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1 **Abstract**

2 Thirty Hanwoo cattle including bulls, cows, and steers (n = 10 each) were slaughtered and investigated for carcass
3 traits (weight, meat color, fat color, yield index, maturity, marbling score, back-fat thickness, and firmness) and meat
4 quality. The meat quality such as: pH, color, cooking loss, fatty acid, thiobarbituric acid reactive substance, warner-
5 bratzler shear force, tensile tests, and texture profiles were analyzed on *Longissimus Lumborum* (LL) muscles of
6 the carcasses at different aging times (3 d and 21 d). The results showed that steers and cows had higher back-fat
7 thickness and marbling score, and a lower firmness ($p < 0.001$) than bulls. Bulls exhibited a lower meat quality
8 indicating by higher cooking loss, thiobarbituric acid reactive substance content, warner-bratzler shear force and
9 tensile test values ($p < 0.01$). Regarding the sensory property, the bull meat also had higher hardness, and lower
10 tenderness, juiciness and flavor scores than the cow or steer meat ($p < 0.01$). Additionally, the bull meat had a higher
11 polyunsaturated fatty acid and a lower monounsaturated fatty acid contents ($p < 0.01$). With increased aging time,
12 the meat tenderness was improved in all the genders. Taken together, the present study demonstrated that the gender
13 and aging time affected the carcass traits, fatty acid and sensory quality of beef. Postmortem aging could improve
14 the meat tenderness of all genders especially bulls.

15

16 **Keywords** : Ageing, Gender, Texture, Tenderness, Sensory, Quality Traits

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Introduction

21 Hanwoo is a native and valuable cattle breed that is very important in the beef industry sector of Korea [1].
22 Compared to other country's beef (USA, New Zealand, Australia, Canada, and Mexico), Hanwoo beef is
23 characterized by a high intramuscular fat (IMF) and lower content of connective tissues, and unique palatability [2,
24 3]. Hanwoo beef has been regarded as the most expensive and premium meat product in Korea [1].

25 Several studies have found that animals of different genders and ages show different tenderness with particular
26 muscles [4, 5]. Studies on beef indicated that beef from older animals is tougher than beef from younger animals [6].
27 Gender, as a function of sexual hormones, is an important factor influencing the growth pattern, fat, and protein
28 depositions in carcass as a function of sexual hormones [7]. Steer and heifer meat generally have a higher marbling
29 level, so they are tender [8], and better eating quality [9] compared to bull meat. Hence, efforts to improve the eating
30 quality especially tenderness of bull meat is needed.

31 Post-mortem aging could improve palatability attributes such as flavor, odor, flavor intensity and tenderness
32 [10], this process occurs naturally in carcass after slaughtered. During the conversion of muscle to meat, proteins
33 and lipids are broken down into smaller and more flavorful fragments by natural enzymes. Moreover, some of key
34 muscle proteolysis contributes to meat tenderization [11]. Furthermore, during the aging process, oxidative may
35 affect the quality of the meat [12]. Such as: the oxidation of myoglobin turns this pigment into brown metmyoglobin,
36 and lipid oxidation results in forming several products, some of them being associated with the flavor even at low
37 concentrations [13].

38 Wet aging refers to meat aged in a sealed barrier package at refrigerated temperatures. The process occurs in
39 vacuum bags and increased in popularity [14], due to the method is convenient and higher yields compared with
40 dry aging. The wet aging method is easy and needs a short time. Everyone can do the packed package meat aside in
41 their refrigerators and allow them to age. The beef is usually kept for a period of 14d to 42d in wet aging. During the
42 wet aging process, the enzymes will break down the fibers as the beef ages, resulting in a tender cut of meat [14].
43 Thus, until now wet aging is popular with producers due to it takes less time and it has no moisture loss.

44 In addition, beef tenderness evaluated is important for the manufacturer due to tenderness is important to the
45 consumer. Till now, different measurements such as; sensory panel [8, 15, 16], texture profile analysis (TPA), tensile

46 test, and shear force [17] have been used to determine the tenderness of meat. Here, our objective was to evaluate
47 the effects of genders (bull, cow, and steer) on carcass traits, texture and quality characteristics of Hanwoo beef
48 during post-mortem aging (3 d and 21 d).

