and its potential for cultivation in the Pacific Northwest,
349 US: A review. Frontiers in Plant Science. 2017;7:1961. <https://doi.org/10.3389/fpls.2016.01961>
- 350 4. Calamai A, Masoni A, Marini L, Dell'acqua M, Ganugi P, Boukail S, et al. Evaluation of the
351 agronomic traits of 80 accessions of proso millet (*Panicum miliaceum* L.) under Mediterranean
352 pedoclimatic conditions. Agriculture. 2020;10(12):578. <https://doi.org/10.3390/agriculture10120578>
353
- 354 5. Amadou I, Gounga ME, Le G-W. Millets: Nutritional composition, some health benefits and
355 processing – A review. Emirates Journal of Food and Agriculture. 2013;501-8.
356 <https://doi.org/10.9755/ejfa.v25i7.12045>
- 357 6. Nematpour A, Eshghizadeh HR, Zahedi M. Comparing the corn, millet and sorghum as silage

- 358 crops under different irrigation regime and nitrogen fertilizer levels. International Journal of Plant
359 Production. 2021;31:351-361. <https://doi.org/10.1007/s42106-021-00142-8>
- 360 7. Lyon DJ, Burgener, PA, DeBoer KL, Harveson RM, Hein GL, Hergert GW, Holman TL, Nelson
361 LA, Johnson JJ, Nleya T. and Krall JM. Producing and marketing proso millet in the Great Plains.
362 Lincoln, NE: University of Nebraska Extension Circular #EC 137.2008.rs-739296/v1
- 363 8. Wei SN, Li YF, Jeong EC, Kim HJ, Kim JG. Effects of formic acid and lactic acid bacteria
364 inoculant on main summer crop silages in Korea. Journal of Animal Science and Technology.
365 2021;63(1):91. <https://doi.org/10.5187/jast.2021.e7>
- 366 9. Wang M, Franco M, Cai Y, Yu Z. Dynamics of fermentation profile and bacterial community of
367 silage prepared with alfalfa, whole-plant corn and their mixture. Animal Feed Science and
368 Technology. 2020;270:114702. <https://doi.org/10.1016/j.anifeedsci.2020.114702>
- 369 10. Weinberg ZG, Muck R. New trends and opportunities in the development and use of inoculants
370 for silage. FEMS Microbiology Reviews. 1996;19(1):53-68. <https://doi.org/10.1111/j.1574-6976.1996.tb00253.x>
371
- 372 11. Wiersma DW. Kernel milkline stage and corn forage yield, quality, and dry matter content. Journal
373 of Production Agriculture. 1993;6:94-99. <https://doi.org/10.2134/jpa1993.0094>
- 374 12. Lynch JP, O’Kiely P, Doyle EM. Yield, quality and ensilage characteristics of whole-crop maize
375 and of the cob and stover components: harvest date and hybrid effects. Grass and Forage Science.
376 2012;67(4):472-487. <https://doi.org/10.1111/j.1365-2494.2012.00868.x>
- 377 13. Ahmadi F, Lee YH, Lee WH, Oh YK, Park K, Kwak WS. Long-term anaerobic conservation of
378 fruit and vegetable discards without or with moisture adjustment after aerobic preservation with
379 sodium metabisulfite. Waste Management. 87;2019:258-267.
380 <https://doi.org/10.1016/j.wasman.2019.02.010>
- 381 14. Broderick G, Kang J. Automated simultaneous determination of ammonia and total amino acids
382 in ruminal fluid and *in vitro* media. Journal of Dairy Science. 1980;63(1):64-75.
383 [https://doi.org/10.3168/jds.S0022-0302\(80\)82888-8](https://doi.org/10.3168/jds.S0022-0302(80)82888-8)
- 384 15. Brock TD, Madigan MT, Martinko JM, Parker J. Brock Biology of Microorganisms: Upper Saddle
385 River (NJ): Prentice-Hall, 2003.
- 386 16. Jean-Baptiste. Jean-Baptiste - André Dumas. Science 1884; 3(72): 750-752.
387 <https://doi.org/10.1126/science.ns-3.72.750>
- 388 17. Van Soest PJ, Robertson J, Lewis B. Methods for dietary fiber, neutral detergent fiber, and
389 nonstarch polysaccharides in relation to animal nutrition. Journal of Dairy Science.
390 1991;74(10):3583-97. [https://doi.org/10.3168/jds.S0022-0302\(91\)78551-2](https://doi.org/10.3168/jds.S0022-0302(91)78551-2)

