Evaluation of black soldier fly larvae reared on different organic substrates on nutrient digestibility and palatability in cats

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Abstract

This study aimed to evaluate the digestibility and palatability of cat diets by substituting poultry meals (PM) with black soldier fly larvae (BSFL) reared on different organic substrates. The experimental treatments are as follows; CON, a basal diet based on the PM; AF3, 3% PM substituted with BSFL reared on animal-based substrates; AF6, 6% PM substituted with BSFL reared on animal-based substrates; PF3, 3% PM substituted with BSFL reared on plant-based substrates; PF6, 6% PM substituted with BSFL reared on plant-based substrates. *In vitro* and *in vivo* methods were used in this study. The *in vitro* experiment simulated gastric digestion using pepsin and small intestinal digestion using bile and pancreatin, with 6 replicates per diet. The *in vivo* experiment used 30 and 16 mixed-sex cats to assess digestibility and palatability, respectively. Fecal samples were collected over 3 d for nutrient digestibility and palatability was assessed in a 1 d. In the *in vitro* experiment, the AF3 had higher *in vitro* ileal digestibility (IVID) of crude protein (CP) than the CON and PF6, and diets supplemented with BSFL groups had higher IVID of gross energy and ether extract (EE) than the CON. In the *in vivo* experiment, the AF3 showed higher CP digestibility than the CON, and diets supplemented with BSFL groups had higher EE digestibility than the CON. In palatability evaluation, the AF6 had a lower intake ratio than the CON. The 3% substitution of BSFL showed a significantly higher first sniffing bout than the 6% substitution in animal and plant-based substrates. Additionally, except for the CON, the AF3 exhibited a higher first sniffing bout than the other groups. In conclusion, the AF3 improved CP digestibility in both *in vitro* and *in vivo* experiments. Also, the AF3 did not show negative effects on palatability. Therefore, this result indicated that substituting 3% of PM with BSFL reared on animal-based substrate in cat diets was the most efficient alternative.

Keywords: Insect, Poultry meal, Feline

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INTRODUCTION

The growing number of companion animals has led to the rapid expansion of the pet food market [1]. At the same time, the demand for protein sources to meet consumer preferences and nutritional requirements in pet food has been increasing. However, the protein sources currently used in pet food compete with human food industries for resources and face limitations in terms of environmental sustainability [2,3]. Furthermore, cats with carnivorous traits are sensitive to amino acid balance and protein content, making the quality and stable supply of protein crucial in cat diet production [4]. For these reasons, developing high-quality and environmentally sustainable protein sources that meet the nutritional needs of cats has emerged as a key challenge in the pet food industry [5].

Insect protein, including black soldier fly larvae (BSFL), housefly larvae, and mealworms, is increasingly recognized as a sustainable protein source due to its high conversion efficiency and reduced greenhouse gas emissions [6]. Among these, BSFL stands out for its ability to be reared on various organic waste, making it a particularly sustainable alternative [7]. Studies have shown that substituting conventional protein sources in pet diets with BSFL does not compromise the digestibility of dry matter (DM), ether extract (EE), or crude protein (CP) [8–10]. Bosch et al. [11] further demonstrated that fully substituting poultry meal (PM) with BSFL in cat diets preserved digestibility and supported gut health. These findings confirm the safety and potential benefits of BSFL as a protein source for pet health and metabolism. The body composition of BSFL, including protein and lipid content, is influenced by the rearing substrate, which may enhance nutrient utilization when incorporated into pet diets [12,13]. However, studies on the use of BSFL as a protein substitute in cat diets exist, while research comparing the differences in protein substitute potential of BSFL reared on different substrates as a protein source in cat diets remains limited.

Therefore, this study examined the feasibility of partially substituting PM with BSFL reared on various organic substrates in cat diets by conducting a preliminary nutrient utilization assessment using *in vitro* methods. Subsequently, based on the *in vitro* results, the effects on *in vivo* digestibility and palatability were investigated.