49

50

Materials and Methods

51 Carcass selection

52 Thirty Hanwoo cattle including bull (n = 10), cow (n = 10), and steer (n = 10) were obtained from the commercial
53 meat processing plant. They were slaughtered at different ages (bull slaughtered average age at 26 months, steer
54 slaughtered average age at 31.2 months, cow slaughtered average age at 46 months). The following day after
55 slaughter, their carcasses were evaluated and graded by an official grader for carcass traits (carcass weight, ribeye
56 area, back fat thickness and yield grade, etc.) according to the beef carcass grading [18].

57 Sample collection

58 *Longissimus Lumborum* (LL) muscles were collected from the left sides of carcasses. Muscles were sealed with
59 vacuum-packaged and aged at 4°C for 3 and 21 days. Each gender contained 10 samples for each aging time. When
60 the aging was completed, they were prepared into sub-sample size depending on analyses. Except for the samples
61 used for the share force, tensile extension, color, and cooking loss, the rests were vacuum-packaged and store at -
62 80°C until use.

63 pH and Color of meat quality parameters

64 The pH values of these samples were determined using a Meat pH meter (HI99163 Hanna instrument, Italy). Each
65 sample was measured 4 times.

66 The color was determined using a Konica Minolta Spectrophotometer (model: CM-2500d), the machine
67 contained an 8-mm measuring port AT, D65 illuminant, and 10° observer (Sinodevices Group, Japan). Each sample
68 was blooming at 4°C for 30 min, then measured at three different locations on the surface. The samples were
69 measured for L*, a*, and b*. L* means the lightness of meat, a* means the redness of meat, and b* means the
70 yellowness of meat.

71 Cooking loss and Objective of meat quality parameters

72 Cooking loss, and texture analysis (WBSF, TPA, and tensile tests) were determined on the samples (3 cm thick steak

73 with the weight of 300 g). Particularly, the samples were immediately placed in plastic bags and cooked in a water
74 bath until their core temperature had increased to 70°C. After cooking the samples cooled in circulating water for 30
75 min. The weight of samples was recorded before and after cooking, then, using the following equation to measure
76 cooking loss:

$$77 \quad \text{Cook loss(\%)} = \frac{\text{weight of uncooked sample(g)} - \text{weight of cooked sample(g)}}{\text{weight of uncooked sample(g)}} * 100$$

78 After cooking loss measurement, the samples were measured for WBSF, TPA, and tensile tests using an Instron
79 Universal Testing Machine (Model 3342; Instron Corporation, Norwood, MA, USA). Every sample was cut into
80 more than 6 trips which are parallel to the muscle fiber direction. The WBSF was evaluated with 1.5 cm diameter
81 samples and sheared at a crosshead speed of 400 mm/min, using a 40 kgf load cell. Tensile testing was conducted
82 with 70 × 10 × 10 mm thick per sample. Stretching was performed at 50 mm/min. TPA (hardness) was done on 3
83 cuts in a rectangular trapezoid shape with 10 × 10 × 40 × 30 mm per sample. Each sample underwent 2 cycles of
84 60% compression at a constant speed of 50 mm/min.

85 **Sensory of meat quality parameters**

86 The sensory evaluation was followed by our lab's previously established protocols [19]. The panel consisted of eight
87 faculty members. Every sample was cut into 4 cm (length) × 3 cm (height) × 0.5 cm (thick) size to be tested. Three
88 sessions were held for every sample at different times. The panelists evaluated contained tenderness, juiciness, flavor,
89 overall acceptability, and overall rating. Each panelist assessed the cooked beef meat samples in a randomized order,
90 and everyone needs to give a value from 1 to 100 (ie: from denotes unacceptable to extremely acceptable) after the
91 test. Each panelist was asked to use distilled water to refresh their mouths in between samples.

92 **Fatty acid and Oxidative stability (TBARS)**

93 A procedure developed by Rule [20] was used to detect the composition of fatty acid. The samples were made into
94 thin slices which were then freeze-dried for 48 h. About 500 mg of each dried sample was placed in a 20 mL vial
95 with 2 mL of 14% boron-trifluoride in methanol and 2 mL of HPLC grade methanol. The vials were sealed with a
96 scrimp cap, then they were placed in a heating block set at 80°C, and vortex mixing every 5 min, for maintained 2 h.
97 Thereafter, 3 mL distilled water and hexane were added, respectively, and followed by centrifuging at 1000 g force
98 for 5 min. Each sample was infused with about 1 mL of hexane and sealed in a vial. The fatty acids composition was
99 determined using an Agilent Gas Chromatography-Mass Spectrometer system (GC-MS) (GC 7890B, MS 5977B