- 391 18. Yemm E, Willis A. The estimation of carbohydrates in plant extracts by anthrone. *Biochemical*
392 *Journal*. 1954;57(3):508-14. <https://doi.org/10.1042/bj0570508>
- 393 19. ANKOM Technology. 2017. Method 3: In Vitro True Digestibility using the DAISY^{II} Incubator.
394 ANKOM Technology, Macedon, NY. Retrieved from
395 http://www.ankom.com/media/documents/IVDMD_0805_D200.pdf (accessed 10 May. 2021).
- 396 20. Goering HK, Van Soest PJ. Forage Fiber Analysis (Apparatus, Reagents, Procedures, and Some
397 Applications). Agricultural Handbook no. 379. US Department Agriculture-Agricultural Research
398 Service (USDA-ARS), Washington, DC. 1970.
- 399 21. Horrocks RD, Valentine JF. Harvested Forages. Academic Press, San Diego, CA, USA. 1999.
- 400 22. Jahansouza MR, Keshavarz Afshar, R., Heidari Zooleh, H., Hashemi, M. Evaluation yield and
401 quality of sorghum and millet as alternative forage crops to corn under normal and deficit irrigation
402 systems. *Jordan Journal of Agricultural Sciences*. 2014(10):699-714.
- 403 23. Kim JG, Jeong EC, Kim MJ, Li YF, Kim HJ, Lee SH. Comparison of growth characteristics and
404 productivity of summer forage crops in Sihwa reclaimed land. *Journal of The Korean Society of*
405 *Grassland and Forage Science*. 2021;41(2):110-8. <https://doi.org/10.5333/KGFS.2021.41.2.110>
- 406 24. Shin J-S, Kim W-H, Lee S-H, Shin H-Y. Comparison of forage yield and feed value of millet
407 varieties in the reclaimed tidelands. *Journal of The Korean Society of Grassland and Forage*
408 *Science*. 2006;26(4):215-20. <https://doi.org/10.5333/KGFS.2006.26.4.215>
- 409 25. Kim J, Lee Y, Kim Y, Ahmadi F, Oh Y, Park J, et al. Effect of microbial inoculant or molasses on
410 fermentative quality and aerobic stability of sawdust-based spent mushroom substrate.
411 *Bioresource Technology*. 2016;216:188-95. <https://doi.org/10.1016/j.biortech.2016.05.056>
- 412 26. Chen L, Yuan X, Li J, Dong Z, Wang S, Guo G, et al. Effects of applying lactic acid bacteria and
413 propionic acid on fermentation quality, aerobic stability and *in vitro* gas production of forage-
414 based total mixed ration silage in Tibet. *Animal Production Science*. 2019;59(2):376-83.
415 <https://doi.org/10.1071/AN16062>
- 416 27. McDonald P, Henderson A, Heron SJE. The Biochemistry of Silage: Chalcombe publications;
417 1991.
- 418 28. Diepersloot EC, Pupo MR, Ghizzi LG, Gusmão JO, Heinzen Jr C, McCary CL, et al. Effects of
419 microbial inoculation and storage length on fermentation profile and nutrient composition of
420 whole-plant sorghum silage of different varieties. *Frontiers in Microbiology*. 2021;12.
421 <https://doi.org/10.3389/fmicb.2021.660567>
- 422 29. Kung Jr L, Shaver R, Grant R, Schmidt R. Silage review: Interpretation of chemical, microbial,
423 and organoleptic components of silages. *Journal of Dairy Science*. 2018;101(5):4020-33.
424 <https://doi.org/10.3168/jds.2017-13909>

- 425 30. Fernández AM, Cabezuelo ABS, de la Roza Delgado MB, Arrojo MG, Gutiérrez AA. Modelling
426 a quantitative ensilability index adapted to forages from wet temperate areas. Spanish Journal of
427 Agricultural Research. 2013(2):455-62. <https://doi.org/10.5424/sjar/2013112-3219>
- 428 31. Arriola K, Queiroz O, Romero J, Casper D, Muniz E, Hamie J, et al. Effect of microbial inoculants
429 on the quality and aerobic stability of bermudagrass round-bale haylage. Journal of Dairy Science.
430 2015;98(1):478-85. <https://doi.org/10.3168/jds.2014-8411>
- 431 32. Lima R, Lourenço M, Díaz RF, Castro A, Fievez V. Effect of combined ensiling of sorghum and
432 soybean with or without molasses and lactobacilli on silage quality and in vitro rumen
433 fermentation. Animal Feed Science and Technology. 2010;155(2-4):122-31.
434 <https://doi.org/10.1016/j.anifeedsci>.
- 435 33. Pahlow G, Muck RE, Driehuis F, Elferink SJO, Spoelstra SF. Microbiology of ensiling. Silage
436 Science and Technology. 2003;42:31-93. <https://doi.org/10.2134/agronmonogr42.c2>
- 437 34. Amer S, Hassanat F, Berthiaume R, Seguin P, Mustafa A. Effects of water-soluble carbohydrate
438 content on ensiling characteristics, chemical composition and in vitro gas production of forage
439 millet and forage sorghum silages. Animal Feed Science and Technology. 2012;177(1-2):23-9.
440 <https://doi.org/10.1016/j.anifeedsci.2012.07.024>
- 441 35. Contreras-Govea FE, Muck RE, Broderick GA, Weimer PJ. Lactobacillus plantarum effects on
442 silage fermentation and *in vitro* microbial yield. Animal Feed Science and Technology.
443 2013;179(1):61-8. <https://doi.org/10.1016/j.anifeedsci.2012.11.008>
- 444 36. Ammar H, López S, González JS, Ranilla MJ. Chemical composition and in vitro digestibility of
445 some Spanish browse plant species. Journal of the Science of Food and Agriculture.
446 2004;84(2):197-204. <https://doi.org/10.1002/jsfa.1635>
- 447 37. Queiroz O, Ogunade I, Weinberg Z, Adesogan A. Silage review: Foodborne pathogens in silage
448 and their mitigation by silage additives. Journal of Dairy Science. 2018;101(5):4132-42.
449 <https://doi.org/10.3168/jds.2017-13901>
- 450 38. Kim J, Chung E, Seo S, Ham J, Kang W, Kim D. Effects of maturity at harvest and wilting days
451 on quality of round baled rye silage. Asian-Australasian Journal of Animal Sciences.
452 2001;14(9):1233-7. <https://doi.org/10.5713/ajas.2001.1233>
- 453 39. Shao T, Ohba N, Shimojo M, Masuda Y. Dynamics of early fermentation of Italian ryegrass
454 (*Lolium multiflorum* Lam.) silage. Asian-Australasian Journal of Animal Sciences.
455 2002;15(11):1606-10. <https://doi.org/10.5713/ajas.2002.1606>
- 456 40. Shao T, Zhang Z, Shimojo M, Wang T, Masuda Y. Comparison of fermentation characteristics of
457 Italian ryegrass (*Lolium multiflorum* Lam.) and guineagrass (*Panicum maximum* Jacq.) during the
458 early stage of ensiling. Asian-Australasian Journal of Animal Sciences. 2005;18(12):1727-34.
459 <https://doi.org/10.5713/ajas.2005.1727>

