

JAST (Journal of Animal Science and Technology) TITLE PAGE

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ARTICLE INFORMATION	Fill in information in each box below
Article Type	Research article
Article Title (within 20 words without abbreviations)	Dietary supplementation with coated zinc oxide enhanced growth performance by modulating systemic immunity and microbial composition in weaned pigs
Running Title (within 10 words)	Effects of coated zinc oxide in weaned pigs
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Competing interests	No potential conflict of interest relevant to this article was reported.
Funding sources State funding sources (grants, funding sources, equipment, and supplies). Include name and number of grant if available.	This research was supported by the High Value-Added Food Technology Development Program of the Korean Institute of Planning and Evaluation for Technology in Food, Agriculture, Forestry, and Fisheries (IPET), the Ministry for Food, Agriculture, Forestry, and Fisheries of the Republic of Korea (321037-05-3-HD030).
Acknowledgements	Not applicable.
Availability of data and material	Upon reasonable request, the datasets of this study can be available from the corresponding author.
Authors' contributions Please specify the authors' role using this form.	Conceptualization: Nam J, Shin I, Cho J, Kim HB, Kyoung H, Song M Data curation: Nam J, Shin I, Cho J, Kim HB, Kyoung H, Song M Formal analysis: Nam J, Ahn J, Kang Y, Kim S, Kim JT, Jung SW Methodology: Nam J, Shin I, Cho J, Kim HB Software: Nam J, Shin I, Cho J, Kim HB Validation: Shin I, Cho J, Kim HB, Kyoung H, Song M Investigation: Nam J, Shin I, Cho J, Kim HB Writing - original draft: Nam J, Shin I, Cho J, Kim HB Writing - review & editing: Nam J, Shin I, Cho J, Kim HB, Ahn J, Kang Y, Kim S, Kim JT, Jung SW, Kyoung, H, Song M

Ethics approval and consent to participate	The experimental protocol used in this experiment was reviewed and approved by the Institutional Animal Care and Use Committee of Chungnam National University, Daejeon, Republic of Korea. (approval# 202103A-CNU-080)
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8 **Abstract (up to 350 words)**

9 Dietary zinc oxide (ZnO) is widely adopted in the swine industry to alleviate post-weaning diarrhea and promote
10 the growth performance of weaned pigs. However, its low rate of absorption in the gastrointestinal (GI) tract leads
11 to excessive excretion, which causes severe environmental pollution. To overcome this limitation, dietary coated
12 ZnO (CZO) has been studied to enhance zinc absorption and mitigate its environmental influence. In the present
13 study, we investigated the effects of dietary CZO on growth performance, frequency of diarrhea, nutrient
14 digestibility, hematological and immune responses, and fecal microbiota of weaned pigs. A total of 72 weaned
15 pigs [7.30 ± 0.01 kg of average initial body weight (BW); 28 days old] were randomly assigned to three dietary
16 treatments (4 pigs/pen; 6 replicates/treatment): basal weaner diet (CON), CON with 2,500 ppm of ZnO (HZO),
17 and CON with 200 mg/kg of coated ZnO (CZO). The experiment was conducted for 6 weeks. The CZO group
18 increased ($p < 0.05$) average daily gain and gain to feed ratio and showed tendency to reduce ($p = 0.094$) the
19 frequency of diarrhea compared with the CON group during the first two weeks after weaning. The CZO group
20 showed a tendency for the lowest levels of serum tumor necrosis factor- α (TNF- α) ($p = 0.072$) on day 7 and
21 hematocrit ($p = 0.083$) on day 14 and the highest ($p = 0.072$) levels of the immunoglobulin M on day 14 among
22 the treatment groups. The HZO group tended to have the lowest ($p = 0.078$) serum cortisol levels on day 7 among
23 all treatments. At the genus level of gut microbiota analysis, the dietary CZO group showed a higher ($p < 0.05$)
24 relative abundance of *Prevotella*, *Eubacterium*, and *Lactobacillus* than pigs treated with the CON diet. The HZO
25 group showed a higher ($p < 0.05$) relative abundance of *Eubacterium* and *Lactobacillus* than the CON group. Our
26 results demonstrated that dietary CZO supplementation enhanced growth performance and reduced the incidence
27 of diarrhea by modulating systemic immune responses and shifting fecal microbial compositions in weaned pigs,
28 suggesting its potential as an alternative to the high dose ZnO diets.

29

30 **Keywords:** Diarrhea, Fecal microbiota, Growth performance, Systemic immunity, Zinc

INTRODUCTION

Weaning is considered a critical and challenging stage for piglets because of their immature intestinal system and underdeveloped immune functions [1,2]. During this stage, piglets are exposed to an increased risk of post-weaning diarrhea (PWD), which is a fatal condition that hampers the development of piglets and impairing intestinal barrier function by disrupting intestinal mucosa and tight junction proteins, thereby reducing feed intake and causing severe growth retardation [3,4]. Dietary zinc is considered an essential trace element for animals primarily to support immunity, growth, and overall health [5,6]. In swine production, zinc oxide (ZnO) is commonly used at high doses (2,000 to 3,000 ppm) and many previous studies have shown that dietary ZnO has been supplemented in nursery diets to alleviate PWD by supporting intestinal development while reducing gut permeability [7]. However, long-term supplementation with high levels of ZnO raised some serious concerns. It increased the resistance of antibacterial activity by selecting specific strains such as *E. coli* that are more metal tolerant, accumulates toxicity in the kidney and liver, and leads to substantial environmental pollution when excreted through swine manure, owing to their low absorption rates [8–10]. To address these issues, the European Union has prohibited the use of high levels of ZnO in the feed and other countries have begun to phase out the use of ZnO to address environmental concerns [11]. Therefore, various studies are being conducted to control dietary ZnO doses and enhance the bioavailability of zinc, aiming to reduce the reliance on high levels of ZnO in weaner diets [12–14].

