JAST (Journal of Animal Science and Technology) TITLE PAGE

ARTICLE INFORMATION	Fill in information in each box below
Article Type	Review article
Article Title (within 20 words without abbreviations)	Identification of lncRNA-mRNA interactions and genome-wide lncRNA annotation in animal transcriptome profiling
Running Title (within 10 words)	IncRNA annotation and IncRNA function in animal transcriptome
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Competing interests	No potential conflict of interest relevant to this article was reported.
Funding sources State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available.	This work was carried out with the support of the Cooperative Research Program for Agriculture Science & Technology Development, of the Rural Development Administration, Republic of Korea (PJ01620403).
Acknowledgements	We would like to thank Joshua S. for English language review.
Availability of data and material	
Authors' contributions Please specify the authors' role using this form.	Conceptualization: Yoon-Been Park and Jun-Mo Kim. Writing - original draft: Yoon-Been Park. Writing - review & editing: Jun-Mo Kim.
Ethics approval and consent to participate	This article does not require IRB/IACUC approval because there are no human and animal experiments.

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1 Abstract

2 Protein-translated mRNA analysis has been extensively used to determine the function of various traits in animals. 3 The non-coding RNA (ncRNA), which was known to be non-functional because it was not encoded as a protein, 4 was re-examined as it was studied to actually function. One of the ncRNAs, long non-coding RNA (lncRNA), is 5 known to have a function of regulating mRNA expression, and its importance is emerging. Therefore, lncRNAs 6 are currently being used to understand the traits of various animals as well as human diseases. However, studies 7 on lncRNA annotation and its functions are still lacking in most animals except humans and mice. lncRNAs have 8 unique characteristics of lncRNAs and interact with mRNA through various mechanisms. In order to make 9 lncRNA annotations in animals in the future, it is essential to understand the characteristics of lncRNAs and the mechanisms by which lncRNAs function. In addition, this will allow lncRNAs to be used for a wider variety of 10 11 traits in a wider range of animals, and it is expected that integrated analysis using other biological information 12 will be possible.

13 Keywords: lncRNA, Animal traits, lncRNA annotation, lncRNA-mRNA interaction, lncRNA function

Introduction

Most of the genes refer to a region of a DNA sequence that functions related to animal traits or diseases. Therefore, gene expression profiling has been used to analyze biological functions (1), and analysis has been conducted by reading RNA sequences from the transcription process of DNA. However, coding RNAs that are translated into protein accounts for only about 4% of RNA, and the fact that non-coding RNAs existing in a vast region, which were treated with no role in the early days, are involved in gene regulation in various ways are being investigated (2).

21 Among them, long non-coding RNA (lncRNA), unlike mRNA, is not translated into a protein, despite its 22 similar sequence structure (3). In a small number of investigations involving animals, plants, and humans, it has 23 been revealed that lncRNA functions in certain diseases or specific environments. It turns out that lncRNAs, 24 previously considered to have no role, play many significant roles, the most important of which is to regulate 25 mRNA expression (4, 5). LncRNAs regulate gene expression in a variety of ways at epigenetic, chromatin 26 remodeling, transcriptional, and translational levels (6). With the development of Next Generation Suquencing, 27 IncRNA identification has been performed in humans and plants but also various species of animals. As the studies 28 progressed, it was found that lncRNA had longitudinal, tissue-specific, and environmental-specific properties, so 29 various case studies began to progress in various animals (4). Prior studies and database construction are 30 insufficient in other animals compared to humans and mice, so efforts are underway to continuously discover 31 lncRNAs with essential functions and to be studied in many livestock animal samples (7-9). However, even after 32 some time since the importance of lncRNA emerged, many lncRNA transcripts have not been identified in 33 livestock animals, or the functions of lncRNAs have not been identified properly. Therefore, lncRNA research is 34 expected to be actively conducted for higher-dimensional bioinformatic analyses and multi-omics integration 35 (MOI).

1. RNA

36

RNA is a polymeric genetic material that plays a vital role in various life phenomena, including control of gene
expression (10, 11). Unlike DNA, RNA is not a pair of double strands but a single-stranded molecule with a short
chain of nucleotides (12). Notably, RNA can be divided into two main categories: messenger RNA (mRNA),
which is coded as protein, and non-cocing RNA (nc RNA), which is not coded (13).

41 **1.1 mRNA**

42 For DNA genetic information to be expressed as a protein, DNA must first be transcribed into RNA, and this RNA 43 transcribed to be translated into protein is called mRNA (14). With the development of sequencing technology, it 44 became possible to examine the transcriptome region and to identify genes representing functions. Therefore, 45 studies on mRNA expression levels under various conditions and bioinformation analysis-related studies using 46 these results are being actively pursued (15-17). In humans, mRNA is mainly used for pharmaceutical and vaccine 47 development by enhancing the understanding of the immune system (18-24). Additionally, various studies in mice 48 are being conducted for use in humans because mice are also very similar in their genes to humans (25-30). 49 Furthermore, it is widely used for various trait studies in livestock animals. Many mRNA-related studies have 50 been conducted mainly for the analysis of animal production traits (31-39) and quality traits (40-44), and they are 51 also used for research in a wide range of areas, such as milk production (45-48), egg production (49-51), nutrients 52 (52-54), stress (55-60), disease (61-64), and reproductive traits (65-71).

53 1.2 NcRNA

NcRNA refers to RNA that is not translated into a protein (72), and there are many different types of ncRNA.
First, ncRNA can be divided into housekeeping and regulatory ncRNA (Figure 1). Housekeeping ncRNAs include
transfer RNA (tRNA), ribosomal RNA (rRNA), small nuclear RNA (snRNA), and small nuclear RNA (snoRNA),
while regulatory ncRNAs include microRNAs (miRNA), small interfering RNA (siRNA), piwi-interacting RNA
(piRNA), and long non-coding RNA (lncRNA) (73).

Housekeeping ncRNAs are essentially expressed and mainly involved in rRNA modification RNA splicing control (74). tRNA has a complementary anticodon in protein synthesis, which carries the amino acid to mRNA (75), and rRNA is an RNA that plays a structural role in ribosome formation and contributes to enzyme activity for protein synthesis (76). Additionally, SnRNA binds with other proteins to form snRNPs, and plays a role in 63 recognizing introns in the splicing process (77). SnoRNA is primarily responsible for chemical transformation,
64 such as rRNA and tRNA (78). Notably, the main difference between the two RNAs is that snRNA influences the
65 alternative splicing of pre-mRNA molecules to determine which sequence should be translated into proteins. In
66 contrast, snoRNA participates in tRNA, rRNA, and mRNA editing and genome imprinting (79).

67 Regulatory ncRNAs can be divided into small RNAs and lncRNAs according to the length of RNA. The 68 small ncRNA includes miRNA, siRNA, piRNA, and the like. MiRNA is an ncRNA composed of about 22 nt and 69 functions in RNA silence and regulation of gene expression after transcription (80). Likewise, SiRNA is an ncRNA 70 composed of approximately 23 nt, which is involved in RNA interference and interferes with gene expression by 71 inhibiting the production of specific proteins (81). The main difference between the two RNAs is that miRNA 72 regulates the expression of several mRNAs, and siRNA inhibits the expression of specific target mRNAs (82). 73 Furthermore, PiRNA consists of about 30 nt, which induces PIWI proteins to cleave the target RNA, promote 74 heterochromatin assembly, methylate DNA, and regulate gene expression (83).

Among regulatory ncRNAs, RNA molecules greater than 200 nt in length are defined as lncRNA (84). Although lncRNAs are very similar in structure to mRNAs, they are not translated into proteins and regulate gene expression through various bases, including epigenetic modification (3). The various lists of annotated lncRNAs based on resemblance to protein-coding mRNAs account for only 0.05–1.12% of cellular RNA, while functional intronic RNAs could constitute as much as 16% (85).

