

JAST (Journal of Animal Science and Technology) TITLE PAGE
Upload this completed form to website with submission

| ARTICLE INFORMATION | Fill in information in each box below |
|--|---|
| Article Type | Research article |
| Article Title (within 20 words without abbreviations) | Dietary spray-dried plasma supplementation in late-gestation and lactation enhanced productive performance and immune responses of lactating sows and their litters |
| Running Title (within 10 words) | Dietary spray-dried plasma for productivity of lactating sows and piglets |
| Author | Kwangwook Kim ^{1#} , Byeonghyeon Kim ^{2#} , Hyunjin Kyoung ^{3#} , Yanhong Liu ¹ , Joy M. Campbell ⁴ , Minhong Song ^{3*} and Peng Ji ^{5*} |
| Affiliation | ¹ Department of Animal Science, University of California, Davis, CA 95616, United States of America ² Animal Nutrition & Physiology Team, National Institute of Animal Science, Rural Development Administration, Wanju 55365, Korea ³ Division of Animal and Dairy Science, Chungnam National University, Daejeon 34134, Korea ⁴ APC, Inc., Ankeny, IA 50021, United States of America ⁵ Department of Nutrition, University of California, Davis, CA 95616, United States of America |
| ORCID (for more information, please visit https://orcid.org) | Kwangwook Kim (https://orcid.org/0000-0001-5854-6047) Byeonghyeon Kim (https://orcid.org/0000-0003-4651-6857) Hyunjin Kyoung (https://orcid.org/0000-0001-5742-5374) Yanhong Liu (https://orcid.org/0000-0001-7727-4796) Joy M. Campbell (https://orcid.org/0000-0003-2499-3644) Minhong Song (https://orcid.org/0000-0002-4515-5212) Peng Ji (https://orcid.org/0000-0002-7447-5688) |
| Competing interests | No potential conflict of interest relevant to this article was reported. |
| Funding sources State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available. | This work was supported by Korea Institute of Planning and Evaluation for Technology in Food, Agriculture and Forestry (IPET) through Useful Agricultural Life Resources Industry Technology Development Program, funded by Ministry of Agriculture, Food and Rural Affairs (MAFRA)(120051-02-2-HD020). |
| Acknowledgements | Not applicable |
| Availability of data and material | Upon reasonable request, the datasets of this study can be available from the corresponding author. |
| Authors' contributions Please specify the authors' role using this form. | Conceptualization: Kim K, Kim B, Kyoung H, Song M, Ji P. Data curation: Kim K, Kim B, Kyoung H, Liu Y, Campbell JM, Song M, Ji P. Formal analysis: Kim K, Kim B. Methodology: Kim K, Kim B, Kyoung H. Software: Kim K, Kim B, Kyoung H. Validation: Liu Y, Campbell JM, Song M, Ji P. Investigation: Kim K, Kim B, Kyoung H. Writing - original draft: Kim K, Kim B, Kyoung H. Writing - review & editing: Kim K, Kim B, Kyoung H, Liu Y, Campbell JM, Song M, Ji P. |
| Ethics approval and consent to participate | This experimental protocol for this research was reviewed and approved by the Institutional Animal Care and Use Committee of Chungnam National University, Daejeon, Korea (approval # CNU-00611). |

CORRESPONDING AUTHOR CONTACT INFORMATION

| For the corresponding author (responsible for correspondence, proofreading, and reprints) | Fill in information in each box below |
|--|--|
| First name, middle initial, last name | Minho Song |
| Email address – this is where your proofs will be sent | mhsong@cnu.ac.kr |
| Secondary Email address | mhsong6@gmail.com |
| Address | Division of Animal and Dairy Science, Chungnam National University, Daejeon 34134, Republic of Korea |
| Cell phone number | +82-10-9254-0931 |
| Office phone number | +82-42-821-5776 |
| Fax number | +82-42-825-9754 |

| For the corresponding author (responsible for correspondence, proofreading, and reprints) | Fill in information in each box below |
|--|--|
| First name, middle initial, last name | Peng Ji |
| Email address – this is where your proofs will be sent | penji@ucdavis.edu |
| Secondary Email address | |
| Address | Department of Nutrition, University of California, Davis, CA 95616, United States of America |
| Cell phone number | |
| Office phone number | +1-530-752-6469 |
| Fax number | |

Abstract

The study was conducted to evaluate the effects of spray-dried plasma (SDP) supplementation during late gestation and lactation on productive performance and immune responses of sows and their litters. Twelve sows (227.78 ± 2.16 kg average body weight; 2.0 average parity) were randomly allotted to two dietary treatments: a basal diet (CON) and the basal diet supplemented with 1% SDP. Sows were fed experimental diets from d 30 before farrowing to weaning of their piglets. Blood samples were collected from sows on d 1, 3, and 7 of lactation and from two randomly selected nursing pigs per litter on d 3 and 7 after birth, and d 1, 3, and 7 after weaning. Productive performance and immune responses of sows and their piglets were measured. There was a trend of less body weight loss in sows supplemented with SDP ($p < 0.10$) during the lactation period and a trend of greater ($p < 0.10$) average daily gain in SDP piglets compared to those in the CON group. Sows in the SDP group tended to have lower ($p < 0.10$) serum concentrations of tumor necrosis factor- α (TNF- α), transforming growth factor- β 1 (TGF- β 1), and cortisol on d 3 and lower serum concentration of TNF- α on d 7 compared with sows in CON group. In comparison with CON piglets, piglets from SDP sows tended to have lower ($p < 0.10$) serum concentrations of TNF- α , TGF- β 1, and cortisol on d 7 after birth, lower ($p < 0.10$) serum TNF- α and C-reactive protein on d 3 and 7 after weaning, and greater ($p < 0.10$) average daily gain after weaning. Moreover, weaned pigs from sows fed SDP had significantly lower ($p < 0.05$) serum concentrations of cortisol and TGF- β 1 on d 3 and 7 postweaning, respectively, than CON piglets. In conclusion, spray-dried plasma supplementation in sow diets from late gestation to weaning improved the productive performance of sows and their offspring; the beneficial effects of SDP may be mediated in part through modulation of immune responses of both sows and piglets.