MATERIALS AND METHODS

Preparations of BSFL and experimental diet

The BSFL was reared on animal and plant-based substrates. The BSFL reared on an animal-based substrate was provided by Chungbuk Agricultural Research and Extension Services (Cheongju, Korea), and the BSFL reared on a plant-based substrate was obtained from Inseong Industry (Jeju, Korea). The BSFL reared on the animal-based substrate were fed milk sludge and feed waste in a 7:3 ratio, at 28 ± 2℃ and 60 ± 10% humidity. The BSFL reared on the plant-based substrate were fed citrus pulp and soybean meal in an 8:2 ratio, at 25 ± 3℃ and 70 ± 5% humidity. All BSFL used in the experiment were 3rd instar larvae reared for 10 d. After rearing, the larvae were air-dried, underwent secondary drying to reduce moisture content to below 1%, followed by grinding for experimental purposes. The chemical composition of the experimental diets and the ingredient profiles are presented in Tables 1 and 2, respectively. The experimental treatments were as follows: CON, basal diet; AF3, 3% BSFL reared on animal-based substrate substituting with PM; AF6, 6% BSFL reared on animal-based substrate substituting with PM; PF3, 3% BSFL reared on plant-based substrate substituting with PM; PF6, 6% BSFL reared on plant-based substrate substituting with PM.

Table 1. Chemical composition of the BSFL

Items (%)	BSFL reared on animal-based substrate	BSFL reared on plant-based substrate
Dry matter	96.68	93.46
Gross energy (kcal/g)	5,603.73	5,753.30
Crude protein	48.37	38.51
Ether extract	23.66	35.65
Crude fiber	8.11	7.78
Crude ash	10.06	10.49
Aspartic acid	5.40	5.55
Threonine	2.56	2.57
Serine	2.95	2.97
Glutamic acid	8.50	8.42
Glycine	2.77	2.78
Alanine	4.42	4.52
Valine	3.31	3.35
Isoleucine	2.32	2.31
Leucine	4.65	4.76
Tyrosine	3.49	3.53
Phenylalanine	2.09	2.07
Lysine	3.82	3.89
Histidine	1.95	1.93
Arginine	3.28	3.31
Cysteine	0.49	0.48
Methionine	0.71	0.65
Proline	4.62	4.72

BSFL, black soldier fly larvae.

Experiment 1
Digestibility assay
In vitro method

The *in vitro* trial described by Soutar et al. [14] was conducted with 6 replicates per diet. The samples were prepared in finely ground (< 1.0 mm) form. In stomach simulation, weigh (5.000 ± 0.005 g) of each sample in 250 mL Erlenmeyer flasks, then add 85 mL of ultra-high-quality water (> 18 MΩ). The pH was adjusted to 2.0 using 1 M HCl and 1 M sodium bicarbonate solution by gradually adding each to reach the desired level. The sample was equilibrated at 39℃ for 15 min, before 10 mL pepsin solution (20 mg/mL, ≥ 250 units/mg; solid, P7000, pepsin from porcine gastric mucosa, Sigma-Aldrich, St. Louis, MO, USA) was added to the flask to simulate stomach digestion in the cat. In addition, 5 mL of chloramphenicol solution (C0378, chloramphenicol, Sigma-Aldrich with 5 g/L ethanol) was added to prevent bacterial fermentation. The flasks were closed with a Parafilm M® film and incubated in a shaking incubator (SWB-35, Hanyang Science Lab, Seoul, Korea) at 39℃ for 1.5 h. In the next step, during the small intestine simulation, a 1 M NaOH solution was added to adjust the pH to 6.8, and then the flask was cooled to room temperature. Subsequently, 20 mL of an 80 mg/mL bile salts (B8756, Sigma Aldrich) solution and 20 mL of a pancreatin solution (1 mg/mL 8 × USP pancreatin composed of amylase [3,720 U/mg], protease [2880 U/mg], and lipase [100–650 U/mg]; P7545, Sigma-Aldrich) were added to the flask to simulate digestion conditions in the cat's small intestine. Then, the flasks were closed

Table 2. Ingredient composition of experimental diets

Ingredients (%)	CON[1]	AF3	AF6	PF3	PF6
Poultry meal	35.00	32.00	29.00	32.00	29.00
BSFL_A	-	3.00	6.00	-	-
BSFL_P	-	-	-	3.00	6.00
Rice	21.36	21.36	21.36	21.36	21.36
Wheat flour	14.74	14.74	14.74	14.74	14.74
Wheat bran	10.00	10.00	10.00	10.00	10.00
Soybean meal	5.00	5.20	5.50	5.70	6.40
DDGS	5.00	5.00	5.00	5.00	5.00
Poultry oil	5.00	4.80	4.50	4.30	3.60
Beet pulp	2.00	2.00	2.00	2.00	2.00
Salt	0.70	0.70	0.70	0.70	0.70
Taurine	0.20	0.20	0.20	0.20	0.20
Min + Vit[2]	1.00	1.00	1.00	1.00	1.00
Total	100.00	100.00	100.00	100.00	100.00
Chemical composition					
ME (kcal/kg)	3,633	3,635	3,632	3,633	3,633
CP (%)	32.01	31.61	31.25	31.55	31.09
EE (%)	10.56	10.74	10.82	10.66	10.77
CF (%)	3.64	3.83	4.03	3.86	4.07
Ash (%)	2.33	2.56	2.80	2.59	2.85
Ca (%)	1.11	1.18	1.26	1.19	1.28
P (%)	0.69	0.68	0.68	0.69	0.68

