1 2	Novel zinc sources as antimicrobial growth promoters for monogastric animals: A review
3	Xin Jian Lei ^{a,b,1} , Zhang Zhuang Liu ^{c,1} , and In Ho Kim ^{b*}
4	
5	^a College of Animal Science and Technology, Northwest A&F University, Yangling, Shaanxi,
6	China
7	^b Department of Animal Resource and Science, Dankook University, Cheonan, 330714, South
8	Korea
9	°College of Veterinary Medicine, Northwest A&F University, Yangling, Shaanxi, China
10	
11	
12	
13	* Corresponding author: In Ho Kim
14	Department of Animal Resource and Science, Dankook University, Cheonan,
15	Chungnam, South Korea
16	E-mail: inhokim@dankook.ac.kr
17	Tel: +82-41-550-3652
18	Fax: +82-41-559-7881
19	¹ These two authors contributed equally to this work.
20	

ORC	<u>))</u>	Xin Jian Lei (https://orcid.org/0000-0001-7348-9239) Zhuangzhuang Liu (https://orcid.org/0000-0001-7669-8853) In Ho Kim (https://orcid.org/0000-0001-6652-2504)
<u>Auth</u> Contrib		Conceptualization: Lei XJ, Kim IH Writing - original draft: Lei XJ, Liu Z Writing - review & editing: Lei XJ, Liu Z, Kim IH
<u>Fund</u> inform		This manuscript has not received any funding
Confli inter		No potential conflict of interest relevant to this article was reported
IRB/IA appro		This manuscript does not require IRB/IACUC approval because there are no human and animal participants.

24 ABSTRACT: The essentiality of zinc for animals has been recognized over 80 years. Zinc is a 25 n essential trace element that is a component of many enzymes and is associated with the vario 26 us hormones. Apart from the nutritional function, zinc has antimicrobial property and often be supplemented in diets in the quantities greater than which is required to meet the nutritional re 27 28 quirement, especially for weaning pigs. This review will focus on the application of pharmacol ogical zinc and its mechanisms which may be responsible for the effects of zinc on performan 29 ce and health of monogastric animals. Various novel sources of zinc in non-ruminant animal p 30 31 roduction will also be discussed. These should assist in more precisely formulating feed to ma 32 ximize the production performance and to maintain the health condition of monogastric animal 33 s.

34 Key words: novel zinc source; pharmacological zinc; monogastric animals



35 INTRODUCTION

36 Zinc, an essential trace element, is a component of many metalloenzymes including DNA and 37 RNA synthetases and many digestive enzymes [1]. The essentiality of zinc for animals has 38 been recognized in 1933 [2]. Besides, zinc has a regulatory role in the immune system and 39 deficiency in zinc may impair the proper immune function [3]. The concentration of zinc in body increases during the initial period of postnatal ontogenesis (2 weeks for chicks and 4 40 41 weeks for piglets), after which it remains approximately constant. Apart from the nutritional 42 function, unlike most other minerals, copper and zinc have antimicrobial properties. Therefore, they are often supplemented in diets in the quantities greater than which is required to meet the 43 nutritional requirement, especially for weaning pigs [4]. In practice, the use of 44 pharmacological levels of zinc ranging from 2,000 to 4,000 mg/kg (that this "practice" is 45 forbidden in the EU) in the form of zinc oxide is a common recommendation to reduce post-46 weaning diarrhea and improve growth performance in weaning pigs [5]. However, the use of 47 high doses zinc oxide has raised serious concern related to microbial resistance development 48 [6-8]. Moreover, due to its low absorption, high amount of zinc excretion has motivated 49 50 environmental concerns [9-11]. Those provoke the question about the reasonability of pharmacological levels of zinc supplementation as a solution of the ban of antimicrobial 51 52 growth promoters.

To improve the bioavailability of zinc, several efforts have been made including: changing conventional zinc oxide powder into porous particles or nanoparticles, organic zinc (eg, zinc lactate, zinc amino acid, zinc chelate), and the application of enteric coating method [12-17]. Phytate from whole grains and some vegetables, by forming indigestible complexes with zinc in intestine strongly interferes with zinc absorption [18]. Therefore, it is suggested that supplementation of phytase in the diet can be a useful strategy to improve bioavailability of 59 zinc [19]. This review will focus on the application of pharmacological zinc and its 60 mechanisms which may be responsible for the effects of zinc on performance and health of 61 monogastric animals. various novel sources of zinc in non-ruminant animal production will 62 also be discussed. These should assist in more precisely formulating feed to maximize the 63 production performance and to maintain the health condition of monogastric animals.