Keywords: spray-dried plasma, lactating sows, productive performance, immune responses, weaned pigs

Introduction

Reproductive performance and health status of sows are economically important factors to the profit of the swine industry. However, numerous environmental risk factors, such as disease and seasonal variation, can affect the reproductive performance of sows and herd productivity [1]. Therefore, various functional dietary supplements or feed additives were investigated for their potential benefits in enhancing the production, health and longevity of sows [2].

Spray-dried plasma (SDP) is a protein-rich feed additive in swine diets with well-balanced amino acid composition and diverse bioactive components, including immunoglobulins, peptides, glycoproteins, and others [3]. It has also been reported that SDP is superior to soybean meal and other plant-derived protein sources with respect to the contents of essential amino acids [4]. SDP is extensively used in nursery pig diet due to its advantages of improvements in health status. Previous studies have shown that supplementation of SDP improved growth performance [5,6], intestinal barrier function [7,8], and immune responses [9,10] of weaned pigs. More prominent positive effects are often observed when SDP is supplemented in diets for younger pigs that have immature immune system or pigs in the unsanitary environment [11], suggesting an immunomodulatory function of SDP. Such immunomodulatory and other beneficial effects of SDP are believed due in part to its high concentration of immunoglobulins [12,13]. Although the precise mechanism is yet unclear, there are several proposed modes of action of SDP including 1) direct effects on the barrier functions of the gastrointestinal tract [14] 2) regulation of gut-associated lymphoid tissue [15], 3) systemic effects on the respiratory [16] and reproductive systems [17]. In addition, SDP products have been suggested as a potential alternative to antibiotics based on their promising beneficial effects on pigs [14].

Supplementation of SDP products in sow diet during gestation or lactation was shown to improve productivity of sow and growth of their offspring. For instance, supplementing 0.5% SDP in gestation diet (from d 28 post-breeding until d 1 after farrowing) of sows increased body weight and average daily gain of litters at weaning [18], which was ascribed to the improved lactation performance in response to SDP supplementation. Moreover, a lower rate of preweaning mortality and greater weaning weight of piglets were reported for multiparous sows fed a diet containing 0.5% of SDP during lactating [19]. A study by Crenshaw et al. [20] also reported that supplementation of 0.5% SDP in lactation diets improved average body weight of weaned pigs and subsequent farrowing rate of sows, while postweaning sow mortality was reduced. However, information on the growth

51 performance and immune responses of nursing and weaned pigs from sows fed SDP during late gestation and
52 lactation period are still limited. Therefore, the objective of this experiment was to investigate not only the effects of
53 SDP supplementation in sow diets on productive performance and immune responses of sows in lactation period but
54 also their litters.

55 **Materials and Methods**

56 Experimental protocol was reviewed and approved by the Animal Care and Use Committee at the
57 Chungnam National University, Daejeon, Republic of Korea (Protocol # CNU-00611). The experiment was
58 performed at the animal research facility at the Chungnam National University.

59 **Animals, housing, experimental design, and diet**

60 A total of twelve lactating sows (Yorkshire × Landrace; 227.78 ± 2.16 kg average body weight; 2.0 average
61 parity) were stratified by parity and randomly assigned to one of the two dietary treatments on d 84 of gestation (6
62 sows/treatment). Sows were individually housed in farrowing crates since d 109 of gestation and had free access to
63 water. Sows were fed either a basal diet (control, CON) or the basal diet supplemented with 1% (as-fed basis) SDP.
64 The SDP used in this experiment was produced by APC Inc. (Ankeny, IA, USA). All diets met the current estimates
65 for nutrient requirements of sows (Table 1) [21]. Sows were restricted fed the experimental diets (3.0 kg/day) from d
66 84 of gestation until farrowing, and then fed the experimental diets on an ad libitum basis from until weaning of
67 their piglets.

68 Litters were weaned at 27 d of age (6.09 ± 0.26 kg average body weight) and each litter was allotted to
69 individual nursery pen. Weaned pigs were subjected to a 3-phase feeding program and fed the phase 1 nursery diet
70 at week 1, phase 2 diet at week 2 and phase 3 diet from week 3 to 6 postweaning. Weaned pigs had free access to
71 water and diets throughout the experiment.

72 **Sample collection and measurements**

73 Lactating sows and their litters' body weight were measured on the day of farrowing and weaning. Body
74 weight change of sows and average daily gain of piglets during the lactation period were calculated. The feed intake
75 for each sow was recorded during lactation period to calculate the average daily feed intake. Measurements of sow
76 backfat depth was performed on P2 position (65 mm down the left side from the spine at the same level as the last
77 rib curve) using a real-time ultrasound scanner (Anyscan BF, SongKang GLC Co., Gyeonggi-do, Republic of

78 Korea) on day of farrowing and weaning. The number of stillborn and liveborn piglets and weaned piglets were
79 measured to calculate productive performance.