- 460 41. Cai Y, Benno Y, Ogawa M, Kumai S. Effect of applying lactic acid bacteria isolated from forage
461 crops on fermentation characteristics and aerobic deterioration of silage. *Journal of Dairy Science*.
462 1999;82(3):520-6. [https://doi.org/10.3168/jds.S0022-0302\(99\)75263-X](https://doi.org/10.3168/jds.S0022-0302(99)75263-X)
- 463 42. Muck R, Kung L. Effects of silage additives on ensiling. *Proceedings from the Silage: Field to*
464 *Feedbunk, North American Conference, Ithaca, NY, Northeast Regional Agricultural Engineering*
465 *Service Publication; 1997.*
- 466 43. Fenton MP. An investigation into the sources of lactic acid bacteria in grass silage. *Journal of*
467 *Applied Bacteriology*. 1987;62(3):181-8. <https://doi.org/10.1111/j.1365-2672.1987.tb02397.x>
- 468 44. Xu Z, He H, Zhang S, Kong J. Effects of inoculants *Lactobacillus brevis* and *Lactobacillus*
469 *parafarraginis* on the fermentation characteristics and microbial communities of corn stover silage.
470 *Scientific Reports*. 2017;7(1):1-9. <https://doi.org/10.1038/s41598-017-14052-1>
- 471 45. Danner H, Holzer M, Mayrhuber E, Braun R. Acetic acid increases stability of silage under aerobic
472 conditions. *Applied and Environmental Microbiology*. 2003;69(1):562-7.
473 <https://doi.org/10.1128/AEM.69.1.562-567.2003>
- 474 46. Schroeder, J.W. *Forage Nutrition for Ruminants*; NDSU Extension Service, North Dakota State
475 University: Fargo, ND, USA, 2004.
- 476 47. Rohweder D, Barnes R, Jorgensen N. Proposed hay grading standards based on laboratory
477 analyses for evaluating quality. *Journal of Animal Science*. 1978;47(3):747-59.
478 <https://doi.org/10.2527/jas1978.473747x>

479 **Table 1.** Forage yield and forage quality of proso millet, corn, and sorghum-sudangrass hybrid.

Items	Forage type			SEM	<i>p</i> value
	Proso millet	Corn	Sorghum-sudangrass hybrid		
Dry matter, g/kg	303 ^a	277 ^b	193 ^c	11.7	<0.01
TDN, g/kg DM	631 ^b	677 ^a	541 ^c	16.2	<0.01
RFV	97 ^b	117 ^a	77 ^c	7.63	<0.01
Yield, tons/ha					<0.01
Fresh matter	25.4 ^c	67.6 ^b	121.7 ^a	8.97	<0.01
Dry matter	7.69 ^c	18.7 ^b	23.5 ^a	1.41	<0.01

480 ^{a-c} means with different letter within each row differ ($p < 0.05$). SEM = standard error of mean.

481 TDN = total digestible nutrients. For proso millet and sorghum-sudangrass hybrid, TDN was calculated
 482 according to the following equation: $[889 - (0.79 \times \text{ADF, g/kg DM})]$. For corn plant, TDN was calculated using
 483 the following equation: $[878.4 - (0.70 \times \text{ADF, g/kg DM})]$ [46].

484 RFV = relative feed value calculated according to the following equation: $[(\text{dry matter intake} \times \text{digestible dry}$
 485 $\text{matter})/1.29]$, where dry matter intake = $120/(\text{NDF}\%)$ and digestible dry matter = $88.9 - (0.779 \times \text{ADF}\%)$ [47].
 486

487 **Table 2.** Dry matter (DM) concentration, DM loss and chemical composition during ensiling. Values were expressed as g/kg DM, unless otherwise
488 stated.