[1]CON, basal diet; AF3, 3% BSFL reared on animal-based substrate substituting with poultry meal; AF6, 6% BSFL reared on animal-based substrate substituting with poultry meal; PF3, 3% BSFL reared on plant-based substrate substituting with poultry meal; PF6, 6% BSFL reared on plant-based substrate substituting with poultry meal.

[2]Provided per kg diet: 10.8 mg copper ($CuSO_4$), 0.36 mg selenium (Na_2SeO_3), 150 mg zinc ($ZnSO_4$, ZnO), 17.4 mg manganese ($MnSO_4$), 284.3 mg iron ($FeSO_4$), 17.2 mg copper ($CuSO_4$), 2.2 mg cobalt ($CoSO_4$), 166.3 mg zinc ($ZnSO_4$), 7.5 mg iodine (KI), and 0.2 mg selenium (Na_2SeO_3), 2,562.8 IU vitamin A, 254 IU vitamin D3, 32.1 IU vitamin E.

BSFL_A, black soldier fly larvae reared on an animal-based substrate; BSFL_P, black soldier fly larvae reared on a plant-based substrate; DDGS, distiller's dried grains with solubles; ME, metabolizable energy;.CP, crude protein; EE, ether extract; CF, crude fiber.

with a Parafilm M® film and incubated in a shaking incubator (SWB-35, Hanyang Science Lab) at 39℃ for 3 h. The collected undigested samples were filtered through pre-dried and pre-weighed filter crucibles (Gooch Type Filter Crucibles, PYREX, Sunderland, UK). During filtration, the flasks were rinsed three times with distilled water. Additionally, 10 mL of 95% ethanol and 10 mL of 99.5% acetone were added twice to the glass filter crucibles. At the end of the *in vitro* trial, the filter crucibles containing the undigested residues were dried at 70℃ for 24 hours and collected to calculate DM.

Chemical analysis and calculations

All diets and residues were crushed on a 1 mm screen and chemically analyzed in 6 replicates. The diets and residues of DM (method 930.15), and EE (method 920.39) were determined using the AOAC [15] method. The gross energy (GE) content was analyzed by bomb calorimeter (Parr 6400 Bomb Calorimeter, Parr Instrument, Moline, IL, USA). The CP content was determined using the dumas (Rapid MAX N-Exceed, Elementar, Langenselbold, Germany).

Calculating the *in vitro* digestibility of DM using the following formula:

"Digestibility (%) = 100 – [(residue weight/sample weight) × 100]

Calculating the *in vitro* digestibility of CP, EE, and GE used the following formula:

"Digestibility (%) = 100 – [Nr × (100 – IDDM) / Nd]"

Nr =nutrient concentration in residues (DM %), Nd = nutrient concentration in diet (DM %), IDDM =*in vitro* digestibility of DM (%).

Experiment 2
In vivo method
Animal ethics
The experimental protocol was approved (CBNUA-24-0039-01) by the Institutional Animal Care and Use Committee of Chungbuk National University, Cheongju, Korea.

Animals and experiment design
30 healthy adult domestic cats of mixed sex cats (12 males and 18 females) were used in a triplicated 5 × 5 Latin square design. The cats, aged 5–7 years, had an average body weight of 5.06 ± 0.89 kg, and their diet was controlled to meet or exceed the nutrient profile for adult cats established by the Association of American Feed Control Officials [16]. Each period consisted of 7 d of diet adaptation and 3 d of total fecal collection. Each cat was housed in an individual cage (0.9 m × 0.9 m × 0.9 m) except for diet Cats were randomly assigned to one of the five experimental diets and were fed to maintain body weight. Water was available *ad libitum*. Cats were reared individually during feeding (two times daily: 08:00 to 10:00 and 15:00 to 17:00) and fecal collection periods but were housed in groups except during the experimental period. The cats were housed on a 12-light cycle with lights off from 19:00 to 07:00.