100 Agilent Technologies, USA) with an auto-sampler. The injection temperature was set at 250°C, the carrier gas with a
101 speed 45 cm/s with a split ratio of 50:1. Fatty acid methyl esters were separated with a 1.0 mL/min helium flow
102 which is on a WCOT-fused silica capillary column (30 m × 0.25 mm × 0.25 μm). The oven was programmed as
103 follows: 150°C/2 min, 150°C to 230°C at 10°C/min, 230°C/15 min. The fatty acids were identified by comparison
104 with the retention time with those of fatty acid standards (F.A.M.E. Mix., CRM 18918, 47015-U, Sigma-Aldrich
105 Supelco., USA). The proportion of the fatty acid calculated use the peak area of each identified fatty acid against
106 total identified peak area.

107 The oxidative stability was determined using the procedure developed by Buege and Aust [21], which was
108 detected the values of TBARS (thiobarbituric acid reactive substance) to measure oxidative stability. Briefly, an
109 Ultra Turrax T25 homogenizer (IKA Labortechnik, Jkika Works (Asia) Sdn., Bhd., Malaysia) was used 2.5 g meat
110 samples with the solution for 15 s at 11,000 rpm. The solution contained 7.5 mL DW (distilled water), 25 μL BHA
111 (Butyl hydroxyanisole) and 10 mL TBA/TCA (thiobarbituric acid solution and trichloroacetic acid solution). After
112 homogenizing, the sample was immediately placed in ice, and added TBA/TCA solution to homogenate until the
113 volume to 30 mL. The samples were heated at 90°C in a water bath for 15 min. Then, taken out and placed in ice to
114 cool for 20 min. The absorbance of the sample is determined at 531 nm against a blank that contains all the reagents
115 minus the lipid on an Ultrospec 2000 spectrophotometer (Pharmacia Biotech, Cambridge, England). Multiply the
116 absorbance reading by 5.88 (mg/kg) to calculate the malondialdehyde concentration in the sample.

117 **Statistical Analysis**

118 All data were analyzed using the General Linear Model Procedure of the SAS version 9.3 program (SAS Institute,
119 Cary, NC, USA) [22]. The breed and aging were considered as the fixed factors while the carcass traits, quality
120 attributes, etc. were considered as the variables. Means were compared using Duncan's Multiple Range Test. The
121 significance level was set at $p < 0.05$.

122

123 **Results and Discussion**

124 **Yield and quality grade traits**

125 The carcass traits as affected by gender are presented in Table 1. It was observed that gender affected carcass weight,
126 back fat thickness and, the rib eye area ($p < 0.01$). Cow showed the lowest carcass weight, probably due to the

127 estrous cycle effect and its specific feeding diet. Studies have shown that compared with bulls, steers gain weight at
128 a significantly slower rate and with less efficiency [4, 5]. **In our study, steers and bulls had no difference in carcass**
129 **weight, this could be attributed to the slaughter age difference.** The back-fat thickness was in the following order:
130 steer > cow > bull. The previous study also reported that castrated animals are easier to deposit fat than non-castrates
131 [23]. Bulls had lower back fat thickness compared with cows, this is due to the influence of testicular hormones that
132 cause a significantly higher proportion of lean and a lower proportion of fat [24]. **The yield index was also affected**
133 **by gender ($p < 0.05$), due to yield index, back fat thickness, ribeye area, and carcass weight has the following**
134 **relation: Yield index = $[68.184 - \{0.625 \times \text{back fat thickness (mm)}\}] + \{0.130 \times \text{rib eye area (cm}^2\}) - \{0.024 \times$**
135 **carcass weight (kg)} + 3.23.** Steers had the highest marbling and the lowest firmness meanwhile bulls had the lowest
136 marbling and the highest firmness ($p < 0.001$). This is due to testosterone can inhibit fat development in bulls. **The**
137 **marbling and firmness values showed no difference between steers and cows. This can be attributed to the marbling**
138 **of Hanwoo steers significantly increasing between 12 and 27 months [25], and higher marbling indicating higher**
139 **softness [26].**