Items	Forage type	Ensiling days									SEM
		1	2	3	5	10	15	20	30	45	
DM, g/kg	Proso millet	284.4 ^{aA}	284.1 ^{aA}	278.6 ^{abA}	278.9 ^{abA}	272.2 ^{bcA}	270.2 ^{cA}	269.6 ^{cA}	266.8 ^{cA}	266.4 ^{cA}	3.22
	Corn	275.2 ^{aB}	274.6 ^{aB}	274.7 ^{aA}	272.5 ^{aA}	272.1 ^{aA}	271.0 ^{aA}	268.4 ^{abA}	264.1 ^{bcA}	260.7 ^{cA}	2.82
	Sorghum-sudangrass hybrid	190.3 ^{aC}	187.5 ^{abC}	183.1 ^{abcB}	180.0 ^{bcB}	174.3 ^{cB}	177.7 ^{cB}	176.7 ^{cB}	178.4 ^{bcB}	174.7 ^{cB}	3.82
DM loss	Proso millet	19.0 ^{cA}	19.30 ^{cA}	24.8 ^{bcA}	24.50 ^{bcA}	31.2 ^{abA}	33.20 ^{aA}	33.8 ^{aA}	36.6 ^{aA}	37.0 ^{aA}	2.95
	Corn	2.14 ^{dB}	2.73 ^{dB}	2.63 ^{dB}	4.78 ^{cdC}	5.20 ^{cdC}	6.28 ^{cdC}	8.94 ^{bcC}	13.2 ^{abB}	16.6 ^{aB}	2.11
	Sorghum-sudangrass hybrid	2.50 ^{dB}	5.30 ^{dcB}	9.70 ^{bcC}	12.8 ^{abB}	18.5 ^{aB}	15.1 ^{aB}	16.1 ^{aB}	15.6 ^{aB}	18.1 ^{aB}	2.23
Crude protein	Proso millet	62.3 ^{aA}	61.0 ^{abA}	59.8 ^{abA}	58.3 ^{abA}	60.3 ^{abA}	59.9 ^{abA}	57.9 ^{bA}	59.6 ^{abA}	57.1 ^{bA}	1.46
	Corn	57.4 ^{aB}	56.5 ^{aB}	58.4 ^{aA}	54.6 ^{abB}	54.5 ^{abB}	53.40 ^{abB}	52.7 ^{abB}	53.0 ^{abB}	50.8 ^{bbB}	2.12
	Sorghum-sudangrass hybrid	53.4 ^{aC}	48.9 ^{abC}	49.5 ^{abB}	49.6 ^{abBC}	48.4 ^{abC}	46.2 ^{bC}	46.6 ^{bC}	46.6 ^{bC}	46.2 ^{bC}	1.99
ADF	Proso millet	324.9 ^{bB}	327.3 ^{bbB}	324.3 ^{bB}	342.4 ^{aB}	344.4 ^{aB}	340.1 ^{abB}	347.1 ^{aB}	330.1 ^{bB}	345.8 ^{aB}	4.76
	Corn	260.5 ^{aC}	256.2 ^{abC}	251.5 ^{bcC}	243.4 ^{dcC}	248.9 ^{bcC}	249.2 ^{bcC}	246.1 ^{cdC}	241.5 ^{dcC}	252.0 ^{bcC}	3.21
	Sorghum-sudangrass hybrid	419.1 ^A	419.7 ^A	420.4 ^A	414.9 ^A	420.5 ^A	422.1 ^A	427.6 ^A	413.2 ^A	415.1 ^A	4.98
NDF	Proso millet	608.5 ^{aB}	610.8 ^{aB}	604.3 ^{aB}	606.6 ^{aB}	601.8 ^{abB}	610.5 ^{aB}	602.5 ^{abB}	586.0 ^{bB}	590.5 ^{bbB}	6.01
	Corn	496.1 ^{aC}	491.0 ^{aC}	467.6 ^{bcC}	445.3 ^{cC}	454.4 ^{cC}	455.8 ^{cC}	455.6 ^{cC}	445.5 ^{cC}	449.2 ^{cC}	4.37
	Sorghum-sudangrass hybrid	674.7 ^{aA}	673.1 ^{aA}	668.0 ^{aA}	665.0 ^{aA}	666.2 ^{aA}	669.8 ^{aA}	671.7 ^{aA}	635.0 ^{bA}	640.1 ^{bA}	5.45

489 ADF: acid detergent fiber, NDF: neutral detergent fiber. Values with different lowercase letters within each row show significant difference among ensiling days
490 with the same forage type. Values with different capital letters within each column show significant differences among forage types in the same ensiling day (p
491 < 0.05).

492 SEM = standard error of mean.

493 **Table 3.** Concentrations of lactic acid and acetic acid as a function of ensiling days.