Nutrient digestibility
Apparent total tract digestibility (ATTD) of DM, CP, and gross energy (GE) were determined using 1% celite as an inert indicator by Scott and Boldaji [17] method. Cats were fed diets mixed with celite from 1 to 3 d and diet samples were also collected. Fresh fecal samples were collected from 2 to 4 d. Fresh fecal and diet samples were stored in a freezer at –20℃ immediately after collection. At the end of the experiment, fecal samples were dried at 70℃ for 72 h and then crushed on a 1 mm screen. All diet and fecal samples were then analyzed for DM, CP, and GE following the procedures by the AOAC [15]. Celite levels were determined using Scott and Boldaji [17] method. The GE of diets and feces were analyzed using an adiabatic oxygen bomb calorimeter (6400 Automatic Isoperibol calorimeter, Parr, USA). For calculating the ATTD of the nutrients, we used the following equation: Digestibility = 1 – [(Nf × Cd)/ (Nd × Cf)] × 100, where Nf = concentration of nutrient in fecal, Nd = concentration of nutrient in the diet, Cd = concentration of celite in the diet, and Cf = concentration of celite in the fecal.

Experiment 3
Palatability test
Animal and experimental procedures
16 healthy adult domestic cats of mixed sex (8 males and 8 females) aged 5–7 years with a body weight of 5.12 ± 0.75 kg, were used to determine palatability when substituting PM with BSFL reared on different organic substrates. The palatability test used the five-bowl test method and

estimated feed intake, intake ratio, first sniffing bout, first eating bout, and time to eat. Five treatment diets were prepared in each bowl, with 16 cats individually assigned a 5-minute feeding time. The position of each bowl was assigned following a Latin Square Design, where each diet was rotated systematically to ensure equal exposure to all cats, minimizing positional bias. Feed intake was calculated by subtracting the remaining diet amount from the initially provided. The intake ratio was determined by dividing feed intake by the amount provided. The first sniffing bouts were recorded as the cumulative instances of cats smelling each diet. The total number of first eating bouts indicated the cumulative cases of feeding behavior observed for each diet. Additionally, the time to eat was measured to quantify the duration of feeding behavior for each diet.

Statistical analysis

Data including the palatability, *in vitro* and *in vivo* digestibility by diet was conducted one-way ANOVA and analyzed with the PROC Generalized Linear Models of the JMP (JMP® Pro version 16.0.0, SAS Institute, Cary, NC, USA). The first sniffing bout and first eating bout were visualized using GraphPad Prism 9.5.1 (GraphPad, San Diego, CA, USA). Differences between treatment means were determined using Tukey's multiple-range test. A probability level of $p < 0.05$ was indicated to be statistically significant, and a level of $0.05 \leq p < 0.10$ was considered to have such a tendency.

RESULTS

In vitro digestibility

The effect of substituting PM with BSFL reared on different organic substrates on *in vitro* digestibility in the cat diet is presented in Table 3. All treatments substituting PM with BSFL showed significantly higher GE and EE digestibility than the CON. Also, the AF3 showed significantly higher CP digestibility than the CON and PF6.

In vivo digestibility

The effect of substituting PM with BSFL reared on different organic substrates on nutrient digestibility in the cat diet is presented in Table 4. All treatments substituting PM with BSFL showed significantly higher EE digestibility than the CON. Also, the AF3 showed significantly higher CP digestibility than the CON.

Palatability

The effect of substituting PM with BSFL reared on different organic substrates on nutrient digestibility in the cat diet is presented in Table 5.

Table 3. *In vitro* intestinal digestibility of cat diets with BSFL reared on different organic substrates (Exp 1)

Items (%)	CON[1]	AF3	AF6	PF3	PF6	SE	*p*-value
DM	79.59	80.06	80.12	79.91	80.15	0.211	0.344
CP	79.75[b]	81.14[a]	80.59[ab]	80.36[ab]	79.96[b]	0.273	0.012
EE	84.62[b]	86.68[a]	86.62[a]	86.73[a]	86.68[a]	0.372	0.001
GE	81.61[b]	83.58[a]	83.69[a]	83.70[a]	83.90[a]	0.227	< 0.001

[1]CON, basal diet; AF3, 3% BSFL reared on animal-based substrate substituting with poultry meal; AF6, 6% BSFL reared on animal-based substrate substituting with poultry meal; PF3, 3% BSFL reared on plant-based substrate substituting with poultry meal; PF6, 6% BSFL reared on plant-based substrate substituting with poultry meal.