140 **Effect of gender and aging on meat quality**

141 Gender showed no influence on color parameters (a^* and b^*) at 3 d aging (Table 2). However, significantly lower L^*
142 and higher pH values were observed in bull meat compared with steers or cow meat (Table 2). **L^* values difference**
143 **can be attributed to the different genders contained different fatness which can influence muscle lightness [27].**
144 **Result of the pH values is in agreement with that color values has a negative correlation with muscle pH values [28],**
145 **meanwhile, Jeremiah et al. [29] found that steers had the lowest ultimate pH values and bulls had the highest**
146 **compared with steers, bulls and heifers.** Regarding aging time, color values increased with increased aging time,
147 especially in bulls and cows ($p < 0.05$). The aging resulted in an increase in lightness, redness, and yellowness
148 values ($p < 0.05$). Previous studies also reported that Bruce et al. [30] and Vitale et al. [31] also showed an increase
149 in L^* , a^* , and b^* values of beef *Longissimus thoracis* after 14d aging. This may be explained due to the higher
150 blooming ability of vacuum-aged meat.

151 **Although, there are no significant differences in cooking loss among genders in Table 2. Bull meat had the**
152 **highest cooking loss (%) (19.5%) compared with steers (16.8%) and cows (16.6%) in both the aging times (3d or**
153 **21d), indicating that the meat of cow and steer had a better water holding capacity. This could be related to the**
154 **chemical composition differences among the beef breeds. The finding is consistent with those of Pogorzelska-**

155 Przybyek et al. [4]. Ozawa et al. [32] reported a lower cooking loss in higher marbling Japanese black steer meat.
156 Cooking loss (%) gradually increased from 3 d to 21 d aging for all genders. Similar results were found in the
157 studies of Boakye and Mittal [33] who showed an increased cooking loss in beef LD (Longissimus dorsi) muscle
158 with increased aging time from day 4 to 16. This may be attributed to an increase in protein denaturation which led
159 to the loss of water holding of the muscle tissues.

160 The TBARS content is often measured and used as an indicator of lipid oxidation levels in foods including
161 meats and meat products [34]. The TBARS concentration is related to the levels of malondialdehyde (MDA) which
162 is the secondary lipid oxidation compound. **TBARS values were significantly influenced by aging time ($p < 0.01$)**
163 **(Table 2)**. At the initial measurement, the meat of the cow showed a lower TBARS content compared to the bulls
164 and steers. This could be related to the fatty acid composition differences among the cattle genders [35]. At 21 d of
165 aging, the TBARS increased in all the samples. ~~Also, the aging caused an increase in the TBARS content.~~ These
166 may be explained by the activity of endogenous or microbiological enzymes. Lipid oxidation produces off-flavors,
167 rancidity, and deterioration in meat and meat products [36]. For example, TBARS of values 2.0 mg MDA/kg were
168 considered to be the lower limit for acceptance of oxidized beef by Campo et al. [37], McKenna et al. [38] adopted
169 1.0 mg MDA/kg as an arbitrary threshold, and Hughes et al. [39] found that TBARS levels between 2.60 and 3.11
170 mg MDA/kg were considered acceptable to consumers in long term aged beef striploin. In our study, the highest
171 TBARS value is 0.45 mg MDA/kg which was much lower than the values reported by these authors.

172 **Effects of gender and aging on texture properties**

173 The mean values of TPA, tensile tests, and WBSF are shown in Table 3. The WBSF, tensile tests, and TPA have been
174 recommended as a tenderness standard by the American Meat Science Association [40] As expected, the WBSF,
175 tensile tests, and TPA values decreased as increasing the aging time in all the samples (Table 3). As increasing the
176 aging time from 3 to 21 d, the bulls, cows, and steers reduced shear force from 4.81, 3.97, and 3.67 kgf to 2.29, 2.08,
177 and 2.26 kgf, respectively. In the same aging time, bulls had the highest WBSF values compared to cows or steers.
178 These differences may be attributed to chemical composition differences such as intermuscular fat (IMF) and
179 subcutaneous fat among the genders [41]. With extending aging time, all the meat samples reduced WBSF values,
180 but the bull meat showed the highest percentage (52.39%) of tenderness improvement compared to the cows
181 (47.61%) and steer meat (38.4%).

182 At the initial measurement, tensile tests values were significantly affected by aging and gender ($p < 0.01$).

183 Although all the tensile tests values showed a decrease with aging time from 3 d to 21 d, bulls still had the highest
184 tensile maximum force values in all the aging times. This can be attributed to adipocytes' excessive development
185 caused disorganization of the perimysial connective tissue [42]. Whilst, tensile strain, tensile extension and tensile
186 maximum force values showed no significant difference among the genders ($p > 0.05$) at 21d aging time. Results
187 can be attributed to the increase myofibrillar fragmentation index with increase aging time [43], and myofibrillar
188 fragmentation index could shows the advancement of myofibrillar degradation.