The extensive sequences that do not encode proteins (i.e., the majority of the vast regions of intronic and intergenic sequences) have been regarded as accumulated evolutionary remains arising from the early assembly of genes and/or the insertion of mobile genetic elements. However, as the aforementioned regulatory ncRNAs show, most of these supposedly inert sequences are transcribed and widely employed for gene regulation in cis and *trans* (86).

2. LncRNA

Although the structure of the lncRNA seems similar to that of the mRNA, lncRNA is not coded and exists as a
 ncRNA rather than mRNA.

88 2.1 Generation of lncRNA formation

89 LncRNAs mostly have a cap at the 5' end and a poly(A) tail at the 3' end, presumed to be transcribed similarly to 90 mRNAs (87). LncRNAs are transcribed by RNA Polymerase II (Pol II) and RNA Polymerase III at several loci 91 of the genome, most of which are transcribed by Pol II (88). Due to the lncRNA having a weak internal splicing 92 signal and having a long distance between the 3' splice site and the junction, the lncRNA is spliced more 93 inefficiently than the mRNA (89-91). The nuclear position and fate of lncRNAs appear to be coordinated by 94 various causes, ranging from transcription to nuclear export via sequence motifs in cis and factors in trans (4, 92). 95 Since the arrangement and size of lncRNAs are diverse, it is not well known precisely what biogenesis pathways they are treated. It is also challenging to understand whether ribosome-related lncRNAs are involved by ribosomes 96 97 for translation, so further research is needed (4).

98 2.2 Genomic characteristic of IncRNA

Unlike small ncRNAs such as siRNAs, miRNAs, and piRNAs, lncRNAs are relatively long and therefore have
poorly conserved properties (93). Compared to mRNA, lncRNA has a shorter transcript length and a smaller
number of exons on average, and many studies have demonstrated these characteristics (94-96). Furthermore,
lncRNA has a shorter open reading frame (ORF) length than mRNA and a relatively low expression level (89, 97,
98).

104 Also, lncRNA can exist at various locations in the genome (Figure 2) (99). The lncRNA can be present in 105 the intron region between exon and exon and in the intergenic region between the protein-coding gene (PCG) and 106 PCG (100). In particular, lncRNAs present in the intergenic region are named long intergenic non-coding RNA 107 (lincRNA). Additionally, because lincRNA does not overlap with PCG domains or other small RNA genes, it is 108 relatively easy to conduct research such as the structure and function of lincRNA (101). Although lincRNAs are 109 similar in many respects to lncRNAs, they are somewhat longer than lncRNAs and are characterized by their 110 presence in mammalian nuclei (101, 102). Furthermore, LncRNA can also exist in an exonic region where the 111 lncRNA transcript overlaps the exon portion of the PCG (103). Notably, there are also antisense lncRNAs characterized by transcription from opposite strands of PCG (104), which regulate the expression of theirendogenous sense genes (105).

IncRNA has well-known tissue-specific, species-specific, and conditional-specific tendencies. Even the same individual can express lncRNA differently depending on what kind of tissue it is, and even the same tissue can express differently depending on the species (96, 106-110). Moreso, the tissue-specific characteristics of lncRNA demonstrated higher results even when compared to mRNA through the tissue specificity index calculated numerically in mammals (96, 111).

119 2.3 Whole genomic of IncRNA identification

120 Most animals, except humans and mice, do not yet have a well-established lncRNA database, so the process of 121 identifying novel lncRNA identification for lncRNA analysis should be conducted. Therefore, a new Gene transfer format (GTF) file is needed to find the novel lncRNA instead of the reference gtf file of the animal containing 122 123 only the information of previously known RNA. A merged GTF file is generated based on the transcripts of the samples to be analyzed and the reference GTF file of the corresponding animal (7, 108, 112). Following the 124 merged GTF file, only transcripts longer than 200 bp and an open reading frame transcript length shorter than 300 125 126 bp are selected. It also considers the positional relationship in the genome between lncRNA and PCG and 127 designates transcripts consistent with the definition of lncRNA (intergenic, intronic, etc.). Subsequently, it filters 128 only transcripts with low probability using various tools that calculate the potential for transcripts to be coded as proteins. Importantly, tools for evaluating coding potential are diverse and can be used flexibly depending on how 129 130 to analyze. The transcripts filtered from the sequencing data can be selected and presented as potential novel 131 lncRNA, and can be used for functional analysis and actual lncRNA sequence verification in the future (113-119).

3. Interaction IncRNA to mRNA

Among the many known lncRNA functions, a representative and key function is to regulate mRNA expression including , such as epigenetic modification (3, 4). Therefore, a method of conducting mRNA and lncRNA analysis is actively used as an experimental design for exploring animal traits. Importantly, it is used in a wide variety of fields, including production traits (120-123) and quality traits (124-127), milk production (128-130), egg production (9, 131, 132), stress (133-135), diseases (136) and reproductive traits (137-140).

138 The types of lncRNA that regulate transcription can be divided into two based on the transcription site and 139 functional location of the lncRNA. It is classified as cis-acting lncRNA if its functional location depends on the 140 transcription site, and trans-acting lncRNA if transcribed to exert functions elsewhere without relying on the 141 transcription site (Figure 3) (141). Notably, a method for obtaining a candidate target gene for cis- and trans-142 acting lncRNA has not yet been fully established. However, the candidate target gene interacting with *cis*-acting 143 lncRNA is primarily a candidate group of PCGs within 100 kb on the same chromosome of lncRNA. In contrast, the candidate gene interacting with trans-acting lncRNA is a candidate group of PCGs on different chromosomes 144 145 (141, 142).

146 **3.1** *Cis*-acting

As mentioned earlier, the *cis*-acting mechanism is preferred because lncRNA is less likely to function normally due to dilution from diffusion and transport to other cellular compartments. After all, the expression level is generally relatively low (141).

150 The cis-acting lncRNA can increase or inhibit the expression of target genes through various mechanisms. 151 The mechanism by which *cis*-acting lncRNA increases gene expression is closely related to enhancers. These 152 lncRNAs can be broadly divided into two categories: 1) lncRNAs derived from and transcribed from the enhancer 153 after mutation or translocation has occurred in the gene enhancer (143, 144), and 2) those transcribed from other 154 sources that act like the enhancer of the target gene or affect the enhancer (145, 146). These both lncRNAs can 155 activate the target gene by influencing the target gene's enhancer or act as the enhancer and activate the target 156 gene. As a first mechanism, lncRNA transcripts regulate enhancer activity by forming or maintaining chromatin 157 loops with target genes (147, 148). Additionally, since the lncRNA transcript affects the nuclear localization of 158 the enhancer, it can increase the expression of the target gene by giving strength to the enhancer as an indirect

mechanism (149). The *cis*-acting lncRNA can activate the expression of a target gene by influencing the enhancer through mechanisms other than spatial interaction. It is an lncRNA that attracts a protein that enhances the enhancer of the target gene (150-152). There are also *cis*-acting lncRNAs that activate gene expression independent of enhancers. The lncRNA is transcribed near the target gene, or the preformed chromatin loop structure locates the lncRNA near the target gene, thereby increasing the expression of the target gene by attracting activating factors to the lncRNA (153).

165 The cis-acting lncRNA not only increases the expression of a target gene but also inhibits it. First, lncRNA 166 near the target gene can suppress the expression by silencing the target gene's promoter through the enhancer 167 competition of the target gene (154, 155). In addition, the lncRNA is transcribed near the target gene, or the 168 preformed chromatin loop structure places the lncRNA near the target gene so that the lncRNA attracts repressive 169 complexes such as Polycomb repressive complex 2, resulting in the same effect as histone modification. Thus, 170 gene expression can be inhibited (156). Another mechanism by which cis-acting lncRNA suppresses gene 171 expression is transcriptional interference. Through nucleosome remodeling, in which nucleosomes are rearranged, 172 nucleosome occupancy is reduced, or multiple epigenetic modifications, lncRNA that approaches or overlaps the 173 target gene suppresses the expression of the target gene (157, 158).