ZnO is highly soluble in acidic environments, causing it to be broken down into zinc ions by gastric acid, resulting in a relatively small amount of ZnO reaching the small intestine [15,16]. In addition, it can bind to substances such as phytate to form insoluble complexes, further reducing its bioavailability [17]. Enteric coating technology acts as a protective barrier and has been developed to enhance the delivery of ZnO into the small intestine. With the application of the coating, the inner core of ZnO reduced the dissociation by gastric acid, allowing it to reach the small intestine where it can effectively perform its function and be released by pancreatic lipase [13,18]. In an *in vitro* study, Shen et al. (2014) reported that the dissociation percentage under acidic conditions was higher for uncoated ZnO than for coated ZnO. Moreover, previous studies showed that dietary inclusion of coated ZnO improved growth rate, nutrient digestibility, and alleviated PWD when compared with a basal nursery diet and achieved similar effects to those observed with pharmacological levels of ZnO [13,18,19]. However, the effects of lipid-coated ZnO still remain controversial, and relatively few studies have examined its effect on the changes of fecal microbiota in the weaning piglets. Therefore, our study aimed to investigate the

60 effects of dietary supplementation with coated ZnO as an alternative to high doses of conventional ZnO on growth
61 performance, nutrient digestibility, hematological parameters, systemic immune responses, and fecal microbiota
62 of weaned pigs.

63

64

MATERIALS AND METHODS

65 *Origins of the tested product*

66 The lipid encapsulated ZnO used in the present study was supplied by (ACC Inc., Seongnam-si,
67 Republic of Korea). According to the protocol from manufacturer, this product is a double-coated form of zinc
68 derived from ZnO in the form of lipid matrix composed of fatty acids and hydrogenated palm oil. The ZnO content
69 of the product was 51%.

70 *Experimental design, animals, and diets*

71 This study was approved by the Institutional Animal Care and Use Committee of Chungnam National
72 University, Daejeon, Republic of Korea. (approval# 202103A-CNU-080). A total of 72 weaned piglets [(Landrace
73 × Yorkshire) × Duroc] with average initial BW of 7.30 ± 0.01 kg around 28 days age were conducted for 6 weeks.
74 Pigs were randomly assigned to three dietary treatments (4 pigs/pen; 6 replicates/treatment) (block= initial BW)
75 with pen as the experimental unit. The dietary treatments were 1) a basal weaner diet based on corn-soybean meal
76 (CON), 2) CON supplemented with 2,500 ppm high levels of conventional ZnO (HZO), and 3) CON
77 supplemented with 200 mg/kg dietary coated ZnO (CZO) for 6 weeks. The basal diet was designed to meet or
78 exceed the nutritional requirements of the National Research Council [20] for weaned pigs. The environments of
79 the pig room were controlled, with the temperature set between 28 to 32°C and allowing *ad libitum* access to feed
80 and water in the pens of the same sized (232 × 175 cm; width × length) during the trial.

81 *Data and sample collection*

82 For measuring growth performance, individual BW and feed residual amount were weighed and noted
83 on days 1, 14, and 42 and the average daily gain (ADG), average daily feed intake (ADFI), and gain to feed ratio
84 (G:F) were calculated. Fecal scores in each pen were visually monitored by two independent observers during the
85 first two weeks. The fecal scores ranged from 1 to 5, with 1 indicating hard and dry feces, 2 indicating soft feces,

86 3 indicating moist feces, 4 indicating mild diarrhea, and 5 indicating watery diarrhea. Diarrhea percentages were
87 determined by recording the number of days each pens received a score of 4 or higher, and these days were
88 expressed as a percentage. In the last week of the trial, 0.2% of chromium oxide (Daejung Chemicals & Metals
89 Co. Ltd., Siheung- si, Gyeonggi- do, Republic of Korea) was added to the diets for the each pen as an indigestible
90 marker and feces were collected during the last 3 days of the experiment via rectal palpation after 4 days of
91 adaptation and stored at -80°C for apparent total tract digestibility (ATTD) analysis. In fecal microbiota analysis,
92 feces were obtained from three randomly selected pigs per treatment on the first and final day of the experiment
93 and stored at -80°C until analysis. Blood samples (10 mL) were collected from the jugular vein of one randomly
94 chosen pig per pen using with or without ethylenediaminetetraacetic acid (EDTA) as the anticoagulant vacutainer
95 tubes (BD Vacutainer Systems, Franklin Lakes, NJ, USA on days 1, 7, and 14. Serum samples were collected from
96 the blood samples in non-EDTA tubes which were allowed to clot at room temperature and centrifuged for 15 min
97 at $3,000 \times g$ at 4°C and stored at -80°C for immune response analysis.

98

99 *Nutrient digestibility analysis*

100 Feed and frozen fecal samples were dried in a forced-air dry oven at 65°C for 72 hours. After drying,
101 these samples were ground using a electric grinder (Hamilton Beach Inc, Virginia, USA) for chemical analysis.
102 All grounded samples were analyzed for dry matter (DM), and energy using a bomb calorimeter (Parr 1281 Bomb
103 Calorimeter, Parr Instrument, Moline, IL, USA), and crude protein (CP) using the Kjeldahl method according to
104 the procedures outlined by the Association of Official Analytical Chemists [22]. Chromium concentrations in the
105 diets and fecal samples were estimated using an absorption spectrophotometer (Hitachi Z-5000 Absorption
106 Spectrophotometer, Hitachi High-Technologies Co., Tokyo, Japan). The ATTD values for DM, CP, and energy in
107 each treatment were determined based on a previous study [23].

108 *Blood samples analysis*

109 Hematological parameters were measured from blood samples collected in EDTA tubes using the Scil
110 Vet abc hematology analyzer (scil Vet abc hematology analyzer, scil Animal Care Company, Altorf, France)
111 adjusted for porcine blood. Measurement included total white blood cell (WBC) count, red blood cell (RBC) count,
112 and hematocrit (HCT) level. Systemic immune responses were analyzed by measuring serum concentrations of
113 tumor necrosis factor- α (TNF- α) and cortisol using ELISA kits (R&D Systems Inc., Minneapolis, MN, USA), as

114 well as immunoglobulin A (IgA), immunoglobulin G (IgG), and immunoglobulin M (IgM) using ELISA kits
115 (Bethyl Laboratories, Inc., Waltham, MA, USA). Absorbance was measured at 450 nm using a microplate reader
116 and the concentration of each sample was determined based on their standard curve.