189 Excepting hardness, all the TPA including, springiness (mm), chewiness (N), hardness (N), and gumminess (N),
190 were not affected by aging. When increasing the aging time up to 21 d, the hardness decreased in all the genders,
191 however, bull meat exhibited the highest hardness values in both aging times. This is consistent with that reported by
192 Lepper-Blilie et al. [44]. Thus, it may be said that aging could improve the tenderness of meat from all cattle genders.

193 **Effects of gender and aging on sensory properties**

194 The effects of gender and aging on sensory attributes such as tenderness, juiciness, flavor, and acceptability are
195 presented in Table 4. There were no differences in the tenderness, juiciness, flavor, and acceptability scores between
196 steer and cow meat. Compared to the meat of these two genders, bull meat had a significant difference for all the
197 sensory traits at 3d aging time ($p < 0.01$). Although, there was no difference for all the sensory traits among the
198 genders at 21d aging time, bull meat had the lowest scores. These differences can be attributed to the IMF content
199 differences (Table 1). Similarly, previous studies have shown that beef with higher marbling is tender, flavorful, and
200 juicier [4, 5, 45]. Extending the aging up to 21 d, no differences in the tenderness, juiciness, and flavor scores
201 occurred among the genders. This means that the eating quality of the meat from all genders was improved during
202 aging. The mechanisms underlying this phenomenon could be related to the breakdown of myofibrils into smaller
203 peptides by the endogenous enzymes, which improved the tenderness and flavor characteristics [46].

204 **Fatty acid profiles**

205 The fatty acid profiles of beef from the three gender are presented in Table 5. The fatty acids composition in muscle
206 tissues play an important role in cooked flavor development [47]. A total of 14 fatty acids (FA) were identified in
207 which the most predominant FAs being oleic acid (C18:1), palmitic acid (C16:0), and stearic acid (C18:0). Our
208 results are consistent with those reported in the previous studies on Hanwoo cattle [48], or American Angus [49],
209 and in Japanese Wagyu [50].

210 Total saturated fatty acids (SFAs) content was similar in all genders. The monounsaturated fatty acids (MUFA)

211 content was lower and the polyunsaturated fatty acids (PUFAs) content was higher in bull meat compared to the
212 other remaining genders ($p < 0.01$). Moreover, bull meat had significantly higher levels of C18:2 and C18:0, and
213 had lower levels of C16:1 and C18:1 than either steers or cows ($p < 0.01$). Our results are in accordance with those
214 of Legako et al. [51] who reported a higher MUFA and lower PUFA content in beef with higher marbling. It has also
215 been reported that the C18:1 and MUFA are positively associated with beef flavor. Thus, the higher flavor score in
216 the steer beef (Table 4) could be due to its higher oleic acid content. Contrastingly, the PUFAs content such as C18:2
217 has been reported to negatively affect the beef flavor [52]. In our study, we found that the C18:2 and MUFA were
218 the highest in the bull beef ($p < 0.01$).

219 **Conclusion**

220 Compared to the cow and steer, bulls had a lower marbling score and back-fat thickness. Regarding meat quality,
221 bull meat had a higher cooking loss, WBSF, and tensile test values compared to those of cows and steers throughout
222 the aging period. Bull meat also exhibited a higher TBARS content during aging. For the sensory quality, the bull
223 meat had lower tenderness, juice, and flavor scores. The meat of steer and cows showed higher C18:1 and MUFA
224 content whereas, the bull meat had higher C18:2 and PUFA content. Aging significantly improved the tenderness of
225 meat from all genders. It may be concluded that gender and aging exhibited a significant effect on carcass and
226 quality of beef, and aging could improve the tenderness of meat from all cattle gender especially bull.

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376

Table 1 Yield and quality traits of Hanwoo beef carcasses subjected to different genders

Quality traits	Bull	Steer	Cow	SEM	F value
Back fat, mm	6.6c	14.6a	11.4b	1.9	8.7***
Rib-eye area, cm ²	80.6b	93.8a	89a	3.6	5.3**
Carcass weight, kg	414a	437a	365b	11.7	11***
Maturity ¹⁾	3b	2c	5a	0.4	23***
Yield index ²⁾	68a	64b	67a	1.2	5.8*
Marbling score ³⁾	1b	5a	4a	0.6	13***
Meat color ⁴⁾	5.2	4.8	5	0.2	2.4
Fat color ⁵⁾	3	3	3	0	-
Firmness ⁶⁾	2a	1.2b	1.4b	0.2	10***
month	26b	31.2b	46a	10.09	18.52***

379 ¹⁾ Maturity: 1 to 9 means the maturity from youthful to mature.