[a,b]Means within columns with different superscripts differ significantly ($p < 0.05$).

BSFL, black soldier fly larvae; DM, dry matter; CP, crude protein; EE, ether extract; GE, gross energy.

Table 4. Nutrient digestibility of cat diets with BSFL reared on different organic substrates (Exp 2)

Item (%)	CON[1]	AF3	AF6	PF3	PF6	SE	*p*-value
DM	83.29	86.91	86.81	86.17	85.82	1.198	0.186
CP	87.31[b]	90.24[a]	89.52[ab]	88.96[ab]	89.15[ab]	0.651	0.049
EE	93.51[b]	95.14[a]	94.82[a]	94.95[a]	94.74[a]	0.264	0.002
GE	81.29	82.83	83.60	81.22	80.18	1.164	0.268

[1]CON, basal diet; AF3, 3% BSFL reared on animal-based substrate substituting with poultry meal; AF6, 6% BSFL reared on animal-based substrate substituting with poultry meal; PF3, 3% BSFL reared on plant-based substrate substituting with poultry meal; PF6, 6% BSFL reared on plant-based substrate substituting with poultry meal.

[a,b]Means within columns with different superscripts differ significantly ($p < 0.05$).

BSFL, black soldier fly larvae; DM, dry matter; CP, crude protein; EE, ether extract; GE, gross energy.

Table 5. Palatability of cat diet with BSFL reared on different organic substrates (Exp 3)

Items	CON[1]	AF3	AF6	PF3	PF6	SE	*p*-value
Feed intake (g)	13.65	8.77	0.00	6.69	2.08	3.463	0.054
Intake ratio	14.49[a]	8.72[ab]	0.00[b]	6.78[ab]	2.08[ab]	3.487	0.039
Time to eat (sec)	146.00	135.00	0.00	76.00	31.00	49.45	0.275

[1]CON, basal diet; AF3, 3% BSFL reared on animal-based substrate substituting with poultry meal; AF6, 6% BSFL reared on animal-based substrate substituting with poultry meal; PF3, 3% BSFL reared on plant-based substrate substituting with poultry meal; PF6, 6% BSFL reared on plant-based substrate substituting with poultry meal.

[a,b]Means within columns with different superscripts differ significantly ($p < 0.05$).

BSFL, black soldier fly larvae.

The AF6 showed a significantly lower intake ratio than the CON. The CON and AF3 showed higher first sniffing bouts than the AF6 and PF6 (Figs. 1 and 2). The CON exhibited a higher first eating bout compared to the AF6.

DISCUSSION

This study was conducted to evaluate the potential of BSFL reared on different substrates as a

Fig. 1. First sniffing of cat diets with BSFL reared on different organic substrates. CON, basal diet; AF3, 3% BSFL reared on animal-based substrate substituting with poultry meal; AF6, 6% BSFL reared on animal-based substrate substituting with poultry meal; PF3, 3% BSFL reared on plant-based substrate substituting with poultry meal; PF6, 6% BSFL reared on plant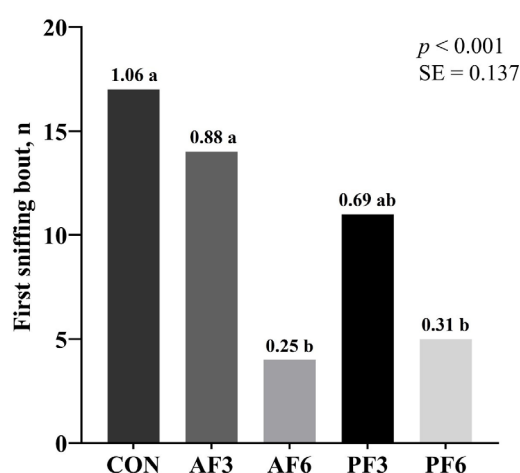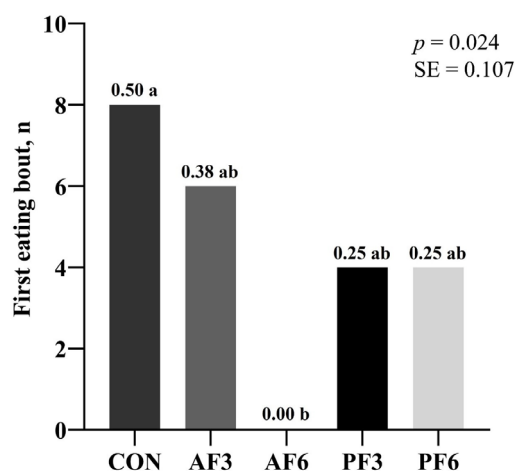-based substrate substituting with poultry meal. [a,b]Means within columns with different superscripts differ significantly ($p < 0.05$). BSFL, black soldier fly larvae.