117

118 *Fecal microbiota analysis*

119 The total DNA was extracted from the use of 200 mg of feces from each sample using the QIAamp Fast
120 DNA Stool Mini Kit (QIAGEN, Hilden, Germany) according to the manufacturer's guidelines. DNA
121 concentrations were then measured with the Colibri Microvolume Spectrometer (Titertek Berthold, Pforzheim,
122 Germany). Samples with the ratio of OD260/280 ranging from 1.80 to 2.15 were processed additionally. To
123 amplify the V5 to V6 regions of the 16S rRNA gene, the sets of polymerase chain reaction (PCR) primers 799F-
124 mod6 and 1114R were used [24]. After amplification, the products were purified using a Wizard® SV Gel and
125 PCR Clean Up System purification kit (Promega, Madison, WI, USA). Purified amplicons of the 16S rRNA gene
126 were sequenced using an Illumina MiSeq platform (Macrogen Inc., Seoul, Republic of Korea). Raw sequence data
127 produced using the Illumina MiSeq platform were subjected to quality assessment using FastQC. Sequencing
128 errors that were inconsistent with the PCR primers contained ambiguous bases, or were less than 200 bp in length
129 were eliminated using Mothur software [25]. The QIIME2 and Microbial Helper pipelines performed de novo
130 operational taxonomic unit (OTU) selection using the Deblur algorithm with a 97% identity cutoff. Alpha
131 (observed OTUs, Chao1, Shannon, and Simpson) and Beta diversity based on principal coordinate analysis (PCoA)
132 using Bray-Curtis indices were measured to assess microbial richness and evenness and comparing microbial
133 communities within each dietary treatment as well as comparing microbial communities across different dietary
134 treatments, respectively.

135 *Statistical analyses*

136 All data except the frequency of diarrhea and fecal microbiota were analyzed using PROC GLM of SAS
137 (v. 9.4; SAS Inst., Cary, NC, USA) in a randomized complete block design (block = initial BW). The experiment
138 unit was the pen. The statistical models for growth performance, digestibility, hematological parameters, and
139 immune responses of pigs included diet as the main effect and initial BW as a covariate. The Chi-square test was
140 used to analyze the frequency of diarrhea. The MicrobiomeAnalyst (<https://www.microbiomeanalyst.ca/>) webtool
141 was used to analyze the alpha and beta diversity statistics using the Kruskal-Wallis test and PERMANOVA,

142 respectively. Taxonomic classification was analyzed by STAMP software v. 2.1.3 [26] using a two-sided Welch's
143 t-test. Results were presented as means \pm SEM, except alpha diversity presented as means \pm SD. Statistically
144 differences and tendency were considered at $p < 0.05$ and $0.05 \leq p < 0.10$, respectively.

145 RESULTS

146 *Growth performance and nutrient digestibility*

147 During the first two weeks of the study, the pigs fed CZO exhibited a significant increase ($p < 0.05$) in
148 ADG and G:F compared with those fed CON (Table 2). However, there were no differences in the growth
149 performance from day 15 to 42 or in the overall period among the dietary treatments. The frequency of diarrhea
150 was measured at 13.87% in CON, 11.05% in HZO, and 10.16% in the CZO group. The pigs in the CZO group
151 tended to have a lower ($p = 0.094$) incidence of diarrhea frequency than those in the CON group. No differences
152 were observed in the ATTD of DM, CP, and energy among the dietary treatments (Table 3).

153 *Hematological parameters and immune responses*

154 No significant differences were observed in WBC and RBC counts on days 1, 7, and 14 among the
155 dietary treatments (Table 4). However, pigs fed with CZO tended to have the lowest hematocrit (HCT) level on
156 day 14 ($p = 0.083$). Regarding systemic immunity, pigs in the CZO group showed a tendency toward the lowest
157 serum TNF- α level on day 7 ($p = 0.072$) and the highest IgM level on day 14 ($p = 0.072$) among the dietary
158 treatments. In addition, pigs in the HZO group tended to have the lowest serum cortisol concentration on day 7 (p
159 $= 0.078$) compared to other treatments. No significant differences were observed in IgG and IgA levels among the
160 dietary treatments.

161 *Fecal microbiota diversities*

162 The alpha diversity indices are presented in Table 5. There were no differences in alpha diversity indices
163 among the dietary treatments on day 1. However, pigs fed dietary CZO showed an increase ($p < 0.05$) in the
164 observed OTUs and Chao1 indices on day 42 compared with those fed HZO. Microbial beta diversity among the
165 dietary treatments was visualized using PCoA plots as shown in 2D and 3D (Figure 1). No differences were found
166 ($r^2 = 0.24$, $p > 0.05$) in the Bray–Curtis distance on day 1 among the dietary treatments. However, differences in
167 the clustering of microbial communities were detected ($r^2 = 0.40$, $p < 0.05$; Figure 1C and 1D) on day 42 among
168 the dietary treatments.

169 *Fecal bacterial taxonomic relative abundances*

170 The relative abundances of bacterial taxa at the phylum and genus levels among the dietary treatments
171 are presented in Figure 2 and 3, respectively. In terms of fecal microbial composition, Firmicutes (CON, 57.42%;
172 HZO, 50.48%; CZO, 58.26%) was the most dominant phylum among the dietary treatments on day 1, followed
173 by Bacteroidetes (CON, 25.46%; HZO, 23.99%; CZO, 23.08%). On day 42, Firmicutes remained the most
174 dominant phylum among the dietary treatments (CON, 69.37%; HZO, 66.14%; CZO, 70.81%). In contrast,
175 Bacteroidetes abundance was decreased (CON, 6.18%; HZO, 18.25%; CZO, 11.11%) and Actinobacteria was
176 increased (CON, 1.52%; HZO, 14.33%; CZO, 2.97%). *Prevotella* was the most dominant fecal genus among the
177 dietary treatments (CON, 24.87%; HZO, 32.67%; CZO, 27.36%) on day 1. On day 42, the dietary CZO group
178 showed a higher ($p < 0.05$) proportions of *Prevotella* (18.62% vs. 7.94%), *Eubacterium* (11.97% vs. 1.32%) and
179 *Lactobacillus* (9.58% vs. 5.13%) than the CON group (Figure 4). In addition, pigs fed CZO had a higher ($p <$
180 0.005) abundance of *Eubacterium* (11.97% vs. 1.42%) than those fed HZO. The dietary HZO group had higher (p
181 < 0.05) *Lactobacillus* abundance (19.67% vs. 5.13 and 9.58%, respectively) than the CON and CZO groups.
182 Additionally, pigs fed HZO had increased ($p < 0.05$) the proportions of *Prevotella* (14.46% vs. 7.94%) compared
183 with those fed CON.