64 **P**I

PHARMACOLOGICAL ZINC

65 Traditionally, antibiotics are included in the diet of young animals for the prevention of postweaning diarrhea and enhancement of growth performance in the worldwide. However, the 66 use of in-feed antibiotics has been fully or gradually banned by many countries including 67 European countries, United States of America, South Korea and China due to the development 68 of antibacterial resistance and antibiotic residues [20-23]. Supplementation of pharmacological 69 doses of zinc in the form of zinc oxide has been proposed as one of the most effective feed 70 additives to replace antibiotics and is already widely commercialized in several countries [24-71 72 27]. Pharmacological levels of zinc oxide have shown antimicrobial properties and are used to fight against post-weaning infections and improve growth performance [28-31]. 73

74 The exact mechanisms regarding pharmacological doses of zinc oxide exert beneficial effects 75 on alleviating post-weaning diarrhea and promoting growth performance are unknown, but 76 potential theories proposed include: (1) pharmacological doses of zinc oxide may reduce 77 intestinal permeability and improve intestinal morphology and appetite [32,33]. Carlson [34] observed that zinc oxide reduced the intestinal mucosal susceptibility to secretagogues which 78 79 activate chloride secretion. In addition, it is suggested that zinc oxide may increase insulin-like 80 growth factor-1, 2 expressions in the small intestinal mucosa [35]. (2) pharmacological doses of zinc oxide have a antimicrobial effects on gram negative bacteria and beneficial effects on 81 82 the stability and diversity of gastrointestinal microbial balance, especially coliforms [36, 37]. 83 (3) pharmacological doses of zinc oxide can improve animals' defense function indicated as 84 improved immune and anti-inflammatory responses [38]. Nevertheless, the use of high doses of zinc is criticized due to the developing bacterial resistance and the excretion of a large 85 amount zinc which may raise environmental concerns [39, 40]. Although the use of 86 87 pharmacological doses of zinc oxide are allowed be in most countries and regions, the 88 European legislation limits the use of zinc oxide in animal production to a maximum of 150 mg/kg [41]. Thus, considerable efforts have been made to improve the bioavailability of zinc, 89 including organic zinc, nanosized zinc oxide, coated zinc oxide, and porous zinc oxide. 90

91 COATED ZINC OXIDE

Enteric coating is a common technology used to protect oral medications against the influence 92 of stomach juices in the field of pharmacy [42]. In feed additives production, enteric coating 93 technology also has been used for a long time [43,44]. Under the protection of outer enteric 94 coating, the inner targeted component can safely pass through the stomach. Whereas, the 95 coating materials can be gradually degraded in the gastrointestinal tract, and the inter zinc 96 oxide will be released slowly within the intestine [13]. Park et al. [9] suggested that the 97 physiological level (100 mg/kg) of zinc from coated zinc oxide showed the similar growth 98 99 promoting effects as pharmacological zinc oxide, but activities of maltase and sucrase in the 100 intestinal mucosa and pancreatic tissue and small intestinal mucosal morphology were not 101 affected by dietary treatments in weaning pigs. However, the effects of coated zinc oxide in 102 pigs were not consistent. Byun et al. [45] indicated that inclusion of coated zinc had no effects 103 on growth performance, intestinal mucosal morphology, and fecal consistency in weaning pigs. 104 Song et al. [46] observed that sub-pharmacological levels (100, 200, or 400 mg/kg) zinc from 105 coated zinc oxide had no effects on growth performance and fecal consistency but reduced

106 villus height and the ratio of villus height to crypt depth in weaning pigs. Shen et al. [18] 107 concluded that low concentrations (380 or 570 mg/kg) of zinc from coated zinc oxide or 2,250 108 mg/kg zinc from conventional zinc oxide could alleviate the occurrence of diarrhea by 109 promoting intestinal morphology, protecting intestinal mucosal barrier from damage, 110 stimulating mucosal immune system, and regulating microbial composition. Moreover, pigs 111 offered diets with coated zinc oxide excreted less zinc in feces compared with those fed those 112 fed conventional zinc oxide. In enterotoxigenic Escherichia coli challenged weaning pigs, 113 Kwon et al. [47] found that both low dose (100 mg/kg) of coated zinc oxide and high dose 114 (2,500 mg/kg) of conventional zinc oxide improved growth rate, reduced Escherichia coli shedding, increased goblet cell density in small intestine, and decreased gastrointestinal tract 115 pH values in post-challenge period. In a recent study, Upadhaya et al. [48] indicated that use 116 of coated zinc oxide in lower doses (250, 500, 750, and 1000 mg/kg) was possible to substitute 117 high dose of conventional zinc oxide in improving growth performance, nutrient digestibility, 118 and intestinal microbial balance in weaning pigs. Importantly, compared with high level of 119 conventional zinc oxide, the zinc excretion was decreased by use of low doses of coated zinc 120 oxide. To our best knowledge, no information is available on the application of coated zinc 121 122 oxide in poultry, at least for broilers and layers. Thus, researches are required to determine the effects of coated zinc oxide in poultry. 123