Organic acids	Forage type	Ensiling days									SEM
		1	2	3	5	10	15	20	30	45	
Lactic acid (LA), g/kg DM	Proso millet	10.1 ^{eB}	14.1 ^{deB}	23.2 ^{cB}	21.6 ^{cdB}	29.6 ^{bcC}	21.8 ^{cdC}	40.0 ^{aB}	37.0 ^{abC}	42.5 ^{aC}	3.64
	Corn	17.6 ^{eB}	28.6 ^{dA}	33.6 ^{dAB}	42.9 ^{cA}	44.3 ^{cB}	48.0 ^{cB}	57.5 ^{bA}	62.0 ^{abB}	66.7 ^{aB}	2.98
	Sorghum-sudangrass hybrid	31.3 ^{eA}	36.1 ^{deA}	39.7 ^{deA}	45.2 ^{dA}	67.9 ^{cA}	71.0 ^{cA}	69.3 ^{cA}	98.4 ^{bA}	126.6 ^{aA}	5.49
Acetic acid (AA), g/kg DM	Proso millet	5.67 ^{eC}	11.1 ^{deB}	14.7 ^{dC}	14.1 ^{dC}	26.3 ^{cC}	25.1 ^{cC}	62.7 ^{aA}	49.0 ^{bB}	41.7 ^{bB}	3.37
	Corn	10.3 ^{dB}	14.8 ^{dB}	22.6 ^{cB}	27.1 ^{cB}	37.6 ^{bB}	34.5 ^{bB}	26.0 ^{cB}	48.4 ^{aB}	38.2 ^{bB}	2.24
	Sorghum-sudangrass hybrid	15.9 ^{eA}	57.6 ^{cdA}	49.6 ^{dA}	63.3 ^{cA}	54.8 ^{cdA}	77.5 ^{bA}	61.6 ^{cA}	83.2 ^{bA}	100.3 ^{aA}	4.63
LA/AA	Proso millet	1.78 ^a	1.27 ^{bB}	1.58 ^{abA}	1.53 ^{abA}	1.13 ^b	0.87 ^{cdB}	0.64 ^{dC}	0.76 ^{dB}	1.02 ^{bcB}	0.15
	Corn	1.71 ^{bc}	1.93 ^{abA}	1.48 ^{cdA}	1.58 ^{bcdA}	1.18 ^d	1.39 ^{cdA}	2.21 ^{aA}	1.28 ^{dA}	1.75 ^{bcA}	0.21
	Sorghum-sudangrass hybrid	1.97 ^a	0.62 ^{bC}	0.80 ^{bbB}	0.71 ^{bbB}	1.24 ^{ab}	0.92 ^{bbB}	1.13 ^{abB}	1.18 ^{abA}	1.26 ^{abB}	0.45

494 Values with different lowercase letters within each row show significant difference among ensiling days with the same forage type. Values with different capital
495 letters within each column show significant differences among forage types in the same ensiling day ($p < 0.05$).

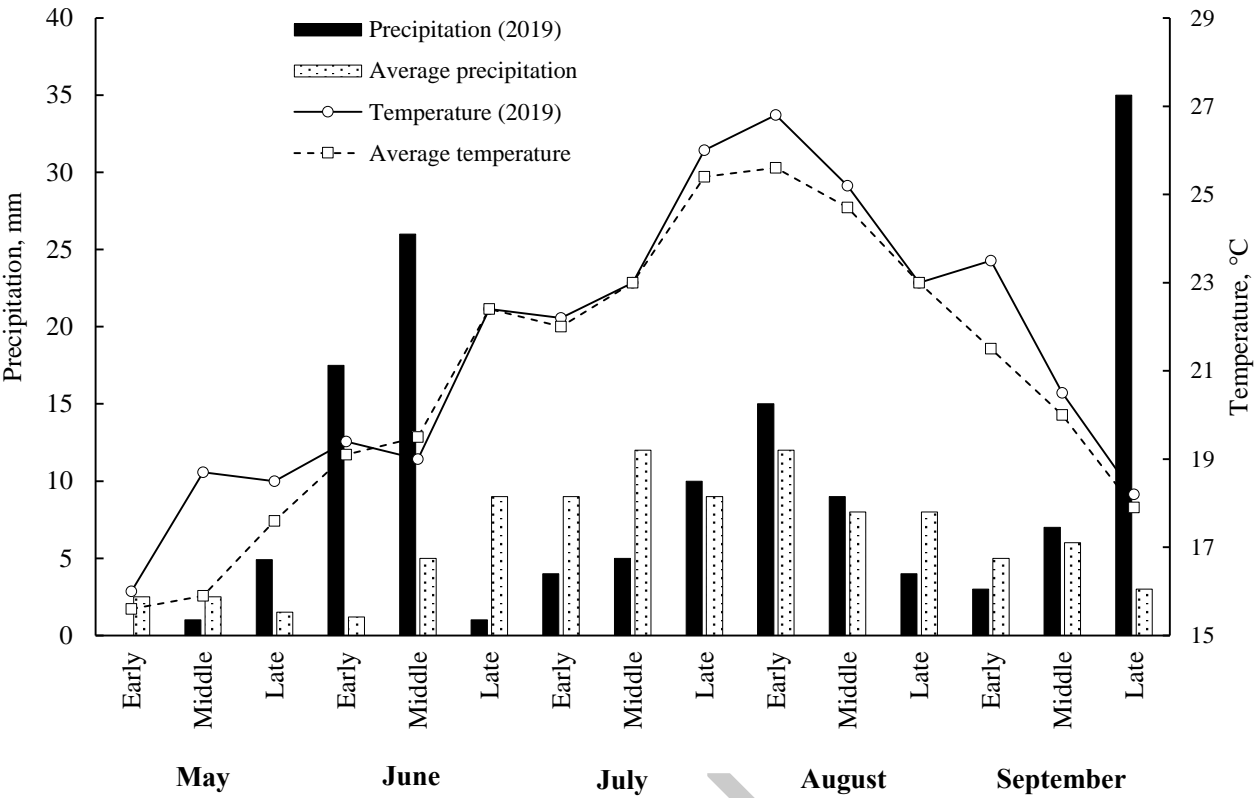
496 SEM = standard error of mean.