184

185

DISCUSSION

186 Due to weaning stress, piglets often suffer from physiological disorders including digestive and immune
187 dysfunction, which reduce feed efficiency, impair nutrient absorption, and delay growth rate [27,28]. To mitigate
188 these challenges and prevent susceptibility to infections, pharmacological levels of ZnO are commonly
189 supplemented in the weaner diets [29–31]. This supplementation not only promotes growth performance but also
190 ameliorates the intestinal barrier environment, enhances systemic immune function, and modulates gut microbiota
191 composition, particularly during the first and second critical weeks of post-weaning [32]. However, inorganic zinc
192 has relatively low bioavailability compared with its organic forms, with most of it being discharged from the feces
193 which could be the main source of soil pollution [33,34]. Therefore, we hypothesized that low doses of coated
194 ZnO could enhance the zinc absorption rate in the gastrointestinal (GI) tract while maintaining an efficacy similar
195 to that of high levels of ZnO. In the present study, the dietary CZO group showed improved ADG and feed
196 efficiency during the first two weeks post-weaning compared with the CON group, which is consistent with

197 previous studies [16,18,35,36]. These results suggest that low doses of coated ZnO promote growth performance
198 through the slow release of Zn by pancreatic lipase and more effective delivery to the small intestine, achieving
199 growth promoting effects similar to those of high levels of conventional ZnO forms. In swine production, ZnO is
200 often added to a weaner diet to alleviate PWD caused by nutritional and physiological stress and *E.coli* infection
201 [37]. In the present study, the CZO group tended to have a lower frequency of diarrhea compared with the CON
202 group during the first two weeks of our study. Additionally, HCT levels in hematological parameters were lower
203 in pigs fed CZO than in those fed CON, which is an indicator of dehydration status typically accompanied by an
204 increase of frequency of diarrhea. Therefore, our results suggest the supplementation with coated ZnO enhanced
205 the intestinal health of pigs, which in turn positively affects their growth performance during the critical period
206 when piglets are most vulnerable to digestive disturbances.

207 Higher ATTD values of DM, CP, and energy were observed in previous studies with the administration
208 of chelated or coated ZnO in the weaner diet [12,16] which can be attributed to the enhanced activations of
209 digestive enzymes [38] and improved morphology of the small intestine, [39] which positively affects nutrient
210 absorption. However, in contrast to previous reports, our results showed no differences in the ATTD of DM, CP,
211 and energy among the dietary treatments. Biswas et al. (2023) [40] reported that the ATTD of DM, nitrogen, and
212 energy were not affected by amino acid-chelated zinc. This finding aligns with that of our study, suggesting that
213 the effects of zinc may vary depending on the animal breed, zinc source, and its concentration.

214 Zinc is an essential trace mineral playing a vital role in regulating the immune systems, including innate
215 and adaptive immunity, and zinc deficiency can negatively affect immune functions, development, and overall
216 health [41]. Disruption of the intestinal barrier and penetration of pathogenic bacteria, because of weaning stress,
217 increase the permeability of intestinal tissues [42,43]. This process activates mucosal immune cells resulting in
218 the upregulation including pro-inflammatory cytokines such as TNF- α , which is a potential marker of
219 inflammatory reactions [44]. In the present study, the serum concentration of TNF- α was decreased in the dietary
220 CZO supplementation compared with the CON diet, but did not differ from the HZO group. This finding suggests
221 that the supplementation of zinc in weaner diets modulated systemic immunity by inhibiting the pathway of
222 nuclear factor- κ B, which is a key transcription factor for the expression of TNF- α [45]. Additionally, a tendency
223 of cortisol concentration was observed on day 7 in the HZO group compared to the CON group. Cortisol, a
224 biomarker of stress, reflects activation of the hypothalamic–pituitary–adrenal (HPA) axis, which is triggered
225 during early weaning stress and leads to the overproduction of cortisol and pro-inflammatory cytokines in

226 response to inflammation [46]. In addition, zinc supplementation in weaned pigs not only regulating immune
227 functions through cell-mediated immunity but also enhancing humoral immunity. IgA, IgM, and IgG are the main
228 antibodies activated by B lymphocytes and are essential for humoral immunity [47]. Our current study indicated
229 that supplemented with CZO group tended to have increased serum IgM levels on day 14. A previous study
230 revealed that ZnO supplementation enhances IgM levels [48], which is consistent with our results. IgM is the first
231 antibody produced in response to antigens and has major functions in the host defense system against infections
232 (Sathe and Cusick, 2022). These results suggest that CZO supplementation is positively affected during early
233 weaning, which is the most critical period for infections. Our findings indicate that supplementation with
234 pharmacological and low doses of coated ZnO alleviates the stress response and modulates immune responses,
235 supporting the potential effect of enhancing the systemic immunity of weaned pigs.

236 The gut ecosystem is affected by the interactions between the immune systems and intestinal
237 microorganisms, leading to the utilization of nutrients and contributing to the maintenance of host homeostasis
238 [49] The intestinal microbiota showed increased diversity and richness after weaning which depends on the species,
239 age, and diet of the animals [50]. In the current study, we observed that dietary HZO reduced the species richness
240 indices (Observed OTUs and Chao1) on day 42 compared with dietary CZO. Previous studies have reported lower
241 richness values in groups treated with high levels of ZnO [15,51], which is in agreements with the results observed
242 in our study. Regarding beta diversity, communities of fecal microbiota were distinctly identified among the
243 dietary treatments on day 42. Based on the results of microbial diversities, the dietary CZO group potentially
244 contributed to the improvement in microbial richness and stable microbial communities among the dietary
245 treatments which may enhance intestinal health [52] . In addition to diversity analysis, the relative taxonomic
246 abundance of fecal microbial communities was investigated to further evaluate the intestinal health of the pigs.
247 Firmicutes and Bacteroidetes phyla were more abundant in the HZO and CZO groups than in the CON group at
248 the phylum level on day 42. These are the most common phyla in the GI tract of humans and pigs and can maintain
249 the balance of energy and promote the production of short chain fatty acids (SCFAs) [53,54]. In the present study,
250 the CZO group showed higher abundance of the genera *Prevotella* and *Eubacterium* compared with the CON
251 group on day 42. *Prevotella* is known as one of the most dominant genera in the GI tract during the weaning
252 period and plays a pivotal role in producing SCFAs that serve as an energy source and help protect against gut
253 inflammation [55,56]. Furthermore, the *Prevotella*-driven enterotype was positively correlated with weight gain
254 and feed intake [57], and the higher abundance of *Prevotella* observed in the present study indicates the beneficial