124 ORGANIC ZINC OXIDE

Organic mineral sources for dietary supplementation, such as proteinate and amino acid chelate, have become popular in feed industry in the past 2 decades due to their higher bioavailability [49]. In weaning pigs, Wang et al. [50] observed that low levels (50 and 100 mg/kg) of zinc from zinc glycine chelate could improve growth rate and serum alkaline

129 phosphatase and copper/zinc superoxide dismutase activities as 3,000 mg/kg of zinc as conventional zinc oxide. But zinc excretion in feces was decreased in pigs offered zinc glycine 130 131 chelate diets compared with pigs receiving diet supplemented with high level conventional 132 zinc oxide. Case and Carlson [51] indicated that weaning pigs fed 500 mg/kg zinc in the form 133 of an organic zinc-polysaccharide complex had comparable production performance but 134 excreted less zinc in feces compared with pigs fed 3,000 mg/kg zinc as zinc oxide. Buff et al. [52] indicated that supplementation of 300 or 450 mg/kg zinc from zinc-polysaccharide 135 136 allowed pigs obtained similar growth performance as 2,000 mg/kg zinc from conventional 137 zinc oxide, but 300 mg/kg zinc as zinc-polysaccharide reduced 76% of fecal zinc compared with 2,000 mg/kg zinc as conventional zinc oxide. Cheng et al. [53] suggested that both 100 138 mg/kg of zinc from zinc lysine complex and zinc sulfate were equally effective in maintaining 139 production performance and zinc absorption of weaning pigs. In sows, Payne et al. [54] found 140 that supplementation of zinc from organic zinc (zinc amino acid complex) could improve the 141 142 development of fetuses during gestation, thus resulting in increased piglets weight at birth and weaning. Moghaddam and Jahanian [55] replaced 75% inorganic zinc with dietary zinc-143 144 methionine and observed improvement in both cellular and humoral functions of the immune 145 system in broilers. Ao et al. [56] reported that chelated zinc proteinate could be used as a zinc 146 source. 12 mg/kg of zinc from chelated zinc proteinate allowed broilers obtained optimal 147 growth performance, whereas when inclusion of phytase in the diet, the optimal dose of zinc 148 from chelated zinc proteinate to meet the requirement of broiler was 7.4 mg/kg. Rossi et al. 149 [57] supplemented graded levels (0, 15, 30, 45, or 60 mg/kg) of organic zinc (chelated zinc 150 proteinate) in male chicks did not affect production performance but improved carcass quality by increasing resistance of skin to tearing. Using the same organic zinc oxide-chelated zinc 151 152 proteinate, Salim et al. [58] reported that inclusion of 25 mg/kg chelated zinc proteinate had no effect on growth performance and skin quality of broiler chickens, but increased the zinccontent in thigh meat and calcium content in plasma.

155 NANOSIZED ZINC OXIDE

156 Nanotechnology has revolutionized the commercial application of nanosized minerals in the 157 areas of engineering, food, biological, and pharmacological applications [11]. Due to better 158 antibacterial property than conventional zinc oxide, nanosized zinc oxide is the third highest 159 produced nanosized metal after nanosized silicon dioxide and nanosized titanium dioxide 160 [59,60]. Reducing the size of the material to the nano scale may modify the physico-chemical 161 properties compared to the same material at larger-size scales, such as much larger surface to mass ratio, improved surface reactivity or increased ion release [61]. The nanosized zinc oxide 162 may expose more molecules of zinc oxide to interact with the gastrointestinal tissues and 163 164 microbial population. In Hendrix laying hens, Tsai et al. [62] observed that supplementation of 20 mg/kg zinc from nanosized zinc oxide (40 mg/kg zinc in basal diet) enhanced zinc 165 retention and serum zinc, eggshell thickness, growth hormone, and carbonic anhydrase activity. 166 Mao and Lien [63] reported that compared with conventional zinc oxide, supplementation 167 with 100 mg/kg nanosized zinc oxide in Hendrix laying hens showed better effects on feed 168 169 intake, serum zinc, ghrelin, metallothionein, immunoglobulin G (IgG), and eggshell strength. 170 Zhao et al. [14] indicated that addition of 20 and 60 mg/kg nanosized zinc oxide improved 171 growth performance (body weight gain) and antioxidant capacity (increased catalase activity 172 and total antioxidant capacity in serum and reduced malondialdehyde in serum and liver) 173 compared with 60 mg/kg conventional zinc oxide, whereas 100 mg/kg nanosized zinc oxide 174 impaired growth rate. Although growth performance were unaffected, Li et al. [64] suggested 175 that supplementation of 120 mg/kg both conventional zinc oxide and nanosized zinc oxide