80 Blood samples were collected from each sow on d 1, 3, and 7 of lactation and two randomly selected
81 piglets (1 barrow and 1 gilt) per litter on d 3 and 7 after birth and d 1, 3 and 7 after weaning using vacutainers
82 containing clot-activator or ethylenediaminetetraacetic acid (EDTA) to harvest whole blood or serum, respectively.
83 Serum samples were collected from whole blood after centrifugation (3,000 rpm for 15 min at 4°C) and kept at -
84 20°C until analysis.

85 **Measurements of white blood cell counts, serum cytokines, acute phase protein, cortisol, and** 86 **Immunoglobulins**

87 Whole blood samples were analyzed by an automatic hematology analyzer (scil Vet abc hematology
88 analyzer, scil animal care company, F-67120 Altorf, France) for total white blood cell (WBC) counts. The serum
89 concentrations of tumor necrosis factor- α (TNF- α ; R&D Systems, Inc., Minneapolis, MN, USA), transforming
90 growth factor- β 1 (TGF- β 1; R&D Systems, Inc., Minneapolis, MN, USA), C-reactive protein (Abnova Corp., Taipei
91 City, Taiwan), cortisol (Cusabio, Wuhan, China), and immunoglobulin G, M, and A (IgG, IgM, and IgA; Abnova
92 Corp., Taipei City, Taiwan) were measured using porcine-specific enzyme-linked immunosorbent assay (ELISA)
93 kits by following the instruction manufacturers' instructions.

94 **Statistical analyses**

95 All data were analyzed using the PROC GLM procedure of SAS (SAS Inst. Inc., Carry, NC, USA) in a
96 completely randomized design with the sow or their litter as an experimental unit, respectively. The model for
97 productive performance and immune responses of sows and their litters included the fixed effect of dietary treatment
98 and sow or litter as random terms; Parity of sows was used as a covariate in the model. Statistical significance and
99 tendency were considered at $p < 0.05$ and $0.05 \leq p < 0.10$, respectively.

100 **Results and discussion**

101 Sows supplemented with 1% SDP tended to have less ($p < 0.10$) body weight losses and improved ($p <$
102 0.10) piglet average daily gain during the time of lactation than those fed CON; however, there were no differences
103 in other productive performances between the groups (Table 2). Sows mobilize body reserves (protein and fat) to
104 support lactation, therefore body weight loss is commonly observed during lactation. However, excessive loss of
105 body weight during lactation can compromise reproductive performance of sows in the subsequent pregnancy,

106 including reduced rate of pregnancy and lower survival rates of embryos [22,23]. Therefore, appropriate feeding
107 practices, such as high-energy rations and nutrient-rich feeds, are required to minimize lactation weight loss [24,25].
108 Previous research reported that supplementation of 0.25% SDP in lactation diets tended to reduce body weight
109 losses of sows and significantly improved average litter weight at weaning [26]. In agreement with this study, results
110 from the current study showed that dietary supplementation of 1% SDP in late gestation and lactation is effective in
111 reducing weight loss of lactating sows and improving the growth of nursing piglets. Taken altogether, SDP may
112 mitigate body weight loss through enhancing nutrient utilization and lactation performance of sows and thus
113 enhance the growth of their litters.

114 Compared with CON sows, addition of 1% SDP tended to reduce ($p < 0.10$) serum TNF- α , TGF- β 1, and
115 cortisol concentrations on d 3 and serum TNF- α concentrations on d 7 after farrowing (Table 3). No difference was
116 observed in the white blood cell counts and serum C-reactive protein throughout the experiments for sow.
117 Parturition is the most stressful event of the reproduction cycle of sows, while improper pre- and peripartal
118 management and nutrition may lead to chronic stress that has deleterious effects on immune functions [27,28]. As a
119 result, these stress-related alterations reflect immunodeficiency, which may contribute to reproductive failure or
120 death of sows and might adversely affect the performance of their litters [29]. It was reported previously that dietary
121 supplementation of 1 or 8% SDP reduced serum concentrations of inflammation- and stress-related mediators (TNF-
122 α , C-reactive protein, and cortisol) and increased concentrations of anti-inflammatory cytokine (TGF- β 1) in the
123 uterine of pregnant mice that suffered from transportation stress [17]. In addition, pregnant mice that received 8%
124 SDP had lower pro-inflammatory cytokines in uterine mucosa and placenta and had reduced lethargic response after
125 LPS challenge [30]. These observations suggest that SDP could attenuate inflammation and enhance immune
126 competence in pregnant mice. Moreover, it was also reported that SDP maintained gut barrier function of weaned
127 pigs [7] and rats [31] in a state of inflammatory status, respectively. Thus, in the present study, the reduced body
128 weight change of sows and greater average daily gain of their litters by SDP supplementation may result from more
129 robust immune functions of sows in response to farrowing and lactation stresses.

130 Piglets from sows supplemented with SDP tended to have lower ($p < 0.10$) serum concentrations of TNF- α ,
131 TGF- β 1, and cortisol on d 7 of lactation (Table 4). However, no differences were found in serum immunoglobulins
132 of nursing piglets during lactation (Table 5). After weaning, pigs from sows supplemented with SDP tended to have
133 greater average daily gain ($p < 0.10$) compared with CON piglets (Table 6). This result was supported by