Fig. 2. First eating of cat diets with BSFL reared on different organic substrates. CON, basal diet; AF3, 3% BSFL reared on animal-based substrate substituting with poultry meal; AF6, 6% BSFL reared on animal-based substrate substituting with poultry meal; PF3, 3% BSFL reared on plant-based substrate substituting with poultry meal; PF6, 6% BSFL reared on plant-based substrate substituting with poultry meal. [a,b]Means within columns with different superscripts differ significantly ($p < 0.05$). BSFL, black soldier fly larvae.

protein substitute in cat diets. In this study, differences in body composition were observed in BSFL reared on different substrates, with those reared on animal-based substrates showing higher CP and lower EE and ash than those reared on plant-based substrates. Similarly, Nyakeri et al. [18] reported that the growth rate and body composition of BSFL varies depending on the rearing substrate, with CP ranging from 36.1% to 45.4% and EE ranging from 18.1% to 38.0% in 16-day-old larvae reared on different organic substrates. Additionally, St-Hilaire et al. [19] demonstrated that the nutrient composition of substrates is directly associated with the body composition of BSFL, with an increased fish waste content in the substrate leading to a higher omega-3 fatty acid in the larvae. These findings suggest that the body composition of BSFL can be modified by rearing substrates, highlighting their potential differentiation as a protein substitute.

In this study, the *in vitro* revealed that AF3 exhibited a higher CP digestibility than PF6. Previous studies have reported a negative correlation between ash content and CP digestibility [20]. Meyer and Mundt [21] observed that higher ash content in diets can increase the pH of digesta, thereby reducing pepsin activity essential for protein breakdown. Likewise, a high-ash diet for dogs (8% of DM) showed a 4% reduction in CP digestibility compared to a low-ash diet (6% of DM) [9]. Consistent with this observation, BSFL reared on animal-based diets exhibited lower ash content and higher CP digestibility than plant-based BSFL. However, the *in vivo* CP digestibility did not show differences for BSFL reared on different organic substrates. This is attributed to the limit of the *in vitro* method. *In vitro* methods, while simulating digestive enzyme activity and intestinal conditions, do not fully reflect the complexity of physiological processes. Moreover, cats are a species sensitive to amino acid imbalances, this may have contributed to the differences observed between the two digestibility evaluation methods [4]. In this study, AF3 demonstrated higher CP digestibility than CON in both *in vitro* and *in vivo* evaluations, despite its lower CP content. These results may suggest that the high CP content, exceeding the requirements, could have reduced digestibility [22]. Similarly, El-Wahab et al. [20] reported a 2% increase in CP digestibility when PM was fully replaced with BSFL in dog diets. However, other studies, including those by Do et al. [23] and Freel et al. [8], indicated no significant differences in CP digestibility when 5% of PM was substituted with BSFL. Consistent with these findings, this study also

observed no significant difference between the PF3 and CON, suggesting that replacing PM with BSFL does not negatively influence CP digestibility in cat diets. These findings suggest that the rearing environment of BSFL may influence its nutritional effects, highlighting the need to explore optimized rearing strategies for its effective utilization. Moreover, the results of this study indicate that substituting 3% of PM with BSFL reared on animal-based organic matter was the most efficient alternative. These findings suggest that the rearing environment of BSFL may influence its nutritional effects, highlighting the need to explore optimized rearing strategies for its effective utilization.