498 **Table 4.** Number of lactic acid bacteria, mold and total microorganisms as a function of ensiling days.

Microbial count ¹	Forage type	Ensiling days									SEM
		1	2	3	5	10	15	20	30	45	
Lactic acid bacteria	Proso millet	6.48 ^{bb}	6.88 ^{aA}	6.94 ^{aB}	6.93 ^{aB}	6.96 ^{aB}	6.48 ^{bb}	5.78 ^{cC}	5.78 ^{cB}	5.34 ^{dB}	0.11
	Corn	6.84 ^{fA}	6.85 ^{fA}	7.23 ^{cdA}	7.30 ^{cA}	7.77 ^{aA}	7.61 ^{abA}	7.04 ^{eA}	6.60 ^{gA}	6.08 ^{hA}	0.08
	Sorghum-sudangrass hybrid	5.08 ^{dC}	5.68 ^{cB}	6.60 ^{bC}	6.53 ^{bC}	6.95 ^{aB}	6.60 ^{bB}	6.95 ^{aB}	6.62 ^{bA}	5.89 ^{cA}	0.12
Molds	Proso millet	3.49 ^{dA}	4.21 ^{cA}	4.30 ^{bcA}	4.20 ^{cA}	5.00 ^{abA}	5.38 ^{aA}	4.34 ^{bcA}	4.04 ^{cB}	4.80 ^{bA}	0.30
	Corn	3.18 ^{cB}	3.00 ^{cC}	4.00 ^{bB}	3.00 ^{cC}	4.05 ^{bb}	4.00 ^{bC}	3.85 ^{bb}	4.67 ^{aA}	4.18 ^{abB}	0.41
	Sorghum-sudangrass hybrid	3.48 ^{cdA}	3.30 ^{dB}	3.85 ^{bB}	3.70 ^{bcB}	3.29 ^{dC}	5.11 ^{aB}	3.60 ^{bcC}	3.31 ^{dC}	3.00 ^{cC}	0.13
Total microorganisms	Proso millet	7.43 ^{bA}	7.51 ^{bA}	7.44 ^{bA}	7.79 ^{aB}	7.86 ^{aB}	7.26 ^{bcA}	7.04 ^{cB}	6.30 ^{dC}	6.60 ^{eB}	0.10
	Corn	7.05 ^{cB}	7.29 ^{cA}	7.18 ^{cB}	8.10 ^{bA}	8.85 ^{aA}	7.11 ^{cAB}	7.85 ^{bA}	7.04 ^{cB}	7.12 ^{cA}	0.19
	Sorghum-sudangrass hybrid	6.57 ^{cC}	6.88 ^{bb}	6.48 ^{cC}	7.01 ^{bC}	7.40 ^{aC}	6.95 ^{bb}	7.04 ^{bb}	7.32 ^{aA}	6.51 ^{cB}	0.08

499 ¹ Microbial count was expressed as the logarithmic number of colony-forming units per gram fresh mass. Values with different lowercase letters within each
500 row show significant difference among ensiling days with the same forage type. Values with different capital letters within each column show significant
501 differences among forage types in the same ensiling day ($p < 0.05$).

502 SEM = standard error of mean.



504

505

506 **Figure 1.** Temperature and precipitation during the growing season (May to September, 2019) and
507 comparison with the average climatic normal. The data were obtained from the Korean Meteorological
508 Administration.