Previous studies have revealed that *cis*-acting lncRNA does not only interact one-to-one with the target gene. One lncRNA may be involved in the transcription of several target genes, and conversely, it appears that several lncRNAs may be involved in transcribing a target gene in unison. (4, 141).

177 **3.2** *Trans*-acting

Unlike cis-acting lncRNAs, trans-acting lncRNAs can interact independently of complementary sequences for target gene regions (99). *Trans*-acting lncRNAs function by binding to proteins, DNA, and other RNAs (159). First, *trans*-acting lncRNAs can act as post-transcriptional regulatory factors by interacting with RNA-binding proteins (RBPs). These lncRNAs interact with RBPs to inhibit mRNA splicing and the stability and translation of mRNAs (160-162). Notably, splicing regulation by lncRNA causes a mutation or transformation in the splicing regulation sequence of the target pre-mRNA, resulting in the mis-splicing of the mRNA (163).

184 *Trans*-acting lncRNAs can also promote or inhibit the stability of mRNA by interacting directly with RNA 185 through base pairing. This is likely due to the ability to attract proteins involved in mRNA decomposition by 186 directly base pairing with other RNAs (164, 165). While its existence has been revealed and its importance as a

- 187 post-transcriptional control factor has emerged, research on *trans*-acting lncRNA is insufficient. Further research
- 188 will be needed to clarify the apparent correlation between *trans*-acting lncRNA and target genes and reveal the
- 189 mechanisms by which several trans-acting lncRNAs interact with RBP.

4. Another function of lncRNA

191 Until recently, interactions between ncRNAs have rarely been studied. However, recent studies have shown that 192 lncRNA can interact with miRNA and mRNA (166). Importantly, the lncRNA acts as a sponge to attract miRNA 193 and competes with PCG, which was supposed to bind to miRNA. This attraction process reduces the target gene 194 regulation effect of miRNA (167-170). Therefore, studies on high-dimensional access to specific biological 195 information are being conducted by analyzing the correlation and interaction of 3 RNAs of lncRNA-miRNA-196 mRNA (171-173). It has also been suggested that some lncRNAs can be preferentially post-processed into 197 snoRNA (99, 174, 175). As mentioned earlier, the possibility of interaction between lncRNAs or other RNAs is 198 still open, such as various lncRNAs involved in regulating one mRNA expression. However, further research is 199 needed as it is unclear. If these mechanisms are revealed, not only will we be able to understand the principles of 200 IncRNA and mRNA interaction that have not yet been accurately identified, but we will also be able to make much 201 more expansive use of MOI network research using lncRNA.



202 **Conclusions**

203 In the past, only studies on mRNA encoded by functional genes were conducted, but now the role of ncRNAs has 204 been re-examined, and research on this topic is being actively conducted. Among them, lncRNA has a high 205 probability of being present in many different places on the genome, and it is known that it has many functions. 206 Therefore, its importance is emerging from these added investigations. As a key function of lncRNA, it can 207 regulate gene expression through various mechanisms. In addition, since it has tissue-specific and species-specific 208 characteristics, it is possible to analyze bioinformation using lncRNA from multiple perspectives in particular 209 tissues of different species. This means that lncRNAs can be used as biomarkers involved in improving 210 reproductive traits and diseases in mammals, including livestock animals. Therefore, lncRNA exploration and functional analysis are being conducted to study various animal traits. However, analysis for identifying lncRNA 211 212 in animal species other than humans and mice is still lacking, and analysis of the mechanism and function of 213 IncRNA is insufficient. If studies that can supplement these areas are conducted, it is likely that high-dimensional 214 MOI analysis using lncRNA will be possible.



Reference 215 Kim K, Zakharkin SO, Allison DB. Expectations, validity, and reality in gene expression profiling. Journal 216 1. of clinical epidemiology. 2010;63(9):950-9. 217 218 2. Patil VS, Zhou R, Rana TM. Gene regulation by non-coding RNAs. Critical reviews in biochemistry and molecular biology. 2014;49(1):16-32. 219 220 Zhang X, Wang W, Zhu W, Dong J, Cheng Y, Yin Z, et al. Mechanisms and functions of long non-coding 3. 221 RNAs at multiple regulatory levels. International journal of molecular sciences. 2019;20(22):5573. 222 4. Statello L, Guo C-J, Chen L-L, Huarte M. Gene regulation by long non-coding RNAs and its biological 223 functions. Nature reviews Molecular cell biology. 2021;22(2):96-118. 224 Im JH, Muschel RJ. New evidence of lncRNA role in tumor progression and metastasis. Hepatobiliary 5. 225 Surgery and Nutrition. 2012;1(1):55-6. Wang W, Min L, Oiu X, Wu X, Liu C, Ma J, et al. Biological function of long non-coding RNA (LncRNA) 226 6. 227 Xist. Frontiers in cell and developmental biology. 2021;9:645647. Chen L, Shi G, Chen G, Li J, Li M, Zou C, et al. Transcriptome analysis suggests the roles of long intergenic 7. 228 229 non-coding RNAs in the growth performance of weaned piglets. Frontiers in genetics. 2019;10:196. 230 Jia L, Wang J, Luoreng Z, Wang X, Wei D, Yang J, et al. Progress in Expression Pattern and Molecular 8. 231 Regulation Mechanism of LncRNA in Bovine Mastitis. Animals. 2022;12(9):1059. 232 Adetula AA, Gu L, Nwafor CC, Du X, Zhao S, Li S. Transcriptome sequencing reveals key potential long 9. non-coding RNAs related to duration of fertility trait in the uterovaginal junction of egg-laying hens. 233 234 Scientific reports. 2018;8(1):1-12.

- 10. Joyce GF. The antiquity of RNA-based evolution. nature. 2002;418(6894):214-21.
- Arraiano CM, Andrade JM, Domingues S, Guinote IB, Malecki M, Matos RG, et al. The critical role of RNA
 processing and degradation in the control of gene expression. FEMS microbiology reviews. 2010;34(5):883 923.
- 239 12. Seetin MG, Mathews DH. RNA structure prediction: an overview of methods. Bacterial regulatory RNA.
 240 2012:99-122.
- Brandenburger T, Somoza AS, Devaux Y, Lorenzen JM. Noncoding RNAs in acute kidney injury. Kidney
 International. 2018;94(5):870-81.
- 243 14. Clancy S, Brown W. Translation: DNA to mRNA to. Nature Education. 2008.