134 inflammatory and stress markers in serum. On d 3, weaned pigs from sows supplemented with SDP had reduced (p
135 < 0.05) serum cortisol concentrations, tended to have reduced ($p < 0.10$) serum TNF- α concentration than those
136 from sows in CON group (Table 7). On d 7, serum concentrations of TGF- β 1 were decreased ($p < 0.05$) and tended
137 to have reduced ($p < 0.10$) C-reactive protein concentrations in weaned pigs from sows fed SDP compared with
138 those from sows fed CON. In order to acquire passive immunity, newborn piglets have to consume a sufficient
139 volume of colostrum which contains high energy sources and immunoglobulins [32]. Previous studies reported that
140 body weight and body conditions of sows during gestation are important factors that affect the quality and quantity
141 of colostrum and milk production [33–35]. In the current study, increased piglet average daily gain at weaning and
142 improved immune responses of nursing and weaned pigs could have been due to improved health status of sows fed
143 SDP. Moreover, the possible modes of action include, but are not limited to 1) increase the growth rate of piglets
144 from sows supplemented with SDP may have been due to increased production of colostrum and milk and thus
145 increased consumption of colostrum and milk by suckling pigs; 2) sows fed SDP may have produced higher
146 concentrations of immunoglobulins in colostrum and milk, however, the amounts of immunoglobulins contained in
147 sow colostrum or milk were not analyzed, nor was colostrum or milk production of sows measured. Future research
148 is needed to elucidate the mechanisms of improved immune responses in pigs from sows supplemented with SDP.

149 **Conclusion**

150 In conclusion, supplementation of spray-dried plasma during late gestation and lactating sows' diets
151 improved productive performance and immune responses of lactating sows and their litters.

152



References

- 153
- 154 1. Stalder KJ, Knauer M, Baas TJ, Rothschild MF, Mabry JW. Sow longevity. *Pig News Inf.* 2004;25:53N-74N.
- 155 2. Kim SW, Weaver AC, Shen Y Bin, Zhao Y. Improving efficiency of sow productivity: Nutrition and health.
- 156 *J Anim Sci Biotechnol.* 2013;4:26. <https://doi.org/10.1186/2049-1891-4-26>.
- 157 3. Pettigrew JE. Reduced use of antibiotic growth promoters in diets fed to weanling pigs: Dietary tools, Part 1.
- 158 *Anim Biotechnol.* 2006;17:207–15. <https://doi.org/10.1080/10495390600956946>.
- 159 4. van Dijk AJ, Everts H, Nabuurs MJA, Margry RJCF, Beynen AC. Growth performance of weanling pigs fed
- 160 spray-dried animal plasma: a review. *Livest Prod Sci.* 2001;68:263–74.
- 161 [https://doi.org/https://doi.org/10.1016/S0301-6226\(00\)00229-3](https://doi.org/https://doi.org/10.1016/S0301-6226(00)00229-3).
- 162 5. Coffey RD, Cromwell GL. The impact of environment and antimicrobial agents on the growth response of early-
- 163 weaned pigs to spray-dried porcine plasma. *J Anim Sci.* 1995;73:2532–9. <https://doi.org/10.2527/1995.7392532x>.
- 164 6. Pierce JL, Cromwell GL, Lindemann MD, Russell LE, Weaver EM. Effects of spray-dried animal plasma and
- 165 immunoglobulins on performance of early weaned pigs. *J Anim Sci.* 2005;83:2876–85.
- 166 <https://doi.org/10.2527/2005.83122876x>.
- 167 7. Nofrarias M, Manzanilla EG, Pujols J, Gibert X, Majo N, Segalés J, Gasa J.. Effects of spray-dried porcine
- 168 plasma and plant extracts on intestinal morphology and on leukocyte cell subsets of weaned pigs. *J Anim Sci.*
- 169 2006;84:2735–42. <https://doi.org/10.2527/jas.2005-414>
- 170 8. Peace RM, Campbell J, Polo J, Crenshaw J, Russell L, Moeser A. Spray-dried porcine plasma influences
- 171 intestinal barrier function, inflammation, and diarrhea in weaned pigs. *J Nutr.* 2011;141:1312–7.
- 172 <https://doi.org/10.3945/jn.110.136796>.
- 173 9. Owusu-Asiedu A, Nyachoti CM, Marquardt RR. Response of early-weaned pigs to an enterotoxigenic
- 174 *Escherichia coli* (K88) challenge when fed diets containing spray-dried porcine plasma or pea protein isolate plus
- 175 egg yolk antibody, zinc oxide, fumaric acid, or antibiotic. *J Anim Sci.* 2003;81:1790–8.
- 176 <https://doi.org/10.2527/2003.81171790x>.
- 177 10. Bosi P, Casini L, Finamore A, Cremokolini C, Merialdi G, Trevisi P, Nobili F, mengheri E. Spray-dried plasma
- 178 improves growth performance and reduces inflammatory status of weaned pigs challenged with enterotoxigenic
- 179 *Escherichia coli* K88. *J Anim Sci.* 2004;82:1764–72. <https://doi.org/10.2527/2004.8261764x>.
- 180 11. Torrallardona D. Spray dried animal plasma as an alternative to antibiotics in weanling pigs: a review. *Asian-*

181 Australasian J Anim Sci. 2009;23:131–48. <https://doi.org/10.5713/ajas.2010.70630>.

182 12. Owen KQ, Nelssen JL, Goodband RD, Tokach MD, Friesen KG, Richert BT, Smith JW, Russell LE. Effects of
183 various fractions of spray-dried porcine plasma on performance of early weaned pigs. *J Anim Sci.* 1995;73:81.
184 <https://doi.org/10.4148/2378-5977.6470>

185 13. Jiang R, Chang X, Stoll B, Fan MZ, Arthington J, Weaver E, Campbell J, Burrin DG. Dietary plasma protein
186 reduces small intestinal growth and lamina propria cell density in early weaned pigs. *J Nutr.* 2000;130:21–6.
187 <https://doi.org/10.1093/jn/130.1.21>.