509

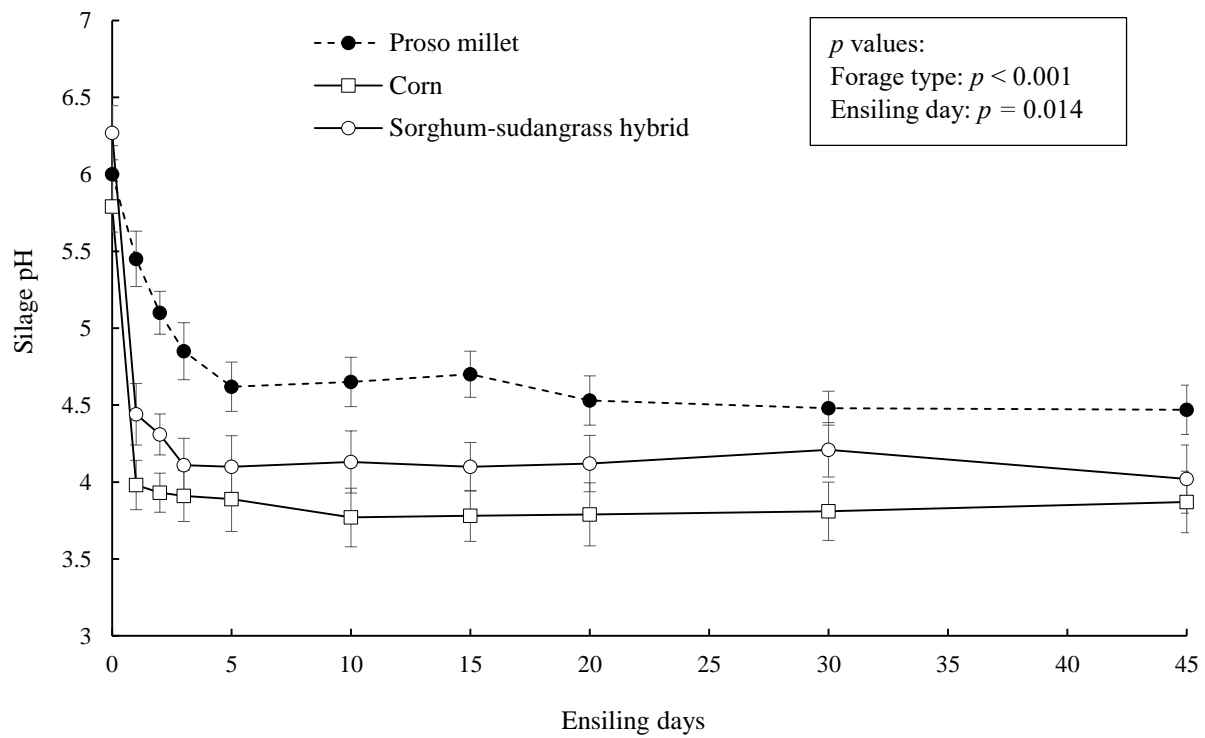
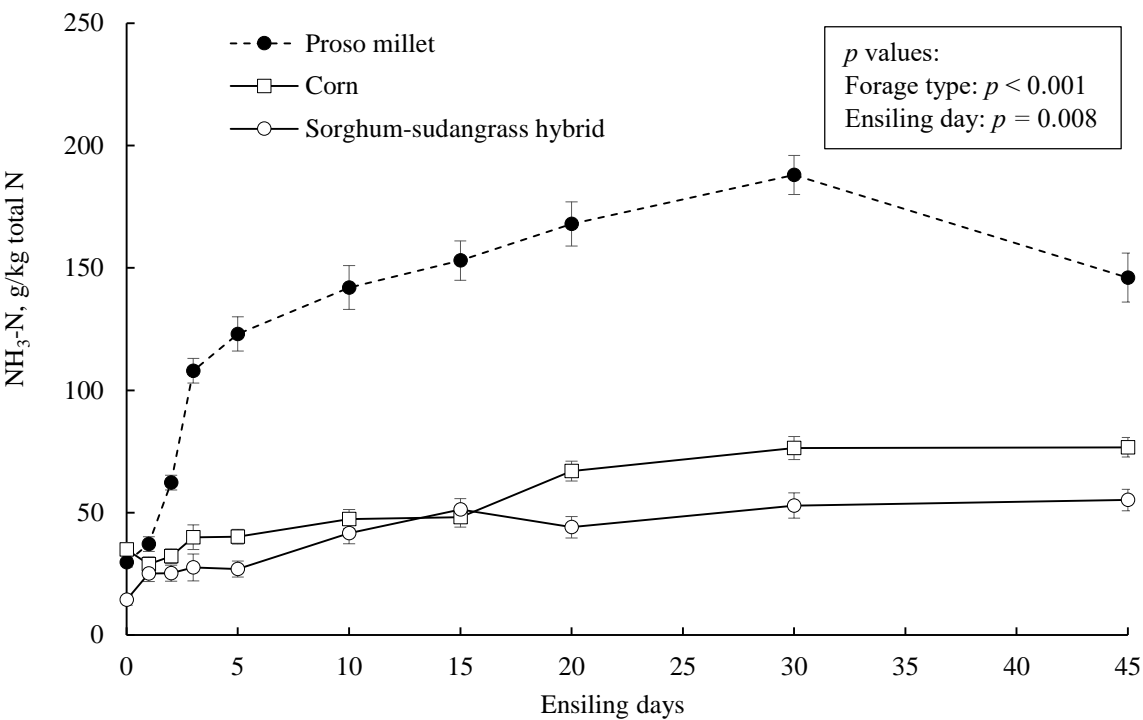


Figure 2. The pH value of proso millet, corn, and sorghum-sudangrass hybrid as a function of ensiling days. Bars indicate standard error.