255 effects of growth performance on weaned pigs. The genus *Eubacterium* has been reported to be a butyrate-
256 producing bacterium as well as a core component of the gut microbiome [58]. Previous studies have shown that
257 *Eubacterium* spp. contributes to maintaining gut integrity by modulating the gut microbiota and are considered to
258 have similar effects to probiotics, such as *Lactobacillus* and *Bacillus* strain in promoting intestinal health [59]. In
259 addition, the higher *Lactobacillus* abundance was observed in the HZO group than in the CZO and CON groups
260 on day 42. Our current results are consistent with those of previous studies reported that only pigs fed
261 pharmacological levels of ZnO showed a higher abundance of *Lactobacillus* than those fed low doses of modified
262 ZnO [12]. This result may vary depending on the form of ZnO and it may be considered that the proportion of
263 *Lactobacillus* is relatively reduced as the proportion of other beneficial bacteria increased in the CZO group.

264 CONCLUSION

265 In summary, our study showed that supplementation with dietary coated ZnO improved growth
266 performance and alleviated the frequency of diarrhea in weaned pigs. Additionally, dietary coated ZnO at low
267 dosages modulated systemic immunity and enhanced the host gut health by altering the microbial communities
268 and shifting the relative microbial compositions of the fecal microbiota. As systemic immune modulation and
269 improved gut health are positively correlated with growth performance, these compounds could potentially replace
270 high levels of ZnO, which mainly contributes to environmental pollution. Further studies should be considered to
271 analyze the metagenomics indicators of zinc to elucidate the potential effects of coated ZnO.

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432 **Table 1.** Composition of basal diet for weaned pigs (as-fed basis)

Item	Basal diet
Ingredient, %	
Corn	49.86
Soybean meal, 44%	25.00
Whey powder	12.50
Soy protein concentrate	6.25
Soybean oil	3.00
Limestone	1.14
Monocalcium phosphate	1.05
Vitamin premix ¹⁾	0.20
Mineral premix ²⁾	0.20
L-Lysine·HCl	0.45
DL-Methionine	0.16
L-Threonine	0.13
L-Valine	0.06
Total	100.00
Calculated energy and nutrient	
Metabolizable energy, kcal/kg	3,465
Crude protein, %	21.26
Calcium, %	0.81
Phosphorous, %	0.65
Lysine, %	1.53
Methionine, %	0.47
Threonine, %	0.95
Tryptophan, %	0.25

433 ¹⁾Vitamin premix provided the following quantities of vitamin per kilogram of complete diet: vitamin A, 12,000
434 IU; vitamin D₃, 2,500 IU; vitamin E, 30 IU; vitamin K₃, 3 mg; D-pantothenic acid, 15 mg; nicotinic acid, 40 mg;
435 choline, 400 mg; and vitamin B₁₂, 12 µg.

436 ²⁾Mineral premix provided the following quantities of mineral per kilogram of complete diet: Fe, 90 mg from
437 iron sulfate; Cu, 8.8 mg from copper sulfate; Zn, 100 mg from zinc oxide; Mn, 54 mg from manganese oxide; I,
438 0.35 mg from potassium iodide; Se, 0.30 mg from sodium selenite.

439 **Table 2.** Effects of dietary coated ZnO on growth performance of weaned pigs¹⁾

Item ²⁾	CON	HZO	CZO	SEM	<i>p</i> -value
Day 1 to 14					
Initial BW, kg	7.29	7.30	7.29	0.32	0.995
Final BW, kg	10.65	11.12	11.75	0.41	0.673
ADG, g/d	240 ^b	273 ^{ab}	319 ^a	20.02	0.045
ADFI, g/d	435	428	424	25.26	0.963
G:F, g/g	0.552 ^b	0.638 ^{ab}	0.752 ^a	0.042	0.021
Day 15 to 42					
Initial BW, kg	10.65	11.12	11.75	0.41	0.673
Final BW, kg	21.75	22.15	22.45	0.34	0.576
ADG, g/d	396	394	382	12.75	0.953
ADFI, g/d	958	954	957	30.45	0.988
G:F, g/g	0.413	0.413	0.399	0.025	0.981
Day 1 to 42					
Initial BW, kg	7.29	7.3	7.29	0.32	0.995
Final BW, kg	21.75	22.15	22.45	0.34	0.576
ADG, g/d	344	354	361	7.12	0.482
ADFI, g/d	784	779	779	23.21	0.962
G:F, g/g	0.439	0.454	0.463	0.017	0.738
Frequency of diarrhea ³⁾ , %	13.87	11.05	10.16		0.094

440 ¹⁾Each value is the mean of 6 replicates (4 pigs/pen).

441 ²⁾CON = basal weaner diet based on corn and soybean meal, HZO = CON + 2,500 ppm ZnO, CZO = CON + 200
 442 mg/kg dietary coated ZnO, BW = body weight, ADG = average daily gain, ADFI = average daily feed intake,
 443 G:F= gain to feed ratio.

444 ³⁾Frequency of diarrhea for the first 2 weeks after weaning (%) = (number of diarrhea score of 4 or higher /
 445 number of pen days) × 100. Data was analyzed using the Chi-square test.