176 improved nutrient (crude protein, crude fat, phosphorus, and zinc) digestibility, blood IgG, and 177 γ -globulin concentrations in weaning pigs. Moreover, pigs fed diet supplemented with 178 nanosized zinc oxide had greater digestibility of phosphorus and zinc, blood phytohemagglutinin skin challenge, IgG, and y-globulin values, growth hormone, carbonic 179 180 anhydrase activity, and zinc concentrations in serum, compared with those fed diet with 181 conventional zinc oxide. Milani et al. [10] suggested that low doses (15, 30, and 60 mg/kg) 182 nanosized zinc oxide were not effective in improving production performance and controlling 183 post-weaning diarrhea of weaning piglets, although increasing nanosized zinc oxide levels 184 increased gain to feed ratio and reduced diarrhea occurrence during first week post-weaning. In addition, supplementation of low doses of nanosized zinc oxide increased dry matter and 185 crude protein digestibility. More recently, Wang et al. [65] showed that supplementation of 186 800 mg/kg nanosized zinc oxide had comparable effects to 3,000 mg/kg conventional zinc 187 oxide on improving growth rate, alleviating diarrhea, and improving intestinal morphology in 188 weaning pigs. Importantly, fecal zinc was lower in pigs fed diets supplemented with low doses 189 nanosized zinc oxide than those fed high dose of conventional zinc oxide. 190

191 TETRABASIC ZINC CHLORIDE

Tetrabasic zinc chloride is produced through a reactive crystallisation process in which zinc chloride is reacted with ammoniated zinc chloride and water. Tetrabasic zinc chloride is intended to supply zinc in final feed for all species [66]. In weaning pigs, Mavromichalis et al. [67] found that with or without antibacterial compound (carbadox), at pharmacological levels, tetrabasic zinc chloride was an effective source of zinc for enhancing growth performance. Zhang and Guo [68] conducted an experiment to evaluate pharmacological doses of zinc from tetrabasic zinc chloride (1500, 2250, and 3000 mg/kg) and conventional zinc oxide (2250 and

199 3000 mg/kg) in weaning pigs. The results reported that supplementation with both tetrabasic 200 zinc chloride and conventional zinc oxide increased daily gain, feed intake, and gain to feed 201 ratio, but decreased fecal scores in weaning pigs. The relative bioavailability of zinc from 202 tetrabasic zinc chloride was 159%, 125%, and 128%, 123%, and 122%, respectively, 203 compared with conventional zinc oxide in broilers [68]. In New Hampshire \times Columbian 204 female chicks, Batal et al. [69] suggested that tetrabasic zinc chloride provided same 205 bioavailable zinc as that of analytical-grade zinc sulfate. However, to promote the growth 206 performance, tetrabasic zinc chloride has to be included at pharmacological levels, and therefore, the problem of high zinc excretion is still present. 207

208 POROUS ZINC

High porosity zinc oxide is a novel form of zinc oxide with enhanced surface area and more 209 210 importantly greater porosity (as much as ten times higher than conventional zinc oxide) and this allows it even stronger efficacy than nanosized zinc as it further increases the possible 211 sites of the interaction among the molecules of zinc oxide, the animals, and microbial 212 population in the gastrointestinal tract (Animine, Sillingy, France) [70]. Vahjen et al. [71] 213 214 reported that porous zinc oxide exhibited higher ex vivo bacterial growth depressing effects 215 than conventional zinc oxide. In weaning pigs, Morales et al. [72] suggested that pigs fed diet 216 supplemented with 110 mg/kg zinc from porous zinc oxide increased growth rate and gain to feed ratio compared with pigs offered diet with 3,000 mg/kg conventional zinc oxide during 217 218 42 to 63 days of age. Long et al. [73] compared the effects of the zinc from nanosized zinc 219 oxide (400 mg/kg), porous zinc oxide (400 mg/kg), and conventional zinc oxide (2,400 mg/kg) 220 in weaning pigs. The results indicated that dietary supplementation with low dose of porous 221 and nanosized zinc oxide had similar effects on improving production performance and

intestinal morphology and reducing diarrhea and intestinal inflammatory as high level of conventional zinc oxide in weaning pigs. In addition, compared with zinc oxide, porous zinc oxide had better effect on reducing diarrhea. Also, no information is published regarding the effects of application of porous zinc oxide in poultry.

226 CONCLUSIONS

Supplementation of pharmacological doses (2,000 to 4,000 mg/kg) of zinc, especially zinc oxide, is an effective method to improve growth performance and alleviate diarrhea postweaning. However, the use of pharmacological doses zinc is criticized due to the large amount of zinc excretion and microbial resistance. To improve the bioavailability of zinc, several novel zinc sources have been developed. However, contradictory effects of those novel zinc sources in animal production have been reported. Therefore, further studies are required to explore the exact mechanisms involved in the antibacterial functions of zinc.