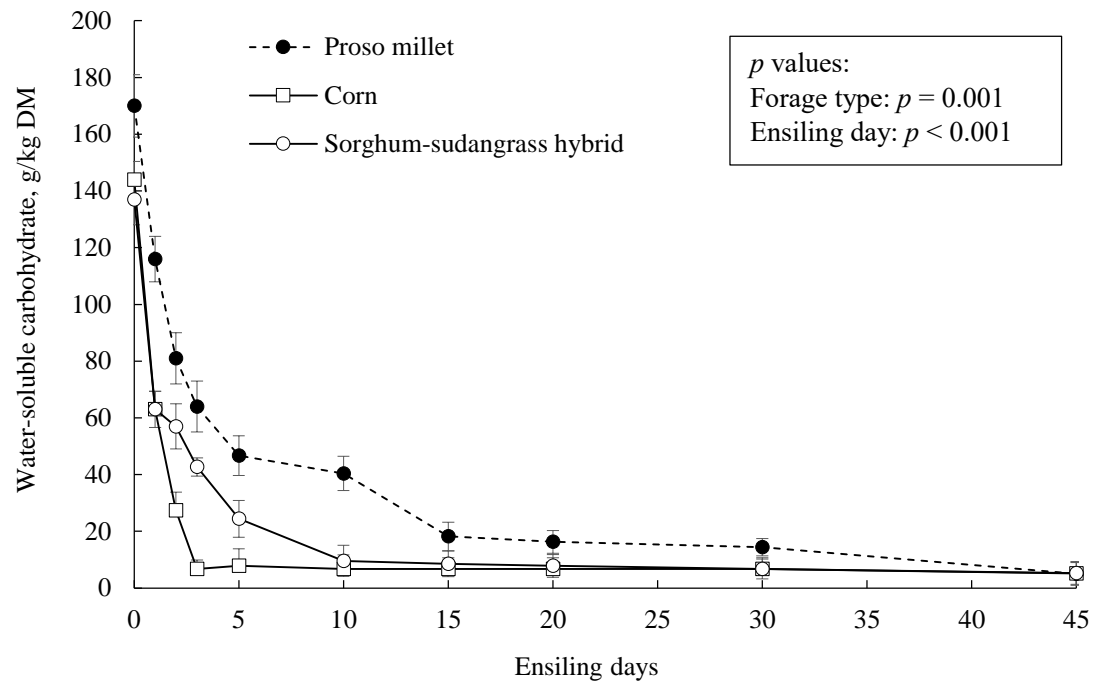
515

516

517 **Figure 3.** Ammonia-nitrogen concentration of proso millet, corn, and sorghum-sudangrass hybrid as a
518 function of ensiling days. Bars indicate standard error.

519

520

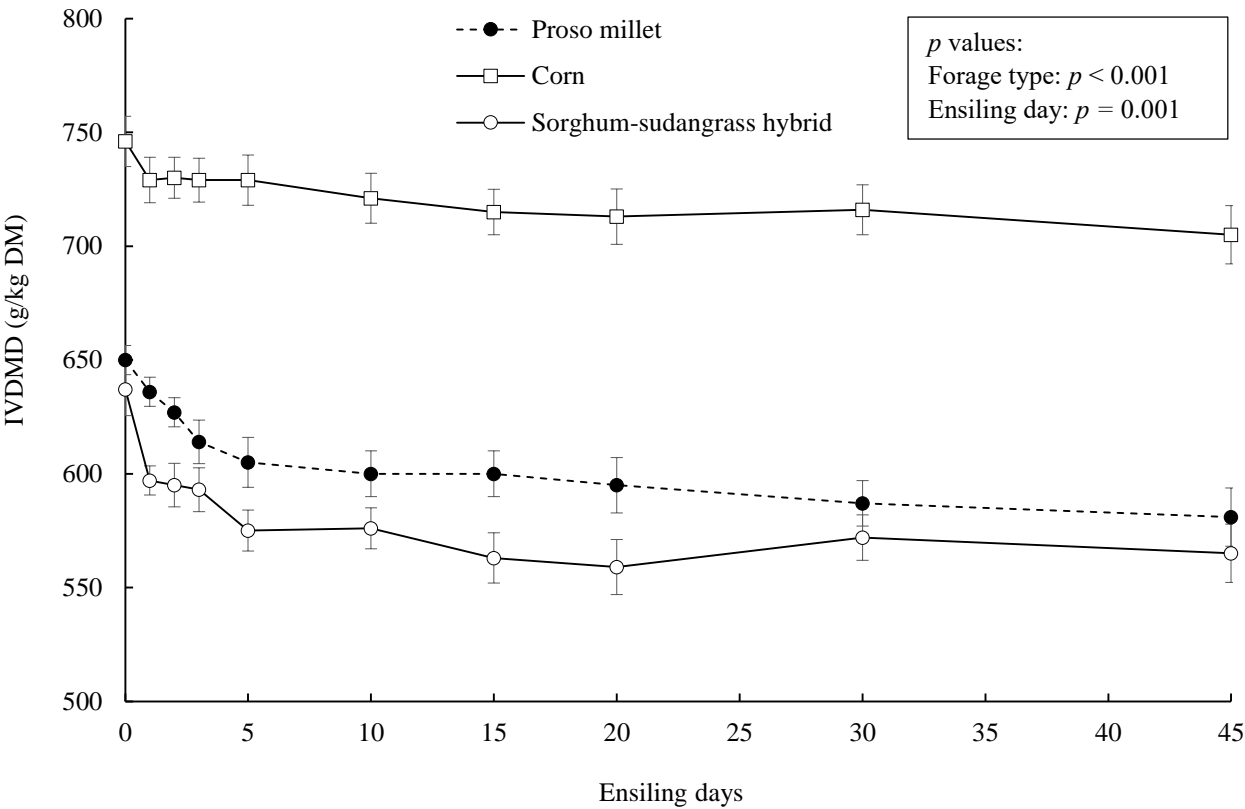


521

522 **Figure 4.** Water-soluble carbohydrate concentration of proso millet, corn, and sorghum-sudangrass
523 hybrid as a function of ensiling days. Bars indicate standard error.

524

525



526

527

528 **Figure 5.** *In vitro* dry matter digestibility (IVDMD) of proso millet, corn, and sorghum-sudangrass
529 hybrid as a function of ensiling days. Bars indicate standard error.