188 14. Pérez-Bosque A, Polo J, Torrallardona D. Spray dried plasma as an alternative to antibiotics in piglet feeds,
189 mode of action and biosafety. *Porc Heal Manag.* 2016;2:16. <https://doi.org/10.1186/s40813-016-0034-1>.

190 15. Lalles JP, Bosi P, Janczyk P, Koopmans SJ, Torrallardona D. Impact of bioactive substances on the
191 gastrointestinal tract and performance of weaned piglets: a review. *Animal.* 2009;3:1625–43.

192 16. Maijó M, Miró L, Polo J, Campbell J, Russell L, Crenshaw J, Weaver E, Moretó M, Pérez-Bosque A. Dietary
193 plasma proteins attenuate the innate immunity response in a mouse model of acute lung injury. *Br J Nutr.*
194 2012;107:867–75. <https://doi.org/10.1017/S0007114511003655>.

195 17. Song M, Liu Y, Lee JJ, Che TM, Soares-Almeida JA, Chun JL, Campbell JM, Polo J, Crenshaw JD, Seo SW,
196 Pettigrew JE. Spray-dried plasma attenuates inflammation and improves pregnancy rate of mated female mice. *J*
197 *Anim Sci.* 2015;93:298–305. <https://doi.org/10.2527/jas.2014-7259>.

198 18. Crenshaw JD, Campbell JM, Russell LE, Greiner LL, Soto J, Connor JF. Effect of spray-dried plasma fed during
199 gestation on pig performance at weaning. *Proc. 37th Allen D. Leman Swine Conf., St. Paul, MN.* p. 193.

200 19. Frugé ED, Roux ML, Lirette RD, Bidner TD, Southern LL, Crenshaw JD. Effects of dietary spray-dried plasma
201 protein on sow productivity during lactation. *J Anim Sci.* 2009;87:960–4. <https://doi.org/10.2527/jas.2008-1353>.

202 20. Crenshaw JD, Campbell JM, Russell LE, Sonderman JP. Effect of spray-dried plasma in diets fed to lactating
203 sows on litter weight at weaning and subsequent farrowing rate. *Proc. 37th Allen D. Leman Swine Conf., St. Paul,*
204 *MN.* p. 47.

205 21. National Research Council (NRC). *Nutrient requirements of swine: Eleventh revised edition.* Washington, DC:
206 The National Academies Press; 2012. <https://doi.org/10.17226/13298>.

207 22. Close WH, Mullan BP. Nutrition and feeding of breeding stock. *Pig Prod.* 1996:169–202.

208 23. Thaker MYC, Bilkei G. Lactation weight loss influences subsequent reproductive performance of sows. *Anim*

209 Reprod Sci. 2005;88:309–18. <https://doi.org/https://doi.org/10.1016/j.anireprosci.2004.10.001>.

210 24. Kirkwood RN, Mitaru BN, Gooneratne AD, Blair R, Thacker PA. The influence of dietary energy intake during
211 successive lactations on sow prolificacy. *Can J Anim Sci.* 1988;68:283–90. <https://doi.org/10.4141/cjas88-029>.

212 25. Bilkei G. Herd health strategy for improving the reproductive-performance of pigs. *Hung Vet J.* 1995;50:766–8.

213 26. Crenshaw JD, Boyd RD, Campbell JM, Russell LE, Moser RL, Wilson ME. Lactation feed disappearance and
214 weaning to estrus interval for sows fed spray-dried plasma. *J Anim Sci.* 2007;85:3442–53.
215 <https://doi.org/10.2527/jas.2007-0220>.

216 27. Cronin GM, Barnett JL, Hodge FM, Smith JA, McCallum TH. The welfare of pigs in two farrowing/lactation
217 environments: cortisol responses of sows. *Appl Anim Behav Sci.* 1991;32:117–27. [https://doi.org/10.1016/S0168-](https://doi.org/10.1016/S0168-1591(05)80036-X)
218 1591(05)80036-X.

219 28. Jarvis S, D'Eath RB, Robson SK, Lawrence AB. The effect of confinement during lactation on the
220 hypothalamic–pituitary–adrenal axis and behaviour of primiparous sows. *Physiol Behav.* 2006;87:345–52.
221 <https://doi.org/10.1016/j.physbeh.2005.10.004>.

222 29. Johnson AK, Marchant-Forde JN. Welfare of pigs in the farrowing environment - The welfare of pigs. In:
223 Marchant-Forde JN, editors. Springer Netherlands; 2009, p. 141–88. [https://doi.org/10.1007/978-1-4020-8909-](https://doi.org/10.1007/978-1-4020-8909-1_5)
224 1_5.

225 30. Liu Y, Choe J, Lee JJ, Kim J, Campbell JM, Polo J, Crenshaw JD, Pettigrew JE, Song M. Spray-dried plasma
226 attenuates inflammation and lethargic behaviors of pregnant mice caused by lipopolysaccharide. *PLoS One.*
227 2018;13:e0203427. <https://doi.org/10.1371/journal.pone.0203427>

228 31. Pérez-Bosque A, Amat C, Polo J, Campbell JM, Crenshaw J, Russell L, Moretó M. Spray-dried animal plasma
229 prevents the effects of *Staphylococcus aureus* enterotoxin B on intestinal barrier function in weaned rats. *J Nutr.*
230 2006;136:2838–2843. <https://doi.org/10.1093/jn/136.11.2838>

231 32. Theil PK, Lauridsen C, Quesnel H. Neonatal piglet survival: impact of sow nutrition around parturition on fetal
232 glycogen deposition and production and composition of colostrum and transient milk. *Animal.* 2014;8:1021–30.
233 <https://doi.org/10.1017/S1751731114000950>.

