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Dietary protease improves growth performance, nutrient digestibility, and intestinal morphology of weaned pigs

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Abstract

This study was conducted to evaluate effects of dietary protease (PR) on growth performance, nutrient digestibility, and intestinal morphology of weaned pigs. A total of 75 weaned pigs [7.06 ± 0.18 kg of average body weight (BW); 28 day old] were randomly allotted to 3 dietary treatments in a randomized complete block design (blocks = BW and sex): a diet based on corn and soybean meal to meet the requirement of crude protein (CP) as a positive control (PC; CP = 24.49%), a low protein diet as a negative control (NC; CP = 22.51%), and NC + 0.02% PR. The PR used in this study was a commercial product containing 75.000 protease units/g derived from Nocardiopsis prasina produced in Bacillus licheniformis. Pigs were fed the dietary treatments for 6 weeks and the diets containing 0.2% chromic oxide for the last week of this study. Blood, feces, ileal digesta, and ileum samples were collected from randomly selected two pigs in each pen on respective time points. Measurements were growth performance, apparent ileal digestibility (AID) and apparent total tract digestibility (ATTD) of dry matter (DM), CP, and energy, frequency of diarrhea, packed cell volume (PCV), and ileal morphology of weaned pigs. Pigs fed PC and PR had higher (p < 0.05) final BW, average daily gain (ADG), and gain to feed ratio (G:F) during overall experimental period than those fed NC. Pigs fed PC and PR had higher (p < 0.05) AID or ATTD of DM, CP, or energy than those fed NC. Moreover, pigs fed PR had higher ratio between villus height and crypt depth (p < 0.05) and number of goblet cells (p < 0.05) than those fed NC. Addition of PR decreased (p < 0.05) frequency of diarrhea for the first two weeks after weaning compared with PC and NC. In addition, pigs fed PR had lower (p < 0.05) PCV on d 14 after weaning than those fed PC and NC. In conclusion, addition of PR in nursery diets with a low protein level significantly improved growth performance, nutrient digestibility, and intestinal morphology of weaned pigs. Keywords: Growth performance, Intestinal morphology, Nutrient digestibility, Protease,

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Competing interests

No potential conflict of interest relevant to this article was reported.

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Not applicable.

Availability of data and material

Upon reasonable request, the datasets of this study can be available from the corresponding author.

Authors' contributions

Conceptualization: Cho JH, Song M. Data curation: Kim HB, Song M. Formal analysis: Cho JY, Perez-Maldonado R. Methodology: Yang BM, Park IH. Software: Oh S, Park DJ. Validation: Lee JJ, Kim S. Investigation: Park S, Yang BM, Kang J. Writing - original draft: Park S, Lee JJ, Park IH, Song M. Writing - review & editing: Park S, Cho JH, Song M.

Ethics approval and consent to participate The protocol for this experiment was reviewed and approved by the Institutional Animal Care and Use Committee of

Chungnam National University, Daejeon, Korea (approval code: CNU-00611).

INTRODUCTION

Soybean meal (SBM) is a major protein source in swine diets due to its excellent balance of amino acid such as lysine, threonine, tryptophan, and others. [1]. In general, SBM is the by-product of whole or dehulled soybeans after extraction of soy oil and contains approximately 44%–48% of crude protein (CP) [2,3]. However, SBM contains relatively high contents of anti-nutritional factors (ANFs) including protease inhibitors, allergens, lectins, phytoestrogens, oligosaccharedses, and phytin. [3–5]. Especially, the protease inhibitors reduce protein availability in the gastrointestinal tract of pigs because they bind and inactivate trypsin and chymotrypsin digestive enzymes [6]. These ANFs also have negative effects on utilization of dietary proteins and the health of pigs [7,8].