Ű

235 **REFERENCES**

- 1. Jensen J, Kyvsgaard NC, Battisti A, Baptiste KE. Environmental and public health related
- risk of veterinary zinc in pig production-Using Denmark as an example. Environ Int.
 2018;114:81-90.
- 239 2. Todd WR, Elvehjem CA, Hart EB. Zinc in the nutrition of the rat. Am. J. Physiol.
 240 1933;107:146-56.
- Maywald M, Rink L. Zinc homeostasis and immunosenescence. J Trace Elem Med Bio.
 2015;29:24-30.
- 4. Liu Y, Espinosa CD, Abelilla JJ, Casas GA, Lagos LV, Lee SA, et al. Non-antibiotic feed
 additives in diets for pigs. Anim Nutr. 2018;4:113-25.
- 5. Gresse R, Chaucheyras-Durand F, Fleury MA, Van de Wiele T, Forano E, Blanquet-Diot S.
 Gut microbiota dysbiosis in postweaning piglets: understanding the keys to health. Trends
 Microbiol. 2017;25:851-73.
- Bednorz C, Oelgeschläger K, Kinnemann B, Hartmann S, Neumann K, Pieper R, et al. The
 broader context of antibiotic resistance: zinc feed supplementation of piglets increases the
 proportion of multi-resistant Escherichia coli in vivo. Int J Med Microbiol. 2013;303:396 403.
- 7. Vahjen W, Pietruszyńska D, Starke IC, Zentek J. High dietary zinc supplementation
 increases the occurrence of tetracycline and sulfonamide resistance genes in the intestine
 of weaned pigs. Gut pathog. 2015;7:23.
- 8. van Alen S, Kaspar U, Idelevich EA, Köck R, Becker K. Increase of zinc resistance in
 German human derived livestock-associated MRSA between 2000 and 2014. Vet
 Microbiol. 2018;214:7-12.
- 258 9. Park BC, Jung DY, Kang SY, Ko, YH, Ha DM, Kwon CH, et al. Effects of dietary

- supplementation of a zinc oxide product encapsulated with lipid on growth performance,
 intestinal morphology, and digestive enzyme activities in weanling pigs. Anim Feed Sci
 Tech. 2015;200:112-7.
- 10. Milani NC, Sbardella M, Ikeda NY, Arno A, Mascarenhas BC, Miyada VS, Dietary zinc
 oxide nanoparticles as growth promoter for weanling pigs. Anim Feed Sci Tech.
 2017;227:13-23.
- 11. Wang C, Zhang L, Su W, Ying Z, He J, Zhang L, et al. Zinc oxide nanoparticles as a
 substitute for zinc oxide or colistin sulfate: Effects on growth, serum enzymes, zinc
 deposition, intestinal morphology and epithelial barrier in weaned piglets. PloS One.
 2017:12:e0181136.
- 12. Kołodziejczak-Radzimska A, Jesionowski T.Zinc oxide from synthesis to application: a
 review. Materials 2014;7:2833-81.
- 13. Shen J, Chen Y, Wang Z, Zhou A, He M, Mao L, et al. Coated zinc oxide improves
 intestinal immunity function and regulates microbiota composition in weaned piglets. Br J
 Nutr. 2014;111:2123-34.
- 14. Zhao CY, Tan SX, Xiao XY, Qiu XS, Pan JQ, Tang ZX. Effects of dietary zinc oxide
 nanoparticles on growth performance and antioxidative status in broilers. Biol Trace Elem
 Res. 2014;160:361-7.
- 15. Sirelkhatim A, Mahmud S, Seeni A, Kaus NHM, Ann LC, Bakhori SKM, et al. Review on
- zinc oxide nanoparticles: antibacterial activity and toxicity mechanism. Nano-Micro
 Letters. 2015;7:219-42.
- 16. Swain PS, Rao SB, Rajendran D, Dominic G, Selvaraju, S. Nano zinc, an alternative to
 conventional zinc as animal feed supplement: A review. Anim Nutr. 2016;2:134-41.
- 282 17. Vahjen W, Roméo A, Zentek J.I mpact of zinc oxide on the immediate postweaning

- colonization of enterobacteria in pigs. J Anim Sci. 2016;94:359-63.
- 18. Revy PS, Jondreville C, Dourmad JY, Nys Y. Effect of zinc supplemented as either an
 organic or an inorganic source and of microbial phytase on zinc and other minerals
 utilisation by weanling pigs. Anim Feed Sci Tech. 2004;116:93-112.
- 19. Blavi L, Sola-Oriol D, Perez JF, Stein HH. Effects of zinc oxide and microbial phytase on
- digestibility of calcium and phosphorus in maize-based diets fed to growing pigs. J Anim
 Sci. 2017;95:847-54.
- 290 20. Casewell M, Friis C, Marco E, McMullin P, Phillips I. The European ban on growth-
- promoting antibiotics and emerging consequences for human and animal health. J
 Antimicrob Chemoth. 2003;52:159-61.
- 293 21. Maron DF, Smith TJ, Nachman KE. Restrictions on antimicrobial use in food animal
 294 production: an international regulatory and economic survey. Global Health 2013;9:48.
- 295 22. Levy S. Reduced antibiotic use in livestock: how Denmark tackled resistance. Environ
 296 Health Perspect. 2014;122:A160-5.
- 297 23. Dowarah R, Verma AK, Agarwal N, Singh P, Singh BR. Selection and characterization of
- probiotic lactic acid bacteria and its impact on growth, nutrient digestibility, health and
 antioxidant status in weaned piglets. PloS One. 2018;13:e0192978.
- 24. Ou D, Li D, Cao Y, Li X, Yin J, Qiao S, et al. Dietary supplementation with zinc oxide
 decreases expression of the stem cell factor in the small intestine of weanling pigs. J Nutr
- 302 Biochem. 2007;18:820-6.
- 303 25. Yin J, Li X, Li D, Yue T, Fang Q, Ni J, et al. Dietary supplementation with zinc oxide
 304 stimulates ghrelin secretion from the stomach of young pigs. J Nutr Biochem.
 305 2009;20:783-90.
- 306 26. Sargeant HR, McDowall KJ, Miller HM, Shaw MA. Dietary zinc oxide affects the