446 ^{a,b)}Means in the same row with different superscripts are different (*p* < 0.05)

447

448 **Table 3.** Effects of dietary coated ZnO on nutrient and energy apparent total tract digestibility of weaned pigs¹⁾

Item²⁾	CON	HZO	CZO	SEM	<i>p</i>-value
DM, %	72.45	75.05	77.01	5.45	0.413
Energy, %	74.26	76.63	78.03	5.19	0.569
CP, %	71.48	73.58	73.55	6.28	0.638

449 ¹⁾ Each value is the mean of 6 replicates (1 pig/pen).

450 ²⁾ CON = basal weaner diet based on corn and soybean meal, HZO = CON + 2,500 ppm ZnO, CZO = CON + 20
451 0 mg/kg dietary coated ZnO; DM= dry matter, CP= crude protein.

452

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Table 4. Effects of dietary coated ZnO on blood profile and immune responses of weaned pigs¹⁾

Item ²⁾	CON	HZO	CZO	SEM	<i>p</i> -value
WBC, ×10³/μL					
Day 1	13.54	13.18	14.60	2.59	0.585
Day 7	23.15	19.81	20.08	1.31	0.429
Day 14	24.65	23.27	18.80	2.48	0.511
RBC, ×10⁶ / μL					
Day 1	4.65	4.89	4.68	0.23	0.405
Day 7	6.05	6.28	6.29	0.41	0.894
Day 14	6.75	6.45	6.40	0.32	0.475
HCT, %					
Day 1	26.08	25.02	25.42	0.82	0.835
Day 7	31.55	28.63	29.54	1.85	0.775
Day 14	36.67 ^a	31.05 ^{ab}	30.25 ^b	1.91	0.083
TNF-α, pg/mL					
Day 1	140.10	132.64	133.35	25.52	0.515
Day 7	109.01 ^a	80.29 ^b	78.57 ^b	9.51	0.072
Day 14	105.44	84.55	85.91	38.91	0.507
Cortisol, ng/mL					
Day 1	115.05	118.74	116.72	18.64	0.691
Day 7	123.43 ^a	81.18 ^b	86.54 ^{ab}	12.52	0.078
Day 14	105.61	98.53	97.53	17.45	0.303
IgG, mg/mL					
Day 1	5.26	4.95	5.01	0.76	0.776
Day 7	3.67	3.79	3.83	1.12	0.675
Day 14	3.75	4.52	4.26	0.31	0.268
IgM, mg/mL					
Day 1	1.47	1.68	1.23	0.32	0.514
Day 7	1.46	1.48	1.47	0.22	0.638
Day 14	1.22 ^b	1.61 ^{ab}	1.65 ^a	0.13	0.072
IgA, mg/mL					
Day 1	0.28	0.31	0.21	0.07	0.496
Day 7	0.33	0.31	0.30	0.10	0.728
Day 14	0.39	0.54	0.44	0.13	0.464

454 ¹⁾ Each value is the mean of 6 replicates (1 pig/pen).455 ²⁾ CON = basal weaner diet based on corn and soybean meal, HZO = CON + 2,500 ppm ZnO, CZO = CON +
456 200 mg/kg dietary coated ZnO, WBC = white blood cell, RBC = red blood cell, HCT = hematocrit, TNF-α =
457 tumor necrosis factor-alpha, IgG = immunoglobulin G, IgM = immunoglobulin M, IgA = immunoglobulin A.458 ^{a,b}Means in the same row with different superscripts are different (*p* < 0.05)

459 **Table 5.** Effects of dietary coated ZnO on bacterial alpha diversity of weaned pigs¹⁾

Item ²	CON	HZO	CZO	<i>p</i> -value
Day 1				
Observed OTUs	492.33 ± 10.30	504.33 ± 122.10	423.00 ± 68.74	0.697
Chao1	494.25 ± 104.20	506.62 ± 123.02	424.81 ± 68.50	0.699
Shannon	4.46 ± 0.35	4.40 ± 0.79	4.06 ± 0.54	0.769
Simpson	0.972 ± 0.010	0.940 ± 0.063	0.943 ± 0.041	0.732
Day 42				
Observed OTUs	417.67 ^{ab} ± 75.21	300.67 ^b ± 20.07	462.33 ^a ± 9.88	0.033
Chao1	420.06 ^{ab} ± 76.18	302.16 ^b ± 19.62	466.43 ^a ± 12.41	0.032
Shannon	3.77 ± 0.35	3.43 ± 0.46	4.09 ± 0.18	0.397
Simpson	0.946 ± 0.016	0.950 ± 0.017	0.952 ± 0.006	0.750