234 33. King RH, Mullan BP, Dunshea FR, Dove H. The influence of piglet body weight on milk production of sows.
235 *Livest Prod Sci.* 1997;47:169–74. [https://doi.org/10.1016/S0301-6226\(96\)01404-2](https://doi.org/10.1016/S0301-6226(96)01404-2)

236 34. King RH. Factors that influence milk production in well-fed sows. *J Anim Sci.* 2000;78:19–25.

237 https://doi.org/10.2527/2000.78suppl_319x.

238 35. Theil PK, Nielsen MO, Sørensen MT, Lauridsen C. Lactation, milk and suckling. *Nutr Physiol Pigs Danish Pig*

239 *Res Centre, Copenhagen, Denmark.* 2012:1–47.

240

ACCEPTED

241 **Table 1. Ingredient composition of experimental diets (as-fed basis)**

| Item ¹ | Gestation diet | | Lactation diet | |
|---------------------------------|----------------|--------|----------------|--------|
| | CON | SDP | CON | SDP |
| Ingredients (%) | | | | |
| Corn | 75.82 | 76.72 | 65.54 | 66.53 |
| Soybean meal | 21.30 | 19.40 | 31.81 | 29.82 |
| SDP ² | - | 1.00 | - | 1.00 |
| Limestone | 0.90 | 0.90 | 0.85 | 0.85 |
| MDCP | 1.58 | 1.58 | 1.40 | 1.40 |
| Vitamin premix ³ | 0.20 | 0.20 | 0.20 | 0.20 |
| Mineral premix ⁴ | 0.20 | 0.20 | 0.20 | 0.20 |
| Total | 100.00 | 100.00 | 100.00 | 100.00 |
| Calculated chemical composition | | | | |
| ME (Mcal/kg) | 3.32 | 3.32 | 3.43 | 3.43 |
| CP (%) | 15.86 | 15.82 | 19.76 | 19.72 |
| Crude fat (%) | 3.09 | 3.05 | 2.86 | 2.80 |
| Crude fiber (%) | 2.97 | 3.00 | 3.33 | 3.36 |
| NDF (%) | 8.71 | 8.78 | 10.78 | 10.81 |
| ADF (%) | 4.18 | 4.20 | 4.63 | 4.65 |
| Calcium (%) | 0.77 | 0.77 | 0.75 | 0.75 |
| Phosphorus (%) | 0.64 | 0.65 | 0.65 | 0.65 |
| Lysine (%) | 0.74 | 0.76 | 1.02 | 1.02 |
| Methionine (%) | 0.25 | 0.25 | 0.30 | 0.30 |
| Threonine (%) | 0.58 | 0.60 | 0.74 | 0.76 |
| Tryptophan (%) | 0.16 | 0.16 | 0.22 | 0.22 |

242 ¹CON, control sow diet based on corn and soybean meal; SDP, CON + 1% spray-dried plasma; MDCP,
 243 monocalcium phosphate; ME, metabolizable energy; CP, crude protein; NDF, neutral detergent fiber; ADF, acid
 244 detergent fiber.

245 ²Spray-dried plasma (APC Inc., Ankeny, IA).

246 ³Provided per kilogram of diet: vitamin A, 10,000 IU; vitamin D₃, 2,000 IU; vitamin E, 48 IU; vitamin K₃,
 247 1.5 mg; riboflavin, 6 mg; niacin, 40 mg; D-pantothenic acid, 17 mg; biotin, 0.2 mg; folic acid, 2 mg; choline, 166
 248 mg; vitamin B₆, 2 mg; and vitamin B₁₂, 28 µg.

249 ⁴Provided per kilogram of diet: Fe, 90 mg from iron sulfate; Cu, 15 mg from copper sulfate; Zn, 50 mg
250 from zinc oxide; Mn, 54 mg from manganese oxide; I, 0.99 mg from potassium iodide; Se, 0.25 mg from sodium
251 selenite.
252

ACCETED

253 **Table 2. Productive performance of lactating sows fed diets supplemented with spray-dried plasma¹**

| Item ² | Dietary treatments | | SEM | p-value |
|--|--------------------|--------|------|---------|
| | CON | SDP | | |
| Parity | 2.1 | 2.0 | 0.30 | 0.733 |
| Lactation days (d) | 28.00 | 26.16 | 0.76 | 0.118 |
| Initial BW on d 1 after farrowing (kg) | 226.14 | 229.42 | 3.12 | 0.471 |
| Final BW at weaning (kg) | 208.80 | 218.61 | 3.73 | 0.092 |
| Sow BW change (kg) | -17.34 | -10.81 | 2.45 | 0.081 |
| Total feed intake (kg) | 183.83 | 180.48 | 4.07 | 0.530 |
| ADFI (kg) | 6.57 | 6.89 | 0.11 | 0.114 |
| Initial backfat depth (mm) | 20.50 | 20.75 | 1.88 | 0.789 |
| Final backfat depth (mm) | 17.20 | 17.26 | 1.53 | 0.987 |
| Backfat depth change (mm) | 3.30 | 3.49 | 1.50 | 0.856 |
| Born alive piglets (n) | 11.16 | 10.20 | 1.42 | 0.655 |
| Dead piglets (n) ³ | 1.50 | 1.20 | 0.58 | 0.437 |
| Pre-weaning mortality (%) | 13.44 | 11.76 | 3.47 | 0.563 |
| Weaned piglets (n) | 9.66 | 9.00 | 0.28 | 0.284 |
| Piglets BW at birth (kg) | 1.60 | 1.80 | 0.07 | 0.145 |
| Piglets BW at weaning (kg) | 5.82 | 6.35 | 0.41 | 0.439 |
| ADG of piglets (g/d) | 150.15 | 173.03 | 7.58 | 0.082 |

254 ¹Values are presented as the least squares mean of 6 replicates.