Addition of exogenous enzymes in animal diets has being successfully used to improve the availability of nutrients and to reduce feed costs in the animal industry [9]. Dietary protease (PR) is an exogenous enzyme able to degrade proteins and has being widely used in swine diets as a part of multi-enzyme products [10]. Several studies have reported supplementation of enzyme cocktails with protease in swine diets enhanced protein digestion and growth performance of weaning, growing, and finishing pigs [11–13]. Recent studies also reported addition of PR alone in swine diets impoved gut development and health of weaned pigs by breaking down protein-bound complexes with other ANFs and affected positively nutrient digestibility and growth performace of pigs [9,11–15]. However, there was limited information about effects of PR when given alone in swine diets. Therefore, the main objectives of this study were to investigate effects of PR on growth performance, nutrient digestibility, and intestinal morphology of weaned pigs.

MATERIALS AND METHODS

The protocol for this experiment was reviewed and approved by the Institutional Animal Care and Use Committee of Chungnam National University, Daejeon, Korea. This experiment was conducted at the Animal Research Center of Chungnam National University.

Experimental design, animals, and diets

A total of 75 weaned pigs [Duroc × (Landrace × Yorkshire); 7.06 ± 0.18 kg of initial body weight (BW); 28 day old] were randomly assigned to 3 dietary treatments (5 pigs with 3 barrows and 2 gilts per pen and 5 replicated pens per treatment) in a randomized completely block design (block = BW and sex). The dietary treatments were 1) a diet based on corn and SBM to meet or exceed the requirement of CP as a positive control (PC; CP = 24.49%), 2) a low protein diet as a negative control (NC; CP = 22.51%), and 3) NC + 0.02% protease (PR) ([2]; Table 1). The PR used in this study was a commercial product (Ronozyme® ProAct, DSM nutrition products, Kaiseraugst, Switzerland) containing 75,000 protease units/g derived from *Nocardiopsis prasina* produced in *Bacillus licheniformis*. The dietary treatments did not include animal plasma, antibiotics, or zinc oxide to avoid their antibacterial or physiological effects. Pigs were fed respective dietary treatments for 6 weeks. All pigs were housed in an environmentally controlled room with a slatted plastic floor and allowed *ad libitum* access to diets and water throughout the entire experiment period.

Data and sample collection

Individual pigs and amount of feed additions and refusals in each pen were weighed and recorded to measure average daily gain (ADG), average daily feed intake (ADFI), and ratio between ADG and ADFI (G:F) of pigs. Diarrhea of each pig was checked and its visual score was recorded by 3 independent evaluators with a score from 1 to 5 (1 = normal hard feces; 2 = slightly soft feces; 3 =

Items	PC	NC
Ingredients (%)		
Corn	56.09	58.09
Soybean meal (44%)	26.00	24.00
Soy protein concentrate	12.00	12.00
Soybean oil	3.00	3.00
Limestone	1.30	1.30
Monocalcium phosphate	1.20	1.20
Vitamin-mineral premix ¹⁾	0.04	0.04
L-Lysine-HCI	0.24	0.24
DL-Methionine	0.09	0.09
L-Threonine	0.04	0.04
Total	100	100
Calculated energy and nutrient contents		
Metabolizable energy (Mcal/kg)	3.53	3.42
Crud protein (%)	24.49	22.51
Calcium (%)	0.81	0.73
Phosphorus (%)	0.69	0.63
Lysine (%)	1.54	1.41

Table 1 Composition	on of basal diet for weaned	nigs (a	s-fed basis)
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¹The vitamin-mineral premix provided the following quantities of vitamins per kilogram of diet: vitamin A, 12,000 IU; vitamin D₃, 2,500 IU; vitamin E, 30 IU; vitamin K₃, 3 mg; D-pantothenic acid, 15 mg; nicotinic acid, 40 mg; choline, 400 mg; and vitamin B₁₂, 12 μg; Fe, 90 mg from iron sulfate; Cu, 8.8 mg from copper sulfate; Zn, 100 mg from zinc oxide; Mn, 54 mg from manganese oxide; I, 0.35 mg from potassium iodide; Se, 0.30 mg from sodium selenite.