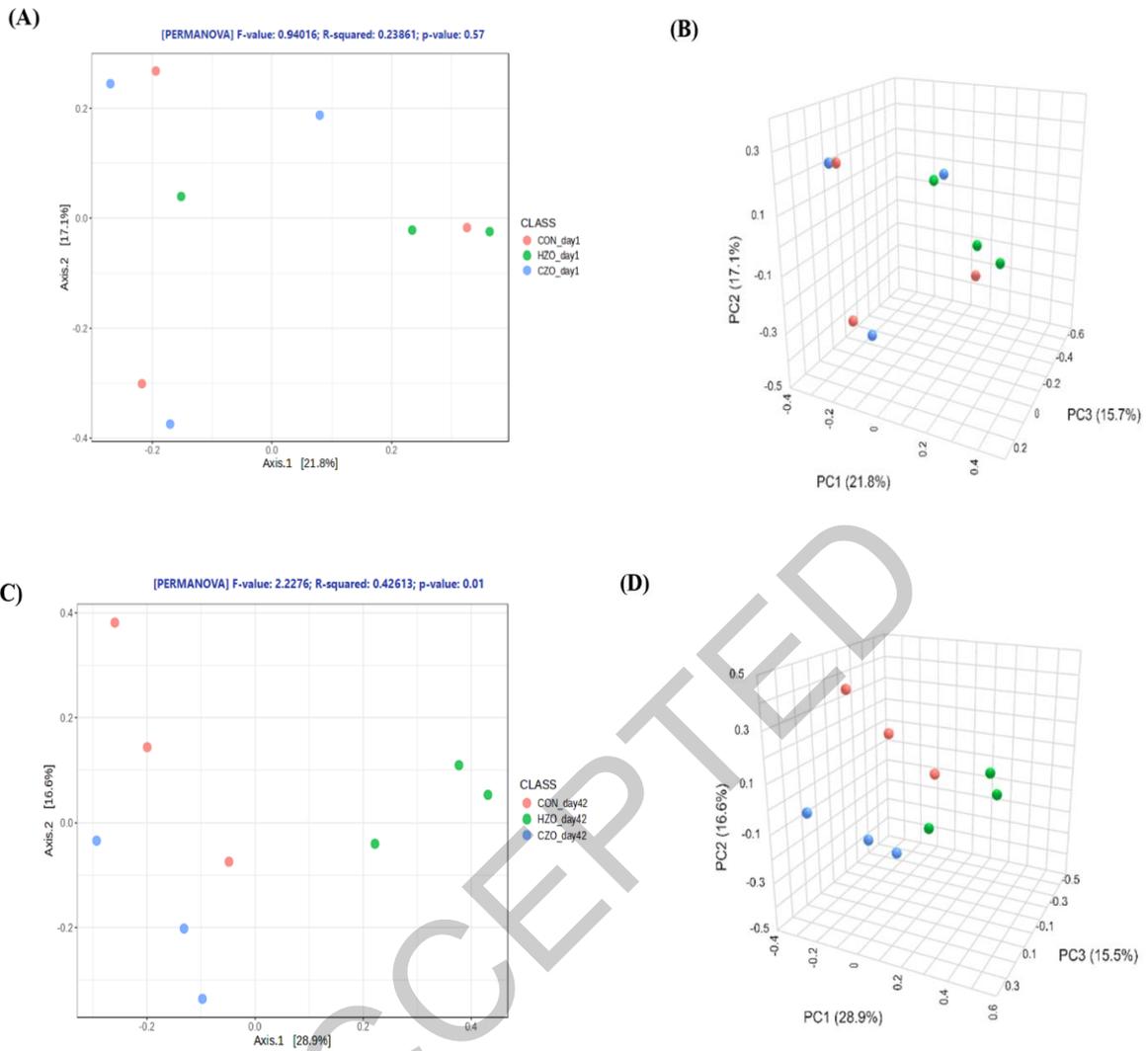
460 ¹⁾ Each value is the mean of 3 replicates and presented as mean ± SD.

461 ²⁾ CON = basal weaner diet based on corn and soybean meal, HZO = CON + 2,500 ppm ZnO, CZO = CON +
 462 200 mg/kg dietary coated ZnO; OTUs, operational taxonomic units.

463 ^{a,b}Means in the same row with different superscripts are different (*p* < 0.05)

464

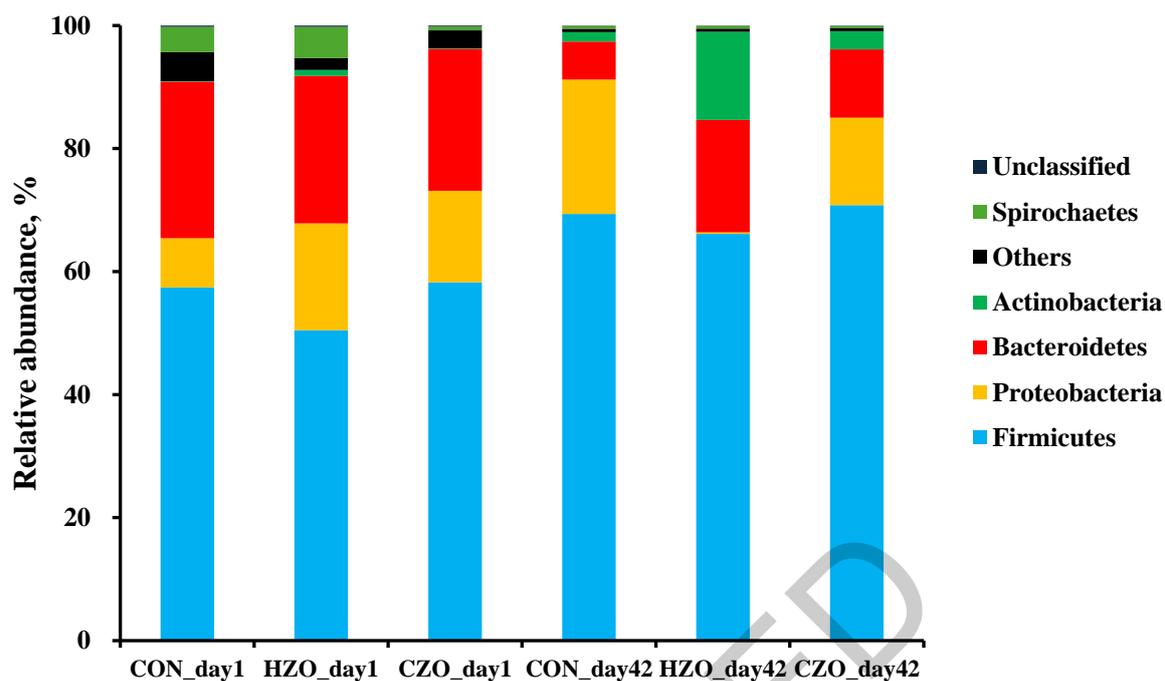
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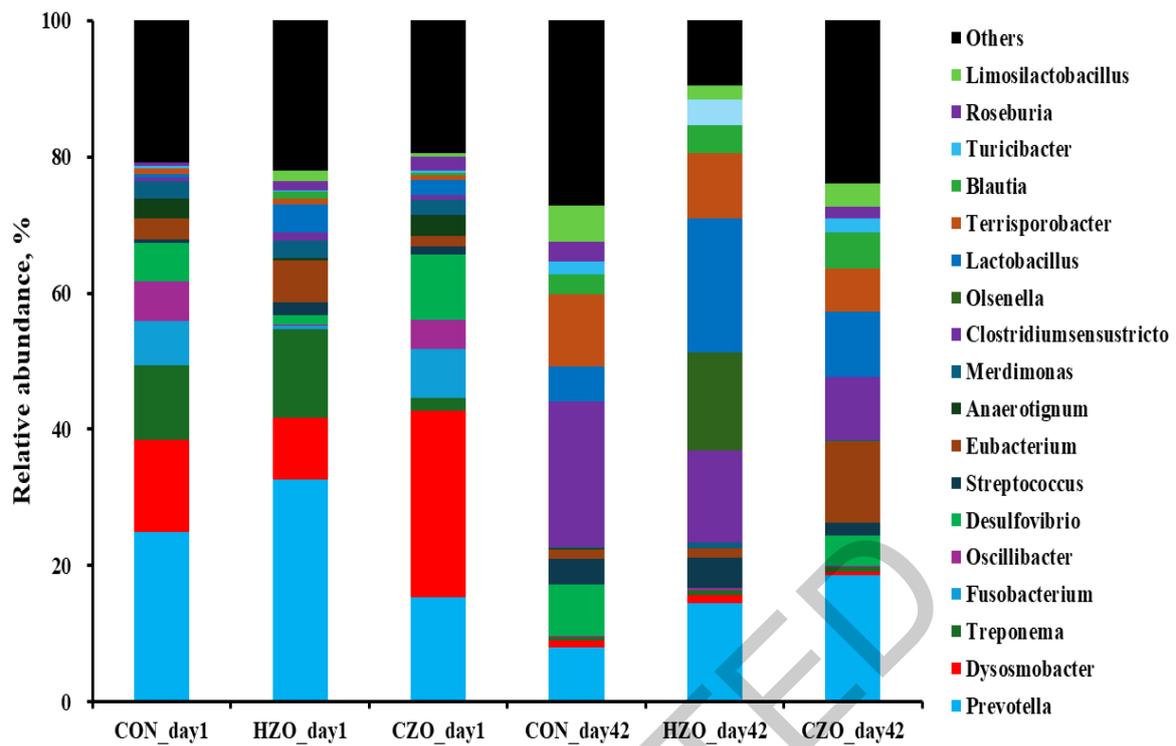
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468 **Figure 1.** Principal coordinates analysis (PCoA) based on Bray-Curtis distance of bacterial communities of
 469 weaned pigs (n = 3). Permutational multivariate analysis of variance (PERMANOVA) was used for statistical
 470 differences of clustering distances. Beta diversity analysis were represented for fecal bacteria (A), (C) at day 1
 471 and day 42 for ordination 2D and (B), (C) at day 1 and day 42 for PCoA 3D. CON = basal weaner diet based on
 472 corn and soybean meal; HZO = CON + 2,500 ppm ZnO, CZO = CON + 200 mg/kg dietary coated ZnO.