380 ²⁾ Yield index = $[68.184 - \{0.625 \times \text{back fat thickness (mm)}\}] + \{0.130 \times \text{rib eye area (cm}^2\}) - \{0.024 \times \text{carcass}$
 381 $\text{weight (kg)}\} + 3.23$.

382 ³⁾ Marbling score: the values from 1 to 9 indicate the marbling is from devoid to abundant.

383 ⁴⁾ Meat color: the values from 1 to 7 indicate the color is from bright cherry to dark red.

384 ⁵⁾ Fat color score: the values from 1 to 7 indicate the fat color is from white to dark yellow.

385 ⁶⁾ Firmness score: the values from 1 to 3 indicate the meat is from soft to firm.

386 * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$.

387 ^{a, b,} indicate significantly different within row with different superscripts.

Table 2 Quality traits of LL muscle subjected to different aging time and genders

Quality traits	Aging	Bull	Steer	Cow	SEM	F value
pH	3 d	5.50a	5.43b	5.46ab	0.04	6.72**
	21 d	5.51a	5.49ab	5.42b	0.07	2.93
	F value	0.1	2.04	2.87		
CIE L*	3 d	33.1b	37.9a	37.2aY	3.32	4.43*
	21 d	36.4	37.9	39.6X	2.64	2.09
	F value	4.69	0	8.44*		
CIE a*	3 d	16.2Y	17.4Y	17.4Y	1.64	0.83
	21 d	20.0X	20.1X	21.0X	1.66	0.49
	F value	10.07*	6.16*	14.66**		
CIE b*	3 d	11.5Y	13.4	13.0Y	1.48	2.83
	21 d	14.7bX	15.3ab	17.8aX	2.42	3.03
	F value	8.67*	5.26	11.59**		
Cooking loss, %	3 d	19.5	16.8	16.6Y	2.68	2.14
	21 d	22.2	19.0	19.6X	2.93	1.9
	F value	1.76	1.56	7.61*		
TBARS, mg MA/kg	3 d	0.21aY	0.17abY	0.13bY	0.09	3.76
	21 d	0.45aX	0.26bX	0.30bX	0.15	11.37*
	F value	89.42***	5.41*	15.53**		

389 ^{a, b}, indicate significantly different within row with different superscripts.

390 ^{X, Y}, indicate significantly different within column with different superscripts.

391 *** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$.

392

Table 3 Shear force and texture characteristics of LL muscle

Trials	Aging	Bull	Steer	Cow	SEM	F value
WBSF, kgf	3 d	4.81X	3.67	3.97X	1.23	1.17
	21 d	2.29Y	2.26	2.08Y	0.33	0.53
	F value	17.37**	5.3	15.67**		
Tensile maximum force, kgf	3 d	3.34aX	2.25bX	2.85abX	0.62	7.34**
	21 d	1.47aY	1.02abY	0.86bY	0.43	3.67
	F value	27.1***	26.47***	256.94***		
Tensile strain, %	3 d	202.3aX	123.7bX	172.2aX	45.42	7.14**
	21 d	60.8Y	82.1Y	83.1Y	23.14	1.6
	F value	41.86***	8.01*	30.44***		
Tensile extension, mm	3 d	20.6aX	12.6bX	17.4aX	4.62	6.96**
	21 d	6.13Y	8.3Y	8.34Y	2.33	1.6
	F value	41.57***	8.99*	29.42**		
Hardness 1, N	3 d	5.12a	3.45b	4.54ab	1.14	4.06*
	21 d	4.56a	3.3b	3.73b	0.65	7.3**
	F value	0.79	0.22	4.8		
Hardness 2, N	3 d	0.05b	0.06ab	0.066ab	0.02	0.13
	21 d	0.07	0.05	0.08	0.03	1
	F value	0.34	1.38	1.16		
Springiness, mm	3 d	0.97	0.85	0.93	0.17	0.65
	21 d	0.95	0.82	1.14	0.28	1.93
	F value	0.01	0.05	4.94		
Gumminess, N	3 d	0.133a	-0.028b	0.032ab	0.09	4.27*
	21 d	0.07	-0.056	0.012	0.11	1.86
	F value	0.42	0.18	0.19		
Adhesiveness, J	3 d	0.006ab	-0.002aX	0.005ab	0.002	2.56*
	21 d	-0.006	-0.005Y	0.005	0.002	0.6
	F value	0.02	8.1*	0.36		
Chewiness, N*mm	3 d	0.162	0.043	0.062	0.11	1.95
	21 d	0.151	0.059	0.134	0.14	0.56
	F value	0.01	0.04	1.44		

394 *** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$.