PC, positive control; NC, negative control.

soft, partially formed feces; 4 = loose, semi-liquid feces; and 5 = watery, mucous-like feces) each day for the first 2 weeks of this experiment [16]. Frequency of diarrhea was calculated by counting pen days with average diarrhea score from individual pigs in each pen of 4 or greater [17,18]. Whole blood samples were collected from the jugular vein of randomly selected 2 pigs in each pen using EDTA tubes (Becton Dickinson Vacutainer Systems, Franklin Lakes, NJ, USA) containing anticoagulant on day 1, 3, 7, and 14 after weaning. For the last week of the experiment period, pigs were fed respective dietary treatments containing 0.2% chromic oxide as an indigestible marker. Fecal samples were collected from randomly selected 2 pigs in each pen by rectal palpation daily for the last 3 days after the 4-d adjustment period. The collected fecal samples were pooled and stored at −20 °C until analysis. Diet samples were also collected and stored at −20 °C until analysis. Randomly selected 2 pigs in each pen were anesthetized by an intra-muscular injection of a 2-mL suxamethonium chloride (Succicholine®, Ilsung Pharm. Co. Ltd., Seoul, Korea) at the end of this experiment. After anesthesia, pigs were euthanized by CO_2 gas [19]. Ileal digesta samples were collected from distal ileum before the ileocecal junction [11]. The collected ileal digesta samples were stored at -20° until analysis. Sections of ileum of 3 cm length were collected, washed gently with distilled water, and fixed in 10% neutral buffered formalin for histological analysis [20,21].

Sample analyses and measurements

Whole blood samples were analyzed to measure packed cell volume (PCV) using a multi-parameter, automated hematology analyzer calibrated for porcine blood (scil Vet abc hematology analyzer, scil animal care company, F-67120 Altorf, France). Frozen ileal digesta and fecal samples were freeze-dried and finely ground through a cyclone mill (Foss Tecator Sycltec 1093, Hillerød, Denmark) before chemical analysis. Diets, fecal samples, and ileal samples were analyzed for dry matter (DM; method 930.15), nitrogen (method 999.03) [22], gross energy using a bomb calorimeter (Parr 1281 Bomb Calorimeter, Parr Instrument Co., Moline, IL, USA), and chromium content using an absorption spectrophotometer (Hitachi Z-5000 Absorption Spectrophotometer, Hitachi High-Technologies Co., Tokyo, Japan) based on the report by Williams et al. [23]. The apparent ileal digestibility (AID) and apparent total tract digestibility (ATTD) of DM, CP, and energy were calculated for each diet according to Stein et al. [24]. The procedures for the measurement of intestinal morphology were based on the report by Liu et al. [20]. The fixed intestinal tissue samples were placed in paraffin, sliced at 5 μ m, and stained with hematoxylin and eosin. The stained slides were scanned by fluorescence microscopy (TE2000, Nikon, Tokyo, Japan) with a charge-coupled device (CCD) camera (DS-Fi1; Nikon, Tokyo, Japan) to measure intestinal morphology such as villus height, width, and area, crypt depth, ratio between villus height and crypt depth (VH:CD), and number of goblet cells by selecting ten straight and integrated villi and their associated crypts and goblet cells. The fluorescent images were processed with NIS-Elements BR software 3.00 (Nikon, Japan).

Statistical analyses

Data were analyzed using the PROC GLM procedure of SAS (SAS Inst. Inc., Cary, NC, USA) in randomized complete block design. The experimental unit was the pen and blocks were BW and sex. The statistical model for growth performance, PCV, AID and ATTD, and number of goblet cells included effects of dietary treatments as a fixed effect and BW and sex as covariates. In addition, pair-wise comparisons were performed among dietary treatments when a main effect of diet was found. The chi-squared test was used for the frequency of diarrhea. Results are given as mean \pm SEM. Statistical significance and tendency were considered at p < 0.05 and $0.05 \le p < 0.10$, respectively.