- expression of genes associated with inflammation: transcriptome analysis in piglets
 challenged with ETEC K88. Vet. Immunol Immunop. 2010;137:120-9.
- 309 27. Hu C, Song J, You Z, Luan Z, Li W. Zinc Oxide-Montmorillonite hybrid influences
- diarrhea, intestinal mucosal integrity, and digestive enzyme activity in weaned pigs. Biol
- 311 Trace Elem Res. 2012;149:190-6.
- 312 28. Walk CL, Wilcock P, Magowan E. Evaluation of the effects of pharmacological zinc oxide
- and phosphorus source on weaned piglet growth performance, plasma minerals and
 mineral digestibility. Animal 2015;9:1145-52.
- 315 29. Han YK, Thacker PA. Effect of antibiotics, zinc oxide and rare earth mineral yeast on
- performance, nutrient digestibility and blood parameters in weaned pigs. Asian-Aust J
 Anim Sci. 2010;23:1057-65.
- 31/ Anim Sci. 2010;25:1057-05.
- 30. Heo JM, Kim JC, Hansen CF, Mullan BP, Hampson DJ, Maribo H, et al. Effects of dietary
 protein level and zinc oxide supplementation on the incidence of post-weaning diarrhoea
 in weaner pigs challenged with an enterotoxigenic strain of *Escherichia coli*. Livest Sci.
- 321 2010;133:210-3.
- 31. Sales J. Effects of pharmacological concentrations of dietary zinc oxide on growth of
 postweaning pigs: A meta-analysis. Biol Trace Elem Res. 2013;152:343-9.
- 324 32. Zhang B. and Guo Y., Supplemental zinc reduced intestinal permeability by enhancing
 325 occludin and zonula occludens protein-1 (ZO-1) expression in weaning piglets. Br J Nutr.
 326 2009;102:687-93.
- 327 33. Pearce SC, Sanz Fernandez MV, Torrison J, Wilson ME, Baumgard LH, Gabler NK.
- Dietary organic zinc attenuates heat stress-induced changes in pig intestinal integrity and
 metabolism. J Anim Sci. 2015;93:4702-13.
- 330 34. Carlson D, Poulsen HD, Sehested J. Influence of weaning and effect of post weaning

- dietary zinc and copper on electrophysiological response to glucose, theophylline and 5-
- HT in piglet small intestinal mucosa. Comp Biochem Physiol. A 2004;137:757-65.
- 333 35. Li XL, Yin JD, Li DF, Chen XJ, Zang JJ, Zhou X. Dietary supplementation with zinc
- oxide increases IGF-I and IGF-I receptor gene expression in the small intestine of
 weanling piglets. J Nutr. 2006;136: 786-91.
- 336 36. Jensen-Waern M, Melin L, Lindberg R, Johanisson A, Peterson L, Wallgren P. Dietary
 337 zinc oxide in weaned pigs-effects on performance, tissue concentrations, morphology,
 338 neutrophil functions and faecal microflora. Res Vet Sci. 1998;64:225-31.
- 339 37. Katouli M, Melin L, Jensen-Waern M, Wallgren P, Mollby R. 1999. The effect of zinc
- oxide supplementation on the stability of the intestinal flora with special reference to
 composition of coliforms in weaned pigs. J Appl Microbiol. 87: 564-73.
- 342 38. Wang, YZ, Xu, ZR, Lin, WX, Huang, HQ, Wang, ZQ. Developmental gene expression
- of antimicrobial peptide PR-39 and effect of zinc oxide on gene regulation of PR-39 in
 piglets. Asian-Australas J Anim Sci. 2004;17:1635-40.
- 345 39. Lichten LA, Cousins RJ. Mammalian zinc transporters: nutritional and physiologic
 346 regulation. Annu Rev Nutr. 2009;29:153-76.
- 40. Brugger D, Windisch WM. Strategies and challenges to increase the precision in feeding
 zinc to monogastric livestock. Anim Nutr. 2017;3:103-8.
- 41. Starke IC, Pieper R, Neumann K, Zentek J, Vahjen W. The impact of high dietary zinc
- 350 oxide on the development of the intestinal microbiota in weaned piglets. FEMS Microbiol.
- 351 Ecol. 2014;87:416-27.
- 42. Yang Q, Ma Y, Zhu J. Applying a novel electrostatic dry powder coating technology to
- 353 pellets. Eur. J. Pharm. Biopharm. 2015;97:118-24.