The BSFL diet had a higher fat content than the CON diet. According to Zuo et al. [24], an increase in dietary fat content enhances fat digestibility, consistent with the higher fat digestibility observed in the BSFL diet in our study. Similarly, Butowski et al. [25] found that high-fat diets (190 g/kg) achieved 99% fat digestibility in cats. Additionally, EE digestibility can be influenced by the carbon chain length and type of lipids [26,27]. With their shorter carbon chains, medium-chain fatty acids (MCFAs) are emulsified and absorbed more efficiently. In contrast, long-chain fatty acids (LCFAs) require longer emulsification and absorption due to their extended chains [28]. BSFL contains a high concentration of MCFAs, such as lauric acid (C12:0) [7,29], while PM mainly consists of LCFAs, like palmitic acid (C16:0) [30,31]. The carbon chain length is critical in emulsification, with MCFAs being more readily emulsified and absorbed than LCFAs [32]. Our study identified differences in fat digestibility in cats due to variations in emulsification. Consistent with our results, Do et al. [23] reported that substituting poultry fat with BSFL oil significantly improved EE digestibility. The MCFAs in BSFL demonstrated higher absorption and permeation through enterocytes than the LCFAs in PM, attributed to their superior emulsifying capacity. This suggests that partially replacing PM with BSFL may enhance EE digestibility. Also, our study indicated the need for future research to compare the fatty acid profiles of these two ingredients.

Although substituting 3% of PM with BSFL reared on animal-based substrates increased CP and EE digestibility, no significant difference was observed in DM digestibility. This result may be related to the high dietary fiber content in BSFL, which is resistant to enzymatic digestion, potentially limiting DM digestibility. Previous studies have similarly reported that, despite a 4% increase in CP digestibility and a 3% increase in fat digestibility in cats fed a high-fiber diet, DM digestibility decreased by approximately 7% [33]. This evidence suggests that a 3% substitution level of PM with BSFL may be more effective than a 6% level, possibly due to the low fiber content at lower substitution levels, which minimizes its impact on DM digestibility.

In the pet food market, palatability is a critical indicator for evaluating how well pets accept and prefer a particular diet, which is directly influenced by their sense of taste and smell [6]. The palatability was assessed through first sniffing, eating, and intake measurements. In this study, AF3 did not significantly influence the intake ratio compared to the CON while AF6 decreased the intake ratio. Furthermore, the first sniffing bout was significantly higher in the 3% BSFL substitution group than in the 6% substitution group. According to previous research, substituting insect proteins (Nauphoeta cinerea, Gromphadorhina portentosa, and Zophobas morio larvae) for about 3% of the PM in cat diet did not significantly affect feed intake [34]. Similarly, Do et al. [23] showed a numerical decrease in feed intake when more than 5% of PM was replaced with whole BSFL. This may be attributed to the high content of MCFAs in BSFL. While cats are known to prefer both animal and plant-based fats, an increase in MCFAs could negatively impact their palatability [35]. Our findings indicate that an increase in the substitution PM with BSFL level reduces palatability, suggesting that a 3% substitution is an optimal level for effective replacement. These findings suggest that the specific ingredients of BSFL may have negatively impacted palatability for cats. Therefore, this study provides useful information into the palatability of cat diet

with 3% and 6% BSFL substitutions, and it illustrates the need for additional research to clarify the mechanisms by which BSFL influences cat feed intake.

CONCLUSION

This study also provides data on substituting PM with BSFL reared on different substrates in cat diets. Substituting PM with BSFL in diets increased EE digestibility without negatively affecting CP digestibility. Additionally, substituting 3% of PM with BSFL reared on an animal-based substrate significantly improved CP digestibility in both *in vitro* and *in vivo* methods. Furthermore, the AF3 showed a significantly higher first sniffing bout in the palatability test compared to other groups, with no significant difference in feed intake compared to the CON group. These findings indicate that BSFL may serve as a suitable protein alternative in cat diets, with BSFL reared on animal-based substrates being effective at a 3% substitution level.

REFERENCES

1. Dust JM, Grieshop CM, Parsons CM, Karr-Lilienthal LK, Schasteen CS, Quigley JD 3rd, et al. Chemical composition, protein quality, palatability, and digestibility of alternative protein sources for dogs. J Anim Sci. 2005;83:22. https://doi.org/10.2527/2005.83102414x

2. Swanson KS, Carter RA, Yount TP, Aretz J, Buff PR. Nutritional sustainability of pet foods. Adv Nutr. 2013;4:141-50. https://doi.org/10.3945/an.112.003335

3. Seo K, Cho HW, Chun J, Jeon J, Kim C, Kim M, et al. Evaluation of fermented oat and black soldier fly larva as food ingredients in senior dog diets. Animals. 2021;11:3509. https://doi.org/10.3390/ani11123509