RESULTS AND DISCUSSION

Present study showed there were differences (p < 0.05) on ADG and G:F of weaned pigs during overall experimental period between PC and NC (Table 2). In addition, weaned pigs fed PR had higher ADG and G:F from d 1 to 7 (p < 0.05) and during overall experimental period (p < 0.05) than those fed NC (Table 2). However, no difference was found on growth performance of weaned pigs during overall experimental period between PC and PR (Table 2). These results are similar to the results from previous studies for weaned pigs [10,11,21] and for growing pigs [14,25]. The reason for these observations may be related to the improved nutrient utilization efficiency by addition of PR [25]. However, some studies reported that no positive effects were found on growth performance of growing-finishing pigs by addition of PR [13,26], maybe because of different growth stages of the pigs with different digestive systems [10].

In the present study, there were differences (p < 0.05) on AID of DM, CP, and energy of weaned pigs between PC and NC (Fig. 1). However, no differences were found on nutrient AID of weaned pigs between PC and PR (Fig. 1). Similar patterns were found (p < 0.05) on ATTD for DM and CP, but not for energy, compared with the results of nutrient AID (Fig. 2). The improved nutrient digestibility in this study was similar to results reported by previous studies [11,27,28]. Addition of exogenous enzymes in non-ruminant diets has been using for several benefits including, minimization of anti-nutritional effects and maximization of nutrient utilization, to improve nutrient utilization and growth performance of non-ruminants [29] because the digestive system and the secretion and activity of digestive enzymes of young non-ruminants are immature and insufficient

	Table 2. Effects of dietary	protease on growth	performance of weaned pigs ¹⁾
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ltem	D	ietary treatment	ts	SEM	<i>p</i> -value ²⁾			
nem	PC	NC	PR		Diet	PC vs. NC	PC vs. PR	NC vs. PF
Day 1 to 7								
Initial BW (kg)	7.01	7.03	7.13	0.09	0.597	0.892	0.356	0.428
Final BW (kg)	7.85	7.47	8.30	0.17	0.018	0.145	0.093	0.005
ADG (g/d)	120.11	63.51	167.20	22.87	0.024	0.100	0.171	0.008
ADFI (g/d)	334.80	321.27	339.14	25.69	0.878	0.716	0.907	0.632
G:F (g/g)	0.359	0.198	0.493	0.08	0.100	0.279	0.242	0.036
Day 8 to 14								
Initial BW (kg)	7.85	7.47	8.30	0.17	0.018	0.145	0.093	0.005
Final BW (kg)	9.82	9.29	10.12	0.31	0.207	0.254	0.512	0.086
ADG (g/d)	281.27	259.18	260.63	35.21	0.885	0.665	0.686	0.977
ADFI (g/d)	502.29	433.79	481.31	32.47	0.344	0.162	0.656	0.321
G:F (g/g)	0.560	0.597	0.541	0.06	0.862	0.714	0.875	0.602
Day 1 to 14								
Initial BW (kg)	7.01	7.03	7.13	0.09	0.597	0.892	0.356	0.428
Final BW (kg)	9.82	9.29	10.12	0.31	0.207	0.254	0.512	0.086
ADG (g/d)	200.69	161.35	213.91	21.80	0.247	0.226	0.676	0.114
ADFI (g/d)	418.54	377.53	410.23	25.18	0.497	0.272	0.819	0.377
G:F (g/g)	0.480	0.427	0.521	0.05	0.617	0.577	0.677	0.337
Day 15 to 21								
Initial BW (kg)	9.82	9.29	10.12	0.31	0.207	0.254	0.512	0.086
Final BW (kg)	12.63	11.23	12.66	0.44	0.071	0.048	0.957	0.043
ADG (g/d)	400.79	277.82	362.74	26.42	0.018	0.007	0.329	0.042
ADFI (g/d)	763.67	804.40	693.09	93.79	0.705	0.764	0.604	0.418
G:F (g/g)	0.525	0.345	0.523	0.04	0.049	0.035	0.916	0.043
Day 1 to 21								
Initial BW (kg)	7.01	7.03	7.13	0.09	0.597	0.892	0.356	0.428
Final BW (kg)	12.63	11.23	12.66	0.44	0.071	0.048	0.957	0.043
ADG (g/d)	267.39	200.17	263.52	20.33	0.066	0.038	0.895	0.048
ADFI (g/d)	533.59	519.82	504.51	38.57	0.869	0.805	0.604	0.784
G:F (g/g)	0.501	0.385	0.522	0.04	0.204	0.086	0.837	0.046
Day 22 to 42								
Initial BW (kg)	12.63	11.23	12.66	0.44	0.071	0.048	0.957	0.043
Final BW (kg)	20.56	16.26	20.69	0.85	0.005	0.004	0.916	0.003
ADG (g/d)	377.51	239.33	382.05	23.46	0.001	0.001	0.894	0.001
ADFI (g/d)	963.40	924.57	912.60	107.34	0.941	0.802	0.744	0.939
G:F (g/g)	0.392	0.259	0.419	0.04	0.027	0.038	0.986	0.019
Day 1 to 42								
Initial BW (kg)	7.01	7.03	7.13	0.09	0.597	0.892	0.356	0.428
Final BW (kg)	20.56	16.26	20.69	0.85	0.005	0.004	0.916	0.003
ADG (g/d)	322.45	219.75	322.79	20.16	0.005	0.004	0.991	0.004
ADFI (g/d)	748.49	722.19	708.56	60.42	0.894	0.764	0.649	0.876
G:F (g/g)	0.431	0.304	0.456	0.03	0.020	0.018	0.810	0.011