- 244 15. de Sousa Abreu R, Penalva LO, Marcotte EM, Vogel C. Global signatures of protein and mRNA expression
 245 levels. Molecular BioSystems. 2009;5(12):1512-26.
- If a Greenbaum D, Colangelo C, Williams K, Gerstein M. Comparing protein abundance and mRNA expression
 levels on a genomic scale. Genome biology. 2003;4(9):1-8.
- 248 17. Chen G, Gharib TG, Huang C-C, Taylor JM, Misek DE, Kardia SL, et al. Discordant protein and mRNA
 249 expression in lung adenocarcinomas. Molecular & cellular proteomics. 2002;1(4):304-13.
- 18. Yakubov E, Rechavi G, Rozenblatt S, Givol D. Reprogramming of human fibroblasts to pluripotent stem
 cells using mRNA of four transcription factors. Biochemical and biophysical research communications.
 2010;394(1):189-93.
- Anderson MG, Nakane M, Ruan X, Kroeger PE, Wu-Wong JR. Expression of VDR and CYP24A1 mRNA
 in human tumors. Cancer chemotherapy and pharmacology. 2006;57(2):234-40.
- 20. Wang X, Reape TJ, Li X, Rayner K, Webb CL, Burnand KG, et al. Induced expression of adipophilin mRNA
 in human macrophages stimulated with oxidized low-density lipoprotein and in atherosclerotic lesions. FEBS
 letters. 1999;462(1-2):145-50.
- 258 21. Dolgin E. The tangled history of mRNA vaccines. Nature. 2021;597(7876):318-24.
- Fujii T, Yamada S, Watanabe Y, Misawa H, Tajima S, Fujimoto K, et al. Induction of choline acetyltransferase
 mRNA in human mononuclear leukocytes stimulated by phytohemagglutinin, a T-cell activator. Journal of
 neuroimmunology. 1998;82(1):101-7.
- 262 23. Faustino NA, Cooper TA. Pre-mRNA splicing and human disease. Genes & development. 2003;17(4):419 263 37.
- 264 24. Hollams EM, Giles KM, Thomson AM, Leedman PJ. MRNA stability and the control of gene expression:
 265 implications for human disease. Neurochemical research. 2002;27(10):957-80.
- 266 25. Venter JC, Adams MD, Myers EW, Li PW, Mural RJ, Sutton GG, et al. The sequence of the human genome.
 267 science. 2001;291(5507):1304-51.
- 268 26. Lee SF, Newton C, Widen R, Friedman H, Klein TW. Differential expression of cannabinoid CB2 receptor
 269 mRNA in mouse immune cell subpopulations and following B cell stimulation. European journal of
 270 pharmacology. 2001;423(2-3):235-41.
- 27. Hörtnagl H, Tasan R, Wieselthaler A, Kirchmair E, Sieghart W, Sperk G. Patterns of mRNA and protein
 expression for 12 GABAA receptor subunits in the mouse brain. Neuroscience. 2013;236:345-72.

 273 28. Carrillo-Vico A, Garcia-Perganeda A, Naji L, Calvo J, Romero M, Guerrero J. Expression of membrane and 274 nuclear melatonin receptor mRNA and protein in the mouse immune system. Cellular and Molecular Life 275 Sciences CMLS. 2003;60(10):2272-8.

- 276 29. Morgan ME, van Bilsen JH, Bakker AM, Heemskerk B, Schilham MW, Hartgers FC, et al. Expression of FOXP3 mRNA is not confined to CD4+ CD25+ T regulatory cells in humans. Human immunology. 2005;66(1):13-20.
- 279 30. Li Y, South T, Han M, Chen J, Wang R, Huang X-F. High-fat diet decreases tyrosine hydroxylase mRNA
 280 expression irrespective of obesity susceptibility in mice. Brain research. 2009;1268:181-9.
- Buzala M, Janicki B. Effects of different growth rates in broiler breeder and layer hens on some productive
 traits. Poultry Science. 2016;95(9):2151-9.
- Shen H, Zhao S, Cao J, Li X, Fan B. Porcine MuRF2 and MuRF3: molecular cloning, expression and association analysis with muscle production traits. Molecular biology reports. 2011;38(8):5115-23.
- Juszczuk-Kubiak E, Bujko K, Grześ M, Cymer M, Wicińska K, Szostak A, et al. Study of bovine Mef2B
 gene: the temporal-spatial expression patterns, polymorphism and association analysis with meat production
 traits. Journal of animal science. 2016;94(11):4536-48.
- Zeng F, Xie L, Pang X, Liu W, Nie Q, Zhang X. Complementary deoxyribonucleic acid cloning of avian
 G0/G1 switch gene 2, and its expression and association with production traits in chicken. Poultry science.
 2011;90(7):1548-54.
- 35. Szreder T, Zwierzchowski L. Estrogen receptors and their genes—potential markers of functional and production traits of farm animals. Molecular biology reports. 2007;34(4):207-11.
- Bai Y, Sun G, Kang X, Han R, Tian Y, Li H, et al. Polymorphisms of the pro-opiomelanocortin and agouti related protein genes and their association with chicken production traits. Molecular biology reports.
 2012;39(7):7533-9.
- 296 37. Cochran SD, Cole JB, Null DJ, Hansen PJ. Discovery of single nucleotide polymorphisms in candidate genes
 297 associated with fertility and production traits in Holstein cattle. BMC genetics. 2013;14(1):1-23.
- 298 38. Liu L, Dou T, Li Q, Rong H, Tong H, Xu Z, et al. Myostatin mRNA expression and its association with body
 299 weight and carcass traits in Yunnan Wuding chicken. Genet Mol Res. 2016;15(10.4238).
- 300 39. Ramayo-Caldas Y, Ballester M, Sánchez JP, González-Rodríguez O, Revilla M, Reyer H, et al. Integrative
 approach using liver and duodenum RNA-Seq data identifies candidate genes and pathways associated with
 feed efficiency in pigs. Scientific reports. 2018;8(1):1-11.
- 40. Lee E, Kim J, Lim K, Ryu Y, Jeon W, Hong K. Effects of variation in porcine MYOD1 gene on muscle fiber
 characteristics, lean meat production, and meat quality traits. Meat science. 2012;92(1):36-43.
- 41. Çinar MU, Fan H, Neuhoff C, Groβe-Brinkhaus C. eQTL Analysis and association of MYF6 mRNA
 expression with meat quality traits in pigs. Kafkas Universitesi Veteriner Fakultesi Dergisi. 2012;18(2).
- 42. Nkrumah J, Li C, Basarab J, Guercio S, Meng Y, Murdoch B, et al. Association of a single nucleotide

- polymorphism in the bovine leptin gene with feed intake, feed efficiency, growth, feeding behaviour, carcass
 quality and body composition. Canadian Journal of Animal Science. 2004;84(2):211-9.
- 43. Li X, Kim S-W, Choi J-S, Lee Y-M, Lee C-K, Choi B-H, et al. Investigation of porcine FABP3 and LEPR
 gene polymorphisms and mRNA expression for variation in intramuscular fat content. Molecular biology
 reports. 2010;37(8):3931-9.
- 44. Seo Y-J, Lim B, Kim D-Y, Lim K-S, Kim J-M. Regulation of Swine Growth by Backfat Tissue during
 Growing and Finishing Stages. Animals. 2021;11(12):3511.
- Raliou M, Dembélé D, Düvel A, Bolifraud P, Aubert J, Mary-Huard T, et al. Subclinical endometritis in dairy
 cattle is associated with distinct mRNA expression patterns in blood and endometrium. PloS one.
 2019;14(8):e0220244.
- Weikard R, Goldammer T, Brunner RM, Kuehn C. Tissue-specific mRNA expression patterns reveal a coordinated metabolic response associated with genetic selection for milk production in cows. Physiological genomics. 2012;44(14):728-39.
- 47. Szewczuk M, Zych S, Czerniawska-Piątkowska E, Wojcik J. Association between IGF1R/i16/TaqI and
 IGF1/SnaBI polymorphisms and milk production traits in Polish Holstein-Friesian cows. Animal Science
 Papers & Reports. 2012;30(1).
- 48. Yang S-H, Bi X-J, Xie Y, Li C, Zhang S-L, Zhang Q, et al. Validation of PDE9A gene identified in GWAS
 showing strong association with milk production traits in Chinese Holstein. International journal of molecular
 sciences. 2015;16(11):26530-42.
- 49. Guo S, Bai Y, Zhang Q, Zhang H, Fan Y, Han H, et al. Associations of CALM1 and DRD1 polymorphisms,
 and their expression levels, with Taihang chicken egg-production traits. Animal Biotechnology. 2021:1-11.
- 50. Yuan Z, Chen Y, Chen Q, Guo M, Kang L, Zhu G, et al. Characterization of chicken MMP13 expression and
 genetic effect on egg production traits of its promoter polymorphisms. G3: Genes, Genomes, Genetics.
 2016;6(5):1305-12.
- 51. Xu H, Shen X, Zhou M, Fang M, Zeng H, Nie Q, et al. The genetic effects of the dopamine D1 receptor gene on chicken egg production and broodiness traits. BMC genetics. 2010;11(1):1-10.
- Vonnahme K, Zhu M, Borowicz P, Geary T, Hess B, Reynolds L, et al. Effect of early gestational
 undernutrition on angiogenic factor expression and vascularity in the bovine placentome. Journal of animal
 science. 2007;85(10):2464-72.
- Wang L, Zhu F, Yang H, Li J, Li Y, Ding X, et al. Effects of dietary supplementation with epidermal growth
 factor on nutrient digestibility, intestinal development and expression of nutrient transporters in early-weaned
 piglets. Journal of animal physiology and animal nutrition. 2019;103(2):618-25.
- Weller M, Albino RL, Marcondes M, Silva W, Daniels K, Campos M, et al. Effects of nutrient intake level on mammary parenchyma growth and gene expression in crossbred (Holstein× Gyr) prepubertal heifers.