354	43. Martín MJ, Lara-Villoslada F, Ruiz MA, Morales ME. Microencapsulation of bacteria: A

- review of different technologies and their impact on the probiotic effects. Innov Food Sci
 Emerg Tech. 2015;27:15-25.
- 44. Pan L, Zhao PF, Yang ZY, Long SF, Wang HL, Tian QY, et al. Effects of coated
- compound proteases on apparent total tract digestibility of nutrients and apparent ileal
 digestibility of amino acids for pigs. Asian-Austral J Anim Sci. 2016;29:1761-7.
- 45. Byun YJ, Lee CY, Kim MH, Jung DY, Han JH, Jang I, et al. Effects of dietary
 supplementation of a lipid-coated zinc oxide product on the fecal consistency, growth, and
 morphology of the intestinal mucosa of weanling pigs. J Anim Sci Tech. 2017;59:29.
- 46. Song YM, Kim MH, Kim HN, Jang I, Han JH, Fontamillas GA, et al. Effects of dietary
 supplementation of lipid-coated zinc oxide on intestinal mucosal morphology and
 expression of the genes associated with growth and immune function in weanling pigs.
 Asian-Austral J Anim Sci. 2018;31:403-9.
- 47. Kwon CH, Lee CY, Han SJ, Kim SJ, Park BC, Jang I, et al. Effects of dietary
 supplementation of lipid-encapsulated zinc oxide on colibacillosis, growth and intestinal
 morphology in weaned piglets challenged with enterotoxigenic *Escherichia coli*. Anim Sci
 J. 2014;85:805-13.
- 48. Upadhaya SD, Ki YM, Lee KY, Kim IH. Use of protected zinc oxide in lower doses in
 weaned pigs in substitution for the conventional high dose zinc oxide. Anim Feed Sci Tech.
 2018;240:1-10.
- 49. Hill GM, Mahan DC, Jolliff JS. Comparison of organic and inorganic zinc sources to
 maximize growth and meet the zinc needs of the nursery pig. J Anim Sci. 2014;92: 158294.

377	50. Wang Y, Tang JW, Ma WQ, Feng J. Dietary zinc glycine chelate on growth performance,
378	tissue mineral concentrations, and serum enzyme activity in weanling piglets. Biol Trace
379	Elem Res. 2010;133:325-34.

381 on growth performance and zinc balance in nursery pigs. J Anim Sci. 2002;80:1917-24.

51. Case CL, Carlson MS. Effect of feeding organic and inorganic sources of additional zinc

- 52. Buff CE, Bollinger DW, Ellersieck MR, Brommelsiek WA, Veum TL. Comparison of
 growth performance and zinc absorption, retention, and excretion in weanling pigs fed
 diets supplemented with zinc-polysaccharide or zinc oxide. J Anim Sci. 2005;83:2380-6.
- 53. Cheng J, Kornegay ET, Schell, T. Influence of dietary lysine on the utilization of zinc
 from zinc sulfate and a zinc-lysine complex by young pigs. J Anim Sci. 1998;76:1064-74.
- 54. Payne RL, Bidner TD, Fakler TM, Southern LL. Growth and intestinal morphology of pigs
 from sows fed two zinc sources during gestation and lactation. J Anim Sci. 2006;84:21419.
- 55. Moghaddam HN, Jahanian R. Immunological responses of broiler chicks can be
 modulated by dietary supplementation of zinc-methionine in place of inorganic zinc
 sources. Asian-Aust J Anim Sci. 2009;22:396-403.
- 56. Ao T, Pierce, JL, Pescatore, AJ, Cantor, AH, Dawson, KA, Ford, MJ, et al. Effects of
 organic zinc and phytase supplementation in a maize–soybean meal diet on the
 performance and tissue zinc content of broiler chicks. Br Poult Sci. 2007;48:690-5.
- 396 57. Rossi P, Rutz F, Anciuti MA, Rech JL, Zauk NH. Influence of graded levels of organic
- zinc on growth performance and carcass traits of broilers. J Appl Poult Res. 2007;16:21925.