¹⁾Each value is the mean of 5 replicates (5 pigs/pen).

²⁾Diet, main diet effect; PC vs. NC, effect between positive and negative controls; PC vs. PR, effect between positive control and negative control + 0.02% dietary protease; NC vs. PR, effect between negative control and negative control + 0.02% dietary protease.

PC, positive control with required level of protein; NC, negative control with low level of protein; PR, negative control + 0.02% dietary protease; BW, body weight; ADG, average daily gain; ADFI, average daily feed intake; G:F, gain to feed ratio; SEM, standard error of means.

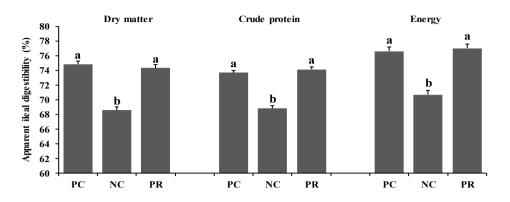
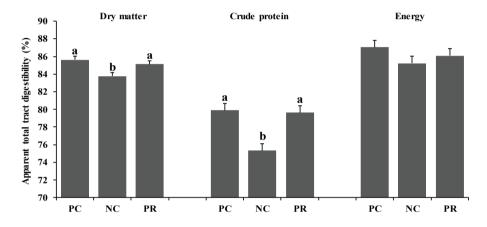
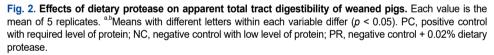


Fig. 1. Effects of dietary protease on apparent ileal digestibility of weaned pigs. Each value is the mean of 5 replicates. ^{a,b}Means with different letters within each variable differ (p < 0.05). PC, positive control with required level of protein; NC, negative control with low level of protein; PR, negative control + 0.02% dietary protease.





to fully utilize nutrients efficiently [29,30]. Similarly, the addition of protease in weaned pig diets may improve their growth performance by increasing nutrient utilization [10,25].

The VH:CD can be an indicator to evaluate nutrient digestion and absorption capacity of the small intestine [31]. The reduction of villus height could induce decreased absorption of nutrients, which may be responsible for the reduced growth performance. The present study showed weaned pigs fed PR had higher VH:CD (p < 0.05) in ileum than those fed NC (Table 3). This result is similar to the result reported by previous studies [12,32]. It may be possible the addition of PR may reduce allergenic reactions that are derived from feeding SBM and that can cause intestinal damage [32,33].