- 342 Journal of Dairy Science. 2016;99(12):9962-73.
- 55. Rhoads M, Kim J, Collier R, Crooker B, Boisclair Y, Baumgard L, et al. Effects of heat stress and nutrition
 on lactating Holstein cows: II. Aspects of hepatic growth hormone responsiveness. Journal of Dairy Science.
 2010;93(1):170-9.
- 56. Deb R, Sajjanar B, Singh U, Kumar S, Brahmane M, Singh R, et al. Promoter variants at AP2 box region of
 Hsp70. 1 affect thermal stress response and milk production traits in Frieswal cross bred cattle. Gene.
 2013;532(2):230-5.
- 57. Uematsu M, Ohara Y, Navas JP, Nishida K, Murphy T, Alexander RW, et al. Regulation of endothelial cell nitric oxide synthase mRNA expression by shear stress. American Journal of Physiology-Cell Physiology. 1995;269(6):C1371-C8.
- 58. Do Amaral B, Connor E, Tao S, Hayen J, Bubolz J, Dahl G. Heat stress abatement during the dry period influences prolactin signaling in lymphocytes. Domestic animal endocrinology. 2010;38(1):38-45.
- Sejian V, Bhatta R, Gaughan J, Dunshea F, Lacetera N. Adaptation of animals to heat stress. Animal.
 2018;12(s2):s431-s44.
- Lim C, Lim B, Kil D, Kim J. Hepatic transcriptome profiling according to growth rate reveals acclimation
 in metabolic regulatory mechanisms to cyclic heat stress in broiler chickens. Poultry Science.
 2022;101(12):102167.
- Park J, Kim M, Na G, Jeon I, Kwon Y-k, Kim J-h, et al. Glucocorticoids modulate NF-κB-dependent gene
 expression by up-regulating FKBP51 expression in Newcastle disease virus-infected chickens. Molecular
 and cellular endocrinology. 2007;278(1-2):7-17.
- 362 62. Ishikawa H, Rahman MM, Yamauchi M, Takashima S, Wakihara Y, Kamatari YO, et al. mRNA profile in milk extracellular vesicles from bovine leukemia virus-infected cattle. Viruses. 2020;12(6):669.
- Hoffmann B, Beer M, Reid SM, Mertens P, Oura CA, van Rijn PA, et al. A review of RT-PCR technologies
 used in veterinary virology and disease control: sensitive and specific diagnosis of five livestock diseases
 notifiable to the World Organisation for Animal Health. Veterinary microbiology. 2009;139(1-2):1-23.
- Lim B, Kim S, Lim K-S, Jeong C-G, Kim S-C, Lee S-M, et al. Integrated time-serial transcriptome networks
 reveal common innate and tissue-specific adaptive immune responses to PRRSV infection. Veterinary
 research. 2020;51(1):1-18.
- Fu Y, Fu J, Wang A. Association of EphA4 polymorphism with swine reproductive traits and mRNA
 expression of EphA4 during embryo implantation. Molecular biology reports. 2012;39(3):2689-96.
- 66. Liu S, Yin H, Li C, Qin C, Cai W, Cao M, et al. Genetic effects of PDGFRB and MARCH1 identified in
 GWAS revealing strong associations with semen production traits in Chinese Holstein bulls. BMC genetics.
 2017;18(1):1-10.

- Hui Y, Zhang Y, Wang K, Pan C, Chen H, Qu L, et al. Goat DNMT3B: An indel mutation detection, association analysis with litter size and mRNA expression in gonads. Theriogenology. 2020;147:108-15.
- Kwon SG, Hwang JH, Park DH, Kim TW, Kang DG, Kang KH, et al. Identification of differentially
 expressed genes associated with litter size in Berkshire pig placenta. PloS one. 2016;11(4):e0153311.
- Fernandez-Rodriguez A, Munoz M, Fernandez A, Pena RN, Tomas A, Noguera JL, et al. Differential gene
 expression in ovaries of pregnant pigs with high and low prolificacy levels and identification of candidate
 genes for litter size. Biology of reproduction. 2011;84(2):299-307.
- Chen X, Li A, Chen W, Wei J, Fu J, Wang A. Differential gene expression in uterine endometrium during
 implantation in pigs. Biology of reproduction. 2015;92(2):52, 1-14.
- Kim J-M, Park J-E, Yoo I, Han J, Kim N, Lim W-J, et al. Integrated transcriptomes throughout swine oestrous
 cycle reveal dynamic changes in reproductive tissues interacting networks. Scientific reports. 2018;8(1):1 14.
- 387 72. Hüttenhofer A, Vogel J. Experimental approaches to identify non-coding RNAs. Nucleic acids research.
 388 2006;34(2):635-46.
- Zhang P, Wu W, Chen Q, Chen M. Non-coding RNAs and their integrated networks. Journal of integrative bioinformatics. 2019;16(3).
- 391 74. Morey C, Avner P. Employment opportunities for non-coding RNAs. FEBS letters. 2004;567(1):27-34.
- 392 75. McClain WH. Rules that govern tRNA identity in protein synthesis. Journal of molecular biology.
 393 1993;234(2):257-80.
- Weiner AM, Maizels N. tRNA-like structures tag the 3'ends of genomic RNA molecules for replication:
 implications for the origin of protein synthesis. Proceedings of the National Academy of Sciences.
 1987;84(21):7383-7.
- Lindsay MA, Griffiths-Jones S, Valadkhan S, Gunawardane LS. Role of small nuclear RNAs in eukaryotic
 gene expression. Essays in biochemistry. 2013;54:79-90.
- 399 78. Dieci G, Preti M, Montanini B. Eukaryotic snoRNAs: a paradigm for gene expression flexibility. Genomics.
 2009;94(2):83-8.
- 401 79. Ghoshal K, Jacob ST. An alternative molecular mechanism of action of 5-fluorouracil, a potent anticancer
 402 drug. Biochemical pharmacology. 1997;53(11):1569-75.
- 403 80. Catalanotto C, Cogoni C, Zardo G. MicroRNA in control of gene expression: an overview of nuclear functions. International journal of molecular sciences. 2016;17(10):1712.