- 58. Salim HM, Lee HR, Jo C, Lee SK, Lee BD. Effect of sex and dietary organic zinc on
 growth performance, carcass traits, tissue mineral content, and blood parameters of broiler
 chickens. Biol Trace Elem Res. 2012;147:120-9.
- 402 59. Padmavathy N, Vijayaraghavan R. Enhanced bioactivity of ZnO nanoparticles- an
 403 antimicrobial study. Sci Technol Adv Mater. 2008;9:1-7.
- 404 60. Piccinno F, Gottschalk F, Seeger S, Nowack B. Industrial production quantities and uses
 405 of ten engineered nanomaterials for Europe and the world. J Nanopart Res. 2012;14:1109406 20.
- 407 61. Peters RJ, Bouwmeester H, Gottardo S, Amenta V, Arena M, Brandhoff P. et al.
 408 Nanomaterials for products and application in agriculture, feed and food. Trends in Food
 409 Sci. Technol. 2016;54:155-164.
- 410 62. Tsai YH, Mao SY, Li MZ, Huang JT, Lien TF. Effects of nanosize zinc oxide on zinc
- 411 retention, eggshell quality, immune response and serum parameters of aged laying hens.
- 412 Anim. Feed Sci. Tech. 2016;213:99-107.
- 413 63. Mao S, Lien T. Effects of nanosized zinc oxide and γ -polyglutamic acid on eggshell
- 414 quality and serum parameters of aged laying hens. Arch Anim Nutr. 2017;71:373-83.
- 415 64. Li MZ, Huang JT, Tsai YH, Mao SY, Fu CM, Lien TF. Nanosize of zinc oxide and the
- 416 effects on zinc digestibility, growth performances, immune response and serum parameters
- 417 of weanling piglets. Anim Sci J. 2016;87:1379-85.
- 418 65. Wang C, Zhang L, Ying Z, He J, Zhou L, Zhang L, et al. Effects of Dietary Zinc Oxide
- 419 Nanoparticles on Growth, Diarrhea, Mineral Deposition, Intestinal Morphology, and
- 420 Barrier of Weaned Piglets. Biol. Trace. Elem. Res. 2018. doi:
- 421 https://doi.org/10.1007/s12011-018-1266-5.

423	Animal Feed). Scientific opinion on the safety and efficacy of tetra-basic zinc chloride for
424	all animal species. EFSA J. 2012;10:2672.
425	67. Mavromichalis I, Webel DM, Parr EN, Baker DH. Growth-promoting efficacy of
426	pharmacological doses of tetrabasic zinc chloride in diets for nursery pigs. Can J Anim Sci.
427	2001;81:387-91.
428	68. Zhang B, Guo Y.Beneficial effects of tetrabasic zinc chloride for weanling piglets and the
429	bioavailability of zinc in tetrabasic form relative to ZnO. Anim Feed Sci Tech.
430	2007;135:75-85.
431	69. Batal AB, Parr TM, Baker DH. Zinc bioavailability in tetrabasic zinc chloride and the
432	dietary zinc requirement of young chicks fed a soy concentrate diet. Poult Sci. 2001;80:87-
433	90.
434	70. Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets. J
434 435	
	70. Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets. J
435	 Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets. J Anim Sci Biotechnol. 2013;4:19.
435 436	 70. Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets. J Anim Sci Biotechnol. 2013;4:19. 71. Vahjen W, Zentek J, Durosoy S. Inhibitory action of two zinc oxide sources on the ex
435 436 437	 70. Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets. J Anim Sci Biotechnol. 2013;4:19. 71. Vahjen W, Zentek J, Durosoy S. Inhibitory action of two zinc oxide sources on the ex vivo growth of porcine small intestine bacteria. J Anim Sci. 2012;90:334-6.
435 436 437 438	 70. Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets. J Anim Sci Biotechnol. 2013;4:19. 71. Vahjen W, Zentek J, Durosoy S. Inhibitory action of two zinc oxide sources on the ex vivo growth of porcine small intestine bacteria. J Anim Sci. 2012;90:334-6. 72. Morales J, Cordero G, Pineiro C, Durosoy S. Zinc oxide at low supplementation level
435 436 437 438 439	 Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets. J Anim Sci Biotechnol. 2013;4:19. Vahjen W, Zentek J, Durosoy S. Inhibitory action of two zinc oxide sources on the ex vivo growth of porcine small intestine bacteria. J Anim Sci. 2012;90:334-6. Morales J, Cordero G, Pineiro C, Durosoy S. Zinc oxide at low supplementation level improves productive performance and health status of piglets. J Anim Sci. 2012;90:436-8.
435 436 437 438 439 440	 Campbell JM, Crenshaw JD, Polo J. The biological stress of early weaned piglets. J Anim Sci Biotechnol. 2013;4:19. Vahjen W, Zentek J, Durosoy S. Inhibitory action of two zinc oxide sources on the ex vivo growth of porcine small intestine bacteria. J Anim Sci. 2012;90:334-6. Morales J, Cordero G, Pineiro C, Durosoy S. Zinc oxide at low supplementation level improves productive performance and health status of piglets. J Anim Sci. 2012;90:436-8. Long L, Chen J, Zhang Y, Liang X, Ni H, Zhang B, et al. Comparison of porous and nano

66. EFSA FEEDAP Panel (EFSA Panel on Additives and Products or Substances used in