255 ²CON, control sow diet based on corn and soybean meal; SDP, CON + 1% spray-dried plasma; BW, body
 256 weight; ADFI, average daily feed intake; ADG, average daily gain; SEM, standard error of mean.

257 ³Stillborn and piglets died before weaning.

258

259 **Table 3. Immune responses of lactating sows fed diets supplemented with spray-dried plasma¹**

| Item ² | Dietary treatments | | SEM | p-value |
|----------------------------|--------------------|--------|-------|---------|
| | CON | SDP | | |
| Day 1 of lactation | | | | |
| WBC (x10 ³ /μL) | 11.93 | 9.55 | 1.77 | 0.893 |
| TNF-α (pg/mL) | 312.30 | 289.80 | 34.19 | 0.809 |
| TGF-β1 (pg/mL) | 357.66 | 238.16 | 59.23 | 0.745 |
| CRP (ng/mL) | 241.43 | 268.87 | 44.60 | 0.846 |
| Cortisol (ng/mL) | 0.51 | 0.49 | 0.06 | 0.345 |
| Day 3 of lactation | | | | |
| WBC (x10 ³ /μL) | 15.33 | 14.08 | 1.58 | 0.465 |
| TNF-α (pg/mL) | 281.96 | 264.94 | 5.06 | < 0.10 |
| TGF-β1 (pg/mL) | 448.07 | 311.37 | 63.21 | < 0.10 |
| CRP (ng/mL) | 139.95 | 132.90 | 18.71 | 0.723 |
| Cortisol (ng/mL) | 0.56 | 0.48 | 0.03 | < 0.10 |
| Day 7 of lactation | | | | |
| WBC (x10 ³ /μL) | 13.71 | 13.98 | 1.64 | 0.687 |
| TNF-α (pg/mL) | 272.15 | 249.35 | 10.21 | < 0.10 |
| TGF-β1 (pg/mL) | 286.20 | 257.32 | 46.00 | 0.854 |
| CRP (ng/mL) | 136.92 | 105.27 | 23.21 | 0.412 |
| Cortisol (ng/mL) | 0.49 | 0.51 | 0.06 | 0.597 |

260 ¹Values are presented as the least squares mean of 6 replicates.

261 ²CON, control sow diet based on corn and soybean meal; SDP, CON + 1% spray-dried plasma; WBC,
 262 white blood cells; TNF-α, tumor necrosis factor-α; TGF-β1, transforming growth factor-β1; CRP, C-reactive
 263 protein; SEM, standard error of mean.

264 **Table 4. Immune responses of nursing piglets from sows fed diets supplemented with spray-dried plasma¹**

| Item ² | Dietary treatments | | SEM | p-value |
|----------------------------|--------------------|--------|--------|---------|
| | CON | SDP | | |
| Day 3 of lactation | | | | |
| WBC (x10 ³ /μL) | 10.15 | 8.54 | 0.92 | 0.687 |
| TNF-α (pg/mL) | 270.39 | 262.67 | 40.86 | 0.741 |
| TGF-β1 (pg/mL) | 788.76 | 607.40 | 140.86 | 0.874 |
| CRP (ng/mL) | 72.14 | 78.15 | 16.87 | 0.812 |
| Cortisol (ng/mL) | 0.83 | 0.45 | 0.23 | 0.345 |
| Day 7 of lactation | | | | |
| WBC (x10 ³ /μL) | 13.15 | 12.98 | 1.30 | 0.465 |
| TNF-α (pg/mL) | 423.57 | 349.87 | 38.99 | < 0.10 |
| TGF-β1 (pg/mL) | 980.41 | 853.49 | 58.99 | < 0.10 |
| CRP (ng/mL) | 119.42 | 115.18 | 26.95 | 0.586 |
| Cortisol (ng/mL) | 1.05 | 0.62 | 0.15 | < 0.10 |

265 ¹Values are presented as the least squares mean of 6 replicates.

266 ²CON, control sow diet based on corn and soybean meal; SDP, CON + 1% spray-dried plasma; WBC,
 267 white blood cells; TNF-α, tumor necrosis factor-α; TGF-β1, transforming growth factor-β1; CRP, C-reactive
 268 protein; SEM, standard error of mean.

269

270 **Table 5. Serum immunoglobulins of nursing piglets from sows fed diets supplemented with spray-dried**
 271 **plasma¹**

| Item ² | Dietary treatments | | SEM | <i>p</i> -value |
|--------------------|--------------------|--------|-------|-----------------|
| | CON | SDP | | |
| Day 3 of lactation | | | | |
| IgG (ng/mL) | 242.13 | 251.47 | 9.95 | 0.818 |
| IgM (ng/mL) | 89.69 | 106.97 | 10.16 | 0.235 |
| IgA (ng/mL) | 30.22 | 25.71 | 5.38 | 0.844 |
| Day 7 of lactation | | | | |
| IgG (ng/mL) | 219.28 | 221.83 | 13.62 | 0.795 |
| IgM (ng/mL) | 58.46 | 69.95 | 7.79 | 0.612 |
| IgA (ng/mL) | 22.65 | 33.69 | 4.40 | 0.692 |

272 ¹Values are presented as the least squares mean of 6 replicates.