- 405 81. Xu W, Jiang X, Huang L. RNA interference technology. Comprehensive Biotechnology. 2019:560.
- 406 82. Lam JK, Chow MY, Zhang Y, Leung SW. siRNA versus miRNA as therapeutics for gene silencing. Molecular 407 Therapy-Nucleic Acids. 2015;4:e252.
- 408
 408 83. Ozata DM, Gainetdinov I, Zoch A, O'Carroll D, Zamore PD. PIWI-interacting RNAs: small RNAs with big functions. Nature Reviews Genetics. 2019;20(2):89-108.
- 410
 41. Zheng H, Brennan K, Hernaez M, Gevaert O. Benchmark of long non-coding RNA quantification for RNA sequencing of cancer samples. GigaScience. 2019;8(12):giz145.
- 412 85. Laurent GS, Wahlestedt C, Kapranov P. The Landscape of long noncoding RNA classification. Trends in genetics. 2015;31(5):239-51.
- 414 86. Mattick JS, Makunin IV. Non-coding RNA. Human molecular genetics. 2006;15(suppl_1):R17-R29.
- 415 87. Quinn JJ, Chang HY. Unique features of long non-coding RNA biogenesis and function. Nature Reviews
 416 Genetics. 2016;17(1):47-62.
- 88. Schlackow M, Nojima T, Gomes T, Dhir A, Carmo-Fonseca M, Proudfoot NJ. Distinctive patterns of transcription and RNA processing for human lincRNAs. Molecular cell. 2017;65(1):25-38.
- 419 89. Guo C-J, Ma X-K, Xing Y-H, Zheng C-C, Xu Y-F, Shan L, et al. Distinct processing of lncRNAs contributes 420 to non-conserved functions in stem cells. Cell. 2020;181(3):621-36. e22.
- 421 90. Melé M, Mattioli K, Mallard W, Shechner DM, Gerhardinger C, Rinn JL. Chromatin environment,
 422 transcriptional regulation, and splicing distinguish lincRNAs and mRNAs. Genome research. 2017;27(1):27423 37.
- 424 91. Zuckerman B, Ulitsky I. Predictive models of subcellular localization of long RNAs. Rna. 2019;25(5):557425 72.
- 426 92. Kung JT, Colognori D, Lee JT. Long noncoding RNAs: past, present, and future. Genetics. 2013;193(3):65169.
- 428 93. Wang KC, Chang HY. Molecular mechanisms of long noncoding RNAs. Molecular cell. 2011;43(6):904-14.
- 429 94. Gong Y, He J, Li B, Xiao Y, Zeng Q, Xu K, et al. Integrated Analysis of lncRNA and mRNA in Subcutaneous
 430 Adipose Tissue of Ningxiang Pig. Biology. 2021;10(8):726.
- 431 95. Song F, Wang L, Zhu W, Dong Z. Long noncoding RNA and mRNA expression profiles following igf3
 432 knockdown in common carp, Cyprinus carpio. Scientific data. 2019;6(1):1-8.

- 433 96. Jiang L, Yang Q, Yu J, Liu X, Cai Y, Niu L, et al. Identification and expression analysis of lncRNA in seven organs of Rhinopithecus roxellana. Functional & Integrative Genomics. 2021;21(5):543-55.
- 435 97. Quinn JJ, Zhang QC, Georgiev P, Ilik IA, Akhtar A, Chang HY. Rapid evolutionary turnover underlies
 436 conserved lncRNA-genome interactions. Genes & development. 2016;30(2):191-207.
- 437 98. Clark MB, Amaral PP, Schlesinger FJ, Dinger ME, Taft RJ, Rinn JL, et al. The reality of pervasive transcription. PLoS biology. 2011;9(7):e1000625.
- 439 99. Ma L, Bajic VB, Zhang Z. On the classification of long non-coding RNAs. RNA biology. 2013;10(6):924440 33.
- 100. Suarez B, Prats-Mari L, Unfried JP, Fortes P. LncRNAs in the type I interferon antiviral response.
 International Journal of Molecular Sciences. 2020;21(17):6447.
- 443 101. Ulitsky I, Bartel DP. lincRNAs: genomics, evolution, and mechanisms. Cell. 2013;154(1):26-46.
- 444 102. Fang Y, Fullwood MJ. Roles, functions, and mechanisms of long non-coding RNAs in cancer. Genomics,
 445 proteomics & bioinformatics. 2016;14(1):42-54.
- Homoson 103. Budak H, Kaya SB, Cagirici HB. Long non-coding RNA in plants in the era of reference sequences. Frontiers
 in plant science. 2020;11:276.
- 104. Ros G, Pegoraro S, De Angelis P, Sgarra R, Zucchelli S, Gustincich S, et al. HMGA2 antisense long non-coding RNAs as new players in the regulation of HMGA2 expression and pancreatic cancer promotion.
 Frontiers in oncology. 2020;9:1526.
- 451 105. Mo S, Zhang L, Dai W, Han L, Wang R, Xiang W, et al. Antisense lncRNA LDLRAD4-AS1 promotes
 452 metastasis by decreasing the expression of LDLRAD4 and predicts a poor prognosis in colorectal cancer.
 453 Cell death & disease. 2020;11(2):1-16.
- 454 106. Grote P, Wittler L, Hendrix D, Koch F, Währisch S, Beisaw A, et al. The tissue-specific lncRNA Fendrr is an
 455 essential regulator of heart and body wall development in the mouse. Developmental cell. 2013;24(2):206456 14.
- 457 107. Zhang Y, Huang X, Liu J, Chen G, Liu C, Zhang S, et al. New insight into long non-coding RNAs associated
 458 with bone metastasis of breast cancer based on an integrated analysis. Cancer cell international. 2021;21(1):1459 17.
- 460 108. Jiang P, Hou Y, Fu W, Tao X, Luo J, Lu H, et al. Characterization of lncRNAs involved in cold acclimation
 461 of zebrafish ZF4 cells. PloS one. 2018;13(4):e0195468.
- 462 109. Simion V, Zhou H, Haemmig S, Pierce JB, Mendes S, Tesmenitsky Y, et al. A macrophage-specific lncRNA
 463 regulates apoptosis and atherosclerosis by tethering HuR in the nucleus. Nature communications.
 464 2020;11(1):1-16.

- 465 110. Jiang Y-Y, Zhu H, Zhang H. Analysis of orthologous lncRNAs in humans and mice and their species-specific
 466 epigenetic target genes. Nan Fang yi ke da xue xue bao= Journal of Southern Medical University.
 467 2018;38(6):731-5.
- 468 111. Yanai I, Benjamin H, Shmoish M, Chalifa-Caspi V, Shklar M, Ophir R, et al. Genome-wide midrange
 469 transcription profiles reveal expression level relationships in human tissue specification. Bioinformatics.
 470 2005;21(5):650-9.
- 471 112. Azlan A, Obeidat SM, Yunus MA, Azzam G. Systematic identification and characterization of Aedes aegypti
 472 long noncoding RNAs (lncRNAs). Scientific reports. 2019;9(1):1-9.
- 473 113. Kong L, Zhang Y, Ye Z-Q, Liu X-Q, Zhao S-Q, Wei L, et al. CPC: assess the protein-coding potential of
 474 transcripts using sequence features and support vector machine. Nucleic acids research.
 475 2007;35(suppl 2):W345-W9.
- 476
 476 114. Wang L, Park HJ, Dasari S, Wang S, Kocher J-P, Li W. CPAT: Coding-Potential Assessment Tool using an alignment-free logistic regression model. Nucleic acids research. 2013;41(6):e74-e.
- 478 115. Li A, Zhang J, Zhou Z. PLEK: a tool for predicting long non-coding RNAs and messenger RNAs based on an improved k-mer scheme. BMC bioinformatics. 2014;15(1):1-10.
- 480
 480 116. Sun L, Luo H, Bu D, Zhao G, Yu K, Zhang C, et al. Utilizing sequence intrinsic composition to classify
 481 protein-coding and long non-coding transcripts. Nucleic acids research. 2013;41(17):e166-e.
- 482
 483
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- 484 118. Guo J-C, Fang S-S, Wu Y, Zhang J-H, Chen Y, Liu J, et al. CNIT: a fast and accurate web tool for identifying
 485 protein-coding and long non-coding transcripts based on intrinsic sequence composition. Nucleic acids
 486 research. 2019;47(W1):W516-W22.
- 487 119. Gong Y, Huang H-T, Liang Y, Trimarchi T, Aifantis I, Tsirigos A. IncRNA-screen: an interactive platform for
 488 computationally screening long non-coding RNAs in large genomics datasets. BMC genomics. 2017;18(1):1 489 18.
- 490 120. Sun J, Xie M, Huang Z, Li H, Chen T, Sun R, et al. Integrated analysis of non-coding RNA and mRNA
 491 expression profiles of 2 pig breeds differing in muscle traits. Journal of animal science. 2017;95(3):1092492 103.
- 493 121. Mohamadipoor Saadatabadi L, Mohammadabadi M, Amiri Ghanatsaman Z, Babenko O, Stavetska R,
 494 Kalashnik O, et al. Signature selection analysis reveals candidate genes associated with production traits in
 495 Iranian sheep breeds. BMC veterinary research. 2021;17(1):1-9.
- Li W, Zheng M, Zhao G, Wang J, Liu J, Wang S, et al. Identification of QTL regions and candidate genes for growth and feed efficiency in broilers. Genetics Selection Evolution. 2021;53(1):1-17.