473

474 **Figure 2.** The relative abundance of the fecal microbiota at the phylum level among dietary treatments on day 1
 475 and day 42. The proportions of top five bacteria are presented at the phylum level in each time period. CON =
 476 basal weaner diet based on corn and soybean meal; HZO = CON + 2,500 ppm ZnO, CZO = CON + 200 mg/kg
 477 dietary coated ZnO.



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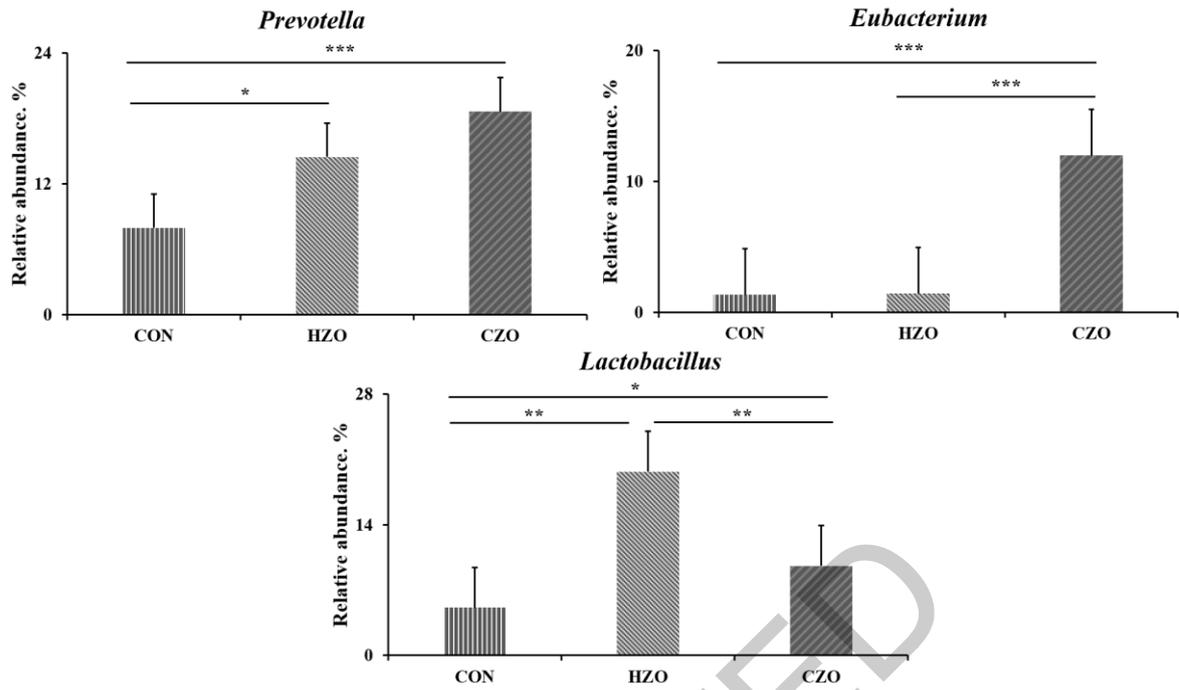
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Figure 3. The relative abundance of the fecal microbiota at the genus level among dietary treatments on day 1 and day 42. The proportions of top five bacteria are presented at the phylum level in each time period. CON = basal weaner diet based on corn and soybean meal; HZO = CON + 2,500 ppm ZnO, CZO = CON + 200 mg/kg dietary coated ZnO.



484

485 **Figure 4.** The bar plots verifying relative abundance differences of fecal microbiota of weaned pigs at the genus
 486 level among dietary treatments. *Prevotella* (A), *Eubacterium* (B), and *Lactobacillus* (C) at day 42. CON = basal
 487 weaner diet based on corn and soybean meal; HZO = CON + 2,500 ppm ZnO, CZO = CON + 200 mg/kg dietary
 488 coated ZnO. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.005$.

489