Diarrhea and infectious disease are serious problems around weaning and usually lead to an impaired growth performance and increased mortality of weaning pigs [34]. However during this study PR treatment showed a reduced (p < 0.05) frequency of diarrhea for the first 2 weeks after weaning (Fig. 3) and PCV on d 14 (Fig. 4) with an increased (p < 0.05) number of goblet cells (Fig. 5) compared with the NC and/or PC. Previous studies have also reported that supplementation of dietary enzymes can reduce diarrhea of pigs. This beneficial effect, maybe attributed to the develop-

Table 3. Effects of dietary protease on ileal morphology of weaned pigs	Table 3. Effect	ts of dietary protease o	n ileal morphology o	f weaned pigs ¹⁾
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Item	D	ietary treatment	ts	SEM	<i>p</i> -value ²⁾			
nem	PC	NC	PR	SEIWI	Diet	PC vs. NC	PC vs. PR	NC vs. PR
Villus height (µm)	312.52	282.70	318.23	11.69	0.110	0.097	0.736	0.053
Crypt depth (µm)	102.32	99.15	88.48	5.37	0.204	0.684	0.094	0.186
VH : CD	3.11	2.87	3.67	0.24	0.099	0.507	0.130	0.040
Villus width (µm)	101.72	93.59	98.82	4.23	0.414	0.200	0.649	0.391
Villus area (µm²)	24,938	23,536	23,815	2,064	0.880	0.640	0.707	0.925

¹⁾Each value is the mean of 5 replicates (5 pigs/pen).

²Diet, main diet effect; PC vs. NC, effect between positive and negative controls; PC vs. PR, effect between positive control and negative control + 0.02% dietary protease; NC vs. PR, effect between negative control and negative control + 0.02% dietary protease.

PC, positive control with required level of protein; NC, negative control with low level of protein; PR, negative control + 0.02% dietary protease; VH, villus height; CD, crypt depth; SEM, standard error of means.

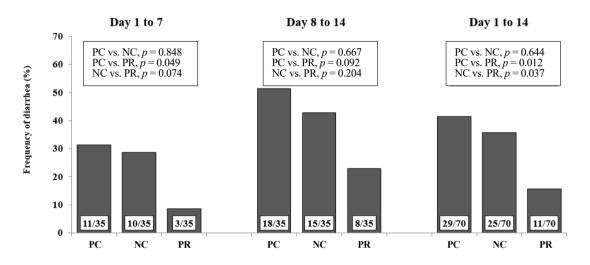


Fig. 3. Effects of dietary protease on frequency of diarrhea of weaned pigs for the first 2 weeks after weaning. Number of diarrhea out of number of pen days is presented in each bar. Frequency of diarrhea (%) = (number of diarrhea / number of pen days) × 100. Data was analyzed by the χ^2 test. Three, four, and one pigs were removed from PC, NC, and PR, respectively (there were no differences among dietary treatments). PC, positive control with required level of protein; NC, negative control with low level of protein; PR, negative control + 0.02% dietary protease.

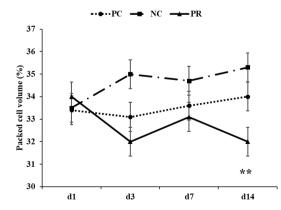


Fig. 4. Effect of dietary protease on packed cell volume of weaned pigs. Each value is the mean of 5 replicates. **Different between PC, NC, and PR (p < 0.05). PC, positive control with required level of protein; NC, negative control with low level of protein; PR, negative control + 0.02% dietary protease.

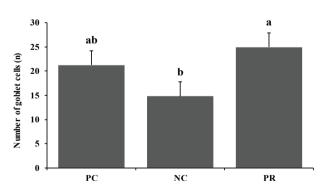


Fig. 5. Effect of dietary protease on number of goblet cells of weaned pigs. Each value is the mean of 5 replicates. ^{a,b}Means with different letters within each variable differ (p < 0.05). PC, positive control with required level of protein; NC, negative control with low level of protein; PR, negative control + 0.02% dietary protease.

ment of the digestive tract, the increase of enzyme activity in the digestive system, and the improvement of nutrient digestibility [10, 35].

CONCLUSION

In conclusion, addition of PR in nursery diets with a low protein level improved growth performance, nutrient digestibility, and intestinal morphology of weaned pigs.

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