273 ²CON, control sow diet based on corn and soybean meal; SDP, CON + 1% spray-dried plasma; IgG,
 274 immunoglobulin G; IgM, immunoglobulin M; IgA, immunoglobulin A; SEM, standard error of mean.

275

276 **Table 6. Growth performance of weaned pigs from sows fed diets supplemented with spray-dried plasma¹**

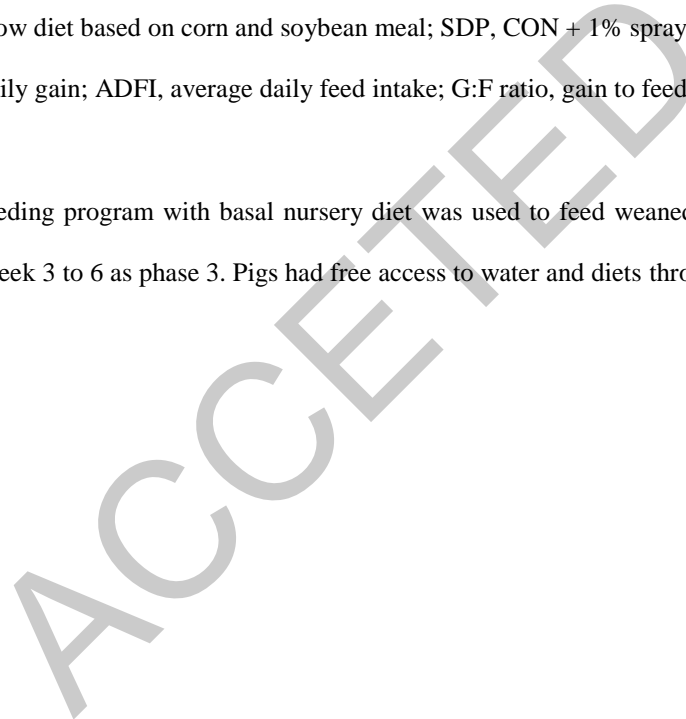
| Item ² | Dietary treatments | | SEM | <i>p</i> -value |
|-------------------------------|--------------------|--------|-------|-----------------|
| | CON | SDP | | |
| Number of weaned pigs/pen (n) | 9.66 | 9.00 | | |
| Initial BW (kg) | 5.82 | 6.35 | 0.41 | 0.439 |
| Final BW ³ (kg) | 23.23 | 26.11 | 0.96 | 0.102 |
| Feed intake (kg) | 25.50 | 26.85 | 0.53 | 0.348 |
| ADG (g/d) | 414.52 | 470.47 | 18.21 | < 0.10 |
| ADFI (g/d) | 607.17 | 639.24 | 0.14 | 0.345 |
| G:F ratio (g/g) | 0.682 | 0.735 | 0.03 | 0.773 |

277 ¹Values are presented as the least squares mean of 6 replicates.

278 ²CON, control sow diet based on corn and soybean meal; SDP, CON + 1% spray-dried plasma; BW, body
 279 weight; ADG, average daily gain; ADFI, average daily feed intake; G:F ratio, gain to feed ratio; SEM, standard error
 280 of mean.

281 ³The 3-phase feeding program with basal nursery diet was used to feed weaned pigs: week 1 as phase 1;
 282 week 2 as phase 2; and week 3 to 6 as phase 3. Pigs had free access to water and diets throughout the experiment.

283



284 **Table 7. Immune responses of weaned pigs from sows fed diets supplemented with spray-dried plasma¹**

| Item ² | Dietary treatments | | SEM | p-value |
|----------------------------|--------------------|---------|--------|---------|
| | CON | SDP | | |
| Day 1 postweaning | | | | |
| WBC (x10 ³ /μL) | 12.91 | 13.42 | 1.46 | 0.412 |
| TNF-α (pg/mL) | 320.24 | 358.90 | 57.21 | 0.847 |
| TGF-β1 (pg/mL) | 1121.44 | 1077.46 | 117.21 | 0.765 |
| CRP (ng/mL) | 87.53 | 76.54 | 14.94 | 0.546 |
| Cortisol (ng/mL) | 1.34 | 1.18 | 0.10 | 0.387 |
| Day 3 postweaning | | | | |
| WBC (x10 ³ /μL) | 16.09 | 13.44 | 2.20 | 0.341 |
| TNF-α (pg/mL) | 449.80 | 344.11 | 38.32 | < 0.10 |
| TGF-β1 (pg/mL) | 1071.75 | 1024.15 | 158.32 | 0.687 |
| CRP (ng/mL) | 126.60 | 103.58 | 17.30 | 0.541 |
| Cortisol (ng/mL) | 1.88 | 1.40 | 0.16 | < 0.05 |
| Day 7 postweaning | | | | |
| WBC (x10 ³ /μL) | 16.95 | 16.15 | 1.25 | 0.741 |
| TNF-α (pg/mL) | 388.84 | 370.12 | 41.89 | 0.601 |
| TGF-β1 (pg/mL) | 836.48 | 718.33 | 41.89 | < 0.05 |
| CRP (ng/mL) | 112.28 | 78.41 | 15.50 | < 0.10 |
| Cortisol (ng/mL) | 1.74 | 1.56 | 0.11 | 0.230 |

285 ¹Values are presented as the least squares mean of 6 replicates.

286 ²CON, control sow diet based on corn and soybean meal; SDP, CON + 1% spray-dried plasma; WBC,
 287 white blood cells; TNF-α, tumor necrosis factor-α; TGF-β1, transforming growth factor-β1; CRP, C-reactive
 288 protein; SEM, standard error of mean.