- 498 123. Alexandre PA, Reverter A, Berezin RB, Porto-Neto LR, Ribeiro G, Santana MH, et al. Exploring the
 499 regulatory potential of long non-coding RNA in feed efficiency of indicine cattle. Genes. 2020;11(9):997.
- 124. Ali A, Al-Tobasei R, Kenney B, Leeds TD, Salem M. Integrated analysis of lncRNA and mRNA expression
 in rainbow trout families showing variation in muscle growth and fillet quality traits. Scientific Reports.
 2018;8(1):1-17.
- Li H, Huang K, Wang P, Feng T, Shi D, Cui K, et al. Comparison of long non-coding RNA expression profiles
 of cattle and buffalo differing in muscle characteristics. Frontiers in genetics. 2020;11:98.
- 505 126. Muniz MMM, Simielli Fonseca LF, Scalez DCB, Vega AS, Silva DBdS, Ferro JA, et al. Characterization of
 506 novel lncRNA muscle expression profiles associated with meat quality in beef cattle. Evolutionary
 507 applications. 2022;15(4):706-18.
- 127. Li R, Li B, Jiang A, Cao Y, Hou L, Zhang Z, et al. Exploring the lncRNAs related to skeletal muscle fiber
 types and meat quality traits in pigs. Genes. 2020;11(8):883.
- 510 128. Özdemir S, Eltas Ö, ÇULHA MH. Expression profiles of lincRNA and mRNA related to milk yield and milk
 511 composition traits in the milk-derived exosomes of Holstein and Doğu Anadolu Kırmızısı cows. Turkish
 512 Journal of Veterinary & Animal Sciences. 2020;44(2):227-34.
- 513 129. Jing H, Chen Y, Qiu C, Guo M-y. LncRNAs Transcriptome Analysis Revealed Potential Mechanisms of
 514 Selenium to Mastitis in Dairy Cows. Biological Trace Element Research. 2022:1-9.
- 515 130. Mu T, Hu H, Feng X, Ma Y, Wang Y, Liu J, et al. Screening and Conjoint Analysis of Key lncRNAs for Milk
 516 Fat Metabolism in Dairy Cows. Frontiers in genetics. 2022;13.
- 517 131. Ouyang Q, Hu S, Wang G, Hu J, Zhang J, Li L, et al. Comparative transcriptome analysis suggests key roles
 518 for 5-hydroxytryptamlne receptors in control of goose egg production. Genes. 2020;11(4):455.
- 519 132. Ning C, Ma T, Hu S, Xu Z, Zhang P, Zhao X, et al. Long non-coding RNA and mRNA profile of liver tissue during four developmental stages in the chicken. Frontiers in genetics. 2020;11:574.
- 133. Li Q, Qiao J, Zhang Z, Shang X, Chu Z, Fu Y, et al. Identification and analysis of differentially expressed
 long non-coding RNAs of Chinese Holstein cattle responses to heat stress. Animal Biotechnology.
 2020;31(1):9-16.
- 134. Li Y, Kong L, Deng M, Lian Z, Han Y, Sun B, et al. Heat stress-responsive transcriptome analysis in the liver
 tissue of Hu sheep. Genes. 2019;10(5):395.
- 135. Ni Y, Wu F, Chen Q, Cai J, Hu J, Shen J, et al. Long noncoding RNA and mRNA profiling of hypothalamic pituitary-mammary gland axis in lactating sows under heat stress. Genomics. 2020;112(5):3668-76.
- Mumtaz PT, Taban Q, Bhat B, Ahmad SM, Dar MA, Kashoo ZA, et al. Expression of lncRNAs in response
 to bacterial infections of goat mammary epithelial cells reveals insights into mammary gland diseases.

- 530 Microbial Pathogenesis. 2022;162:105367.
- 137. Feng X, Li F, Wang F, Zhang G, Pang J, Ren C, et al. Genome-wide differential expression profiling of mRNAs and lncRNAs associated with prolificacy in Hu sheep. Bioscience Reports. 2018;38(2).
- 138. Lian Z, Zou X, Han Y, Deng M, Sun B, Guo Y, et al. Role of mRNAs and long non-coding RNAs in regulating
 the litter size trait in Chuanzhong black goats. Reproduction in Domestic Animals. 2020;55(4):486-95.
- 139. Wang Y, Hua R, Xue S, Li W, Wu L, Kang T, et al. mRNA/lncRNA expression patterns and the function of
 fibrinogen-like protein 2 in Meishan pig endometrium during the preimplantation phases. Molecular
 Reproduction and Development. 2019;86(4):354-69.
- 140. Wichman L, Somasundaram S, Breindel C, Valerio DM, McCarrey JR, Hodges CA, et al. Dynamic
 expression of long noncoding RNAs reveals their potential roles in spermatogenesis and fertility. Biology of
 reproduction. 2017;97(2):313-23.
- 541 141. Gil N, Ulitsky I. Regulation of gene expression by cis-acting long non-coding RNAs. Nature Reviews
 542 Genetics. 2020;21(2):102-17.
- 543 142. Marques AC, Ponting CP. Intergenic lncRNAs and the evolution of gene expression. Current opinion in genetics & development. 2014;27:48-53.
- 545 143. Carelli FN, Liechti A, Halbert J, Warnefors M, Kaessmann H. Repurposing of promoters and enhancers during mammalian evolution. Nature communications. 2018;9(1):1-11.
- 144. Xiang J-F, Yin Q-F, Chen T, Zhang Y, Zhang X-O, Wu Z, et al. Human colorectal cancer-specific CCAT1-L
 IncRNA regulates long-range chromatin interactions at the MYC locus. Cell research. 2014;24(5):513-31.
- 549 145. Ulitsky I. Evolution to the rescue: using comparative genomics to understand long non-coding RNAs. Nature
 550 Reviews Genetics. 2016;17(10):601-14.
- 146. Hezroni H, Ben-Tov Perry R, Meir Z, Housman G, Lubelsky Y, Ulitsky I. A subset of conserved mammalian
 long non-coding RNAs are fossils of ancestral protein-coding genes. Genome biology. 2017;18(1):1-15.
- 147. Lai F, Orom UA, Cesaroni M, Beringer M, Taatjes DJ, Blobel GA, et al. Activating RNAs associate with
 Mediator to enhance chromatin architecture and transcription. Nature. 2013;494(7438):497-501.
- 148. Werner MS, Sullivan MA, Shah RN, Nadadur RD, Grzybowski AT, Galat V, et al. Chromatin-enriched
 lncRNAs can act as cell-type specific activators of proximal gene transcription. Nature structural &
 molecular biology. 2017;24(7):596-603.
- 149. Isoda T, Moore AJ, He Z, Chandra V, Aida M, Denholtz M, et al. Non-coding transcription instructs
 chromatin folding and compartmentalization to dictate enhancer-promoter communication and T cell fate.
 Cell. 2017;171(1):103-19. e18.