395 ^{a, b} indicate significantly different within row with different superscripts.

396 ^{x, y} indicate significantly different within column with different superscripts.

397

Table 4 Sensorial characteristics of LL muscle as affected by aging and gender

Traits	Aging	Bull	Steer	Cow	SEM	F value
Tenderness ¹⁾	3 d	26.4bY	50.4aY	50.4aY	15.67	7.59**
	21 d	80.2X	81.8X	86.6X	7.06	1.13
	F value	273.06***	16.88**	36.18***		
Juiciness ²⁾	3 d	32.6bY	47.8aY	54.4aY	12.66	7.55**
	21 d	75.4X	79.4X	81.8X	7.05	1.06
	F value	174.13***	22.69**	30.4***		
Flavor ³⁾	3 d	34.6bY	59.2aY	60aY	13.62	24.43***
	21 d	72.8X	80.6X	79.6X	6.30	2.88
	F value	165.82***	37.85***	16.38**		
Overall acceptability ⁴⁾	3 d	33.4bY	57.2aY	56.4aY	14.52	9.74***
	21 d	74.6X	81.6X	81.4X	6.25	2.46
	F value	264.4***	14.96**	21.24**		
Overall rating ⁵⁾	3 d	31.4bY	55.8aY	55.4aY	15.28	8.91**
	21 d	73bX	82.4aX	79.8abX	6.87	3.32
	F value	305.75***	13.5**	20.53**		

400 ¹⁾ Tenderness rating: the values from 0 to 100 indicate the tenderness is from not tender to very tender.

401 ²⁾ Juiciness rating: the values from 0 to 100 indicate the juiciness is from not juicy to very juicy.

402 ³⁾ Flavor rating: the values from 0 to 100 indicate the flavor is from dislike to like extremely.

403 ⁴⁾ Overall acceptability: the values from 0 to 100 indicate overall acceptability is from dislike to like extremely.

404 ⁵⁾ Overall rating: the values from 0 to 100 indicate overall rating is from unsatisfactory to satisfactory extremely.

405 ^{a, b,} indicate significantly different within row with different superscripts.

406 ^{X, Y,} indicate significantly different within column with different superscripts.

407 *** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$.

409

410

Table 5 Fatty acid content of LL muscle subjected to different genders

Fatty acid	Bull	Steer	Cow	SEM	F value
C8:0	0.01b	0.02a	0.013ab	0.002	7.2*
C10:0	0.08	0.13	0.11	0.02	2
C12:0	0.17	0.24	0.15	0.03	1.8
C14:0	4.41	5.36	5.4	0.39	1.4
C16:0	19.2	18.4	19.63	0.52	0.98
C16:1	5.91b	9.75a	10.86a	0.68	9.6**
C18:0	19.05a	14.99a	16.63b	0.77	5.1*
C18:1	37.9	42.8	38.95	1.81	1.4
C18:2	11.58a	7.31b	7.07b	0.67	9.6**
C18:3	0.33b	0.27c	0.27c	0.02	7.9**
C20:0	0.29	0.27	0.24	0.03	0.7
C20:0	0.49	0.29	0.3	0.06	2.4
C22:1	0.07		0.03		
C22:4	0.52a	0.16b	0.38ab	0.06	5.5*
SFA	44.24	39.88	42.88	1.39	1.8
MUFA	43.84b	52.53a	49.81a	1.45	6.3**
PUPA	11.91a	7.578b	7.34b	0.68	9.8**

411 * means $p < 0.05$; ** means $p < 0.01$; *** means $p < 0.001$.

412 SFA mean saturated fatty acids.

413 MUFA mean monounsaturated fatty acids.

414 PUFA mean polyunsaturated fatty acids.

415 ^{a, b}, indicate significantly different within row with different superscripts.