- 150. Han X, Zhang J, Liu Y, Fan X, Ai S, Luo Y, et al. The lncRNA Hand2os1/Uph locus orchestrates heart development through regulation of precise expression of Hand2. Development. 2019;146(13):dev176198.
- 151. Ntini E, Louloupi A, Liz J, Muino JM, Marsico A, Ørom UAV. Long ncRNA A-ROD activates its target gene
 DKK1 at its release from chromatin. Nature communications. 2018;9(1):1-16.
- 152. Wang KC, Yang YW, Liu B, Sanyal A, Corces-Zimmerman R, Chen Y, et al. A long noncoding RNA maintains active chromatin to coordinate homeotic gene expression. Nature. 2011;472(7341):120-4.
- 567 153. Fanucchi S, Fok ET, Dalla E, Shibayama Y, Börner K, Chang EY, et al. Immune genes are primed for robust
 568 transcription by proximal long noncoding RNAs located in nuclear compartments. Nature genetics.
 569 2019;51(1):138-50.
- 570 154. Cho SW, Xu J, Sun R, Mumbach MR, Carter AC, Chen YG, et al. Promoter of lncRNA gene PVT1 is a tumor-suppressor DNA boundary element. Cell. 2018;173(6):1398-412. e22.
- 572 155. Wang F, Yuan JH, Wang SB, Yang F, Yuan SX, Ye C, et al. Oncofetal long noncoding RNA PVT1 promotes
 573 proliferation and stem cell-like property of hepatocellular carcinoma cells by stabilizing NOP2. Hepatology.
 574 2014;60(4):1278-90.
- 575 156. Kotzin JJ, Spencer SP, McCright SJ, Kumar DBU, Collet MA, Mowel WK, et al. The long non-coding RNA
 576 Morrbid regulates Bim and short-lived myeloid cell lifespan. Nature. 2016;537(7619):239-43.
- 577 157. Latos PA, Pauler FM, Koerner MV, Şenergin HB, Hudson QJ, Stocsits RR, et al. Airn transcriptional overlap,
 578 but not its lncRNA products, induces imprinted Igf2r silencing. Science. 2012;338(6113):1469-72.
- 579 158. Thebault P, Boutin G, Bhat W, Rufiange A, Martens J, Nourani A. Transcription regulation by the noncoding
 580 RNA SRG1 requires Spt2-dependent chromatin deposition in the wake of RNA polymerase II. Molecular
 581 and cellular biology. 2011;31(6):1288-300.
- 582 159. Elcheva IA, Spiegelman VS. The role of cis-and trans-acting rna regulatory elements in leukemia. Cancers.
 583 2020;12(12):3854.
- 160. Yap K, Mukhina S, Zhang G, Tan JS, Ong HS, Makeyev EV. A short tandem repeat-enriched RNA assembles
 a nuclear compartment to control alternative splicing and promote cell survival. Molecular cell.
 2018;72(3):525-40. e13.
- 161. Lee S, Kopp F, Chang T-C, Sataluri A, Chen B, Sivakumar S, et al. Noncoding RNA NORAD regulates
 genomic stability by sequestering PUMILIO proteins. Cell. 2016;164(1-2):69-80.
- 162. Tichon A, Perry RB-T, Stojic L, Ulitsky I. SAM68 is required for regulation of Pumilio by the NORAD long noncoding RNA. Genes & Development. 2018;32(1):70-8.
- 163. Amodio N, Raimondi L, Juli G, Stamato MA, Caracciolo D, Tagliaferri P, et al. MALAT1: a druggable long
 non-coding RNA for targeted anti-cancer approaches. Journal of hematology & oncology. 2018;11(1):1-19.

- 164. Kretz M, Siprashvili Z, Chu C, Webster DE, Zehnder A, Qu K, et al. Control of somatic tissue differentiation
 by the long non-coding RNA TINCR. Nature. 2013;493(7431):231-5.
- 595 165. Gong C, Maquat LE. lncRNAs transactivate STAU1-mediated mRNA decay by duplexing with 3' UTRs via
 596 Alu elements. Nature. 2011;470(7333):284-8.
- 597 166. Jalali S, Bhartiya D, Lalwani MK, Sivasubbu S, Scaria V. Systematic transcriptome wide analysis of IncRNA-miRNA interactions. PloS one. 2013;8(2):e53823.
- 599 167.Paraskevopoulou MD, Hatzigeorgiou AG. Analyzing miRNA–lncRNA interactions. Long non-coding RNAs:
 600 Springer; 2016. p. 271-86.
- 168. Wang J, Liu X, Wu H, Ni P, Gu Z, Qiao Y, et al. CREB up-regulates long non-coding RNA, HULC expression
 through interaction with microRNA-372 in liver cancer. Nucleic acids research. 2010;38(16):5366-83.
- 169. Klein U, Lia M, Crespo M, Siegel R, Shen Q, Mo T, et al. The DLEU2/miR-15a/16-1 cluster controls B cell
 proliferation and its deletion leads to chronic lymphocytic leukemia. Cancer cell. 2010;17(1):28-40.
- 170. Cai X, Cullen BR. The imprinted H19 noncoding RNA is a primary microRNA precursor. Rna.
 2007;13(3):313-6.
- 171. Wang J-y, Yang Y, Ma Y, Wang F, Xue A, Zhu J, et al. Potential regulatory role of lncRNA-miRNA-mRNA
 axis in osteosarcoma. Biomedicine & Pharmacotherapy. 2020;121:109627.
- 172. Zhou R-S, Zhang E-X, Sun Q-F, Ye Z-J, Liu J-W, Zhou D-H, et al. Integrated analysis of lncRNA-miRNA mRNA ceRNA network in squamous cell carcinoma of tongue. BMC cancer. 2019;19(1):1-10.
- 611 173. Fan C-N, Ma L, Liu N. Systematic analysis of lncRNA-miRNA-mRNA competing endogenous RNA
 612 network identifies four-lncRNA signature as a prognostic biomarker for breast cancer. Journal of translational
 613 medicine. 2018;16(1):1-12.
- Kino T, Hurt DE, Ichijo T, Nader N, Chrousos GP. Noncoding RNA gas5 is a growth arrest-and starvation associated repressor of the glucocorticoid receptor. Science signaling. 2010;3(107):ra8-ra.
- 616 175. Derrien T, Johnson R, Bussotti G, Tanzer A, Djebali S, Tilgner H, et al. The GENCODE v7 catalog of human
 617 long noncoding RNAs: analysis of their gene structure, evolution, and expression. Genome research.
 618 2012;22(9):1775-89.
- 619

Tables and Figures



- 624 Figure 1. A schematic diagram of RNA classification.



Α IncRNA IncRNA PCG1 PCG2 Enhancer ≤ 100 kb I L 636 В **⊖/X**⇒ PCG1 ChrA -IncRNA **>/X**> RBP ChrB • IncRNA PCG2 637

Figure 3. Two mechanisms by which lncRNAs affect PCG expression. (A) lncRNA activates or represses the
expression of pcg in the *cis*-acting condition. A representative method for PCG regulation of *cis*-acting lncRNAs
is to affect enhancers. (B) lncRNA activates or represses the expression of pcg in the *trans*-acting condition. A

641 representative method for pcg regulation of *trans*-acting lncRNAs is to affect RBPs.

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