4. Li P, Wu G. Amino acid nutrition and metabolism in domestic cats and dogs. J Anim Sci Biotechnol. 2023;14:19. https://doi.org/10.1186/s40104-022-00827-8

5. Li Y, Liu M, Wei Y, Li L, Ma D, Weng Y, et al. Influence of a mixture of protein hydrolysate from black soldier fly larvae and schizochytrium on palatability, plasma biochemistry, and antioxidative and anti-inflammatory capacity in cat diets. Animals. 2024;14:751. https://doi.org/10.3390/ani14050751

6. Pezzali JG, Shoveller AK. Short communication: the effects of a semi-synthetic diet with inclusion of black soldier fly larvae meal on health parameters of healthy adult cats. J Anim Sci. 2021;99:skab290. https://doi.org/10.1093/jas/skab290

7. Spranghers T, Ottoboni M, Klootwijk C, Ovyn A, Deboosere S, De Meulenaer B, et al. Nutritional composition of black soldier fly (Hermetia illucens) prepupae reared on different organic waste substrates. J Sci Food Agric. 2017;97:2594-600. https://doi.org/10.1002/jsfa.8081

8. Freel TA, McComb A, Koutsos EA. Digestibility and safety of dry black soldier fly larvae meal and black soldier fly larvae oil in dogs. J Anim Sci. 2021;99:skab047. https://doi.org/10.1093/jas/skab047

9. Penazzi L, Schiavone A, Russo N, Nery J, Valle E, Madrid J, et al. In vivo and in vitro digestibility of an extruded complete dog food containing black soldier fly (Hermetia illucens) larvae meal as protein source. Front Vet Sci. 2021;8:653411. https://doi.org/10.3389/fvets.2021.653411

10. Jian S, Zhang L, Ding N, Yang K, Xin Z, Hu M, et al. Effects of black soldier fly larvae as protein or fat sources on apparent nutrient digestibility, fecal microbiota, and metabolic profiles in beagle dogs. Front Microbiol. 2022;13:1044986. https://doi.org/10.3389/

fmicb.2022.1044986

11. Bosch G, Loureiro BA, Schokker D, Kar SK, Paul A, Sluczanowski N. Black soldier fly larvae meal in an extruded food: effects on nutritional quality and health parameters in healthy adult cats. J Insects Food Feed. 2024;10:1595-606. https://doi.org/10.6084/m9.figshare.25533625

12. Raksasat R, Lim JW, Kiatkittipong W, Kiatkittipong K, Ho YC, Lam MK, et al. A review of organic waste enrichment for inducing palatability of black soldier fly larvae: wastes to valuable resources. Environ Pollut. 2020;267:115488. https://doi.org/10.1016/j.envpol.2020.115488

13. Fitriana E, Laconi E, Jayanegara A. Influence of various organic wastes on growth performance and nutrient composition of black soldier fly larvae (Hermetia illucens): a meta-analysis. IOP Conf Ser Earth Environ Sci. 2021;788:012051. https://doi.org/10.1088/1755-1315/788/1/012051

14. Soutar L, Coltherd JC, Steele VR, Staunton R, Carvell-Miller L, Hughes KR, et al. Comparisons of in vitro and in vivo digestibility assays for phosphorus in feline diets and associations with dietary nutrient content. J Agric Food Chem. 2021;69:10688-99. https://doi.org/10.1021/acs.jafc.1c03308

15. AOAC (Association of Official Analytical Chemists) International. Official methods of analysis of AOAC International. 16th ed. Washington, DC: AOAC Intenational; 2000.

16. AAFCO (Association of American Feed Control Officials). 2021 Official publication. Champaign, IL: AAFCO; 2021. p. 111-236.

17. Scott TA, Boldaji F. Comparison of inert markers [chromic oxide or insoluble ash (Celite)] for determining apparent metabolizable energy of wheat-or barley-based broiler diets with or without enzymes. Poult Sci. 1997;76:8. https://doi.org/10.1093/ps/76.4.594

18. Nyakeri E, Ogola H, Ayieko M, Amimo F. Valorisation of organic waste material: growth performance of wild black soldier fly larvae (Hermetia illucens) reared on different organic wastes. J Insects Food Feed. 2017;3:193-202.

19. St-Hilaire S, Cranfill K, McGuire MA, Mosley EE, Tomberlin JK, Newton L, et al. Fish offal recycling by the black soldier fly produces a foodstuff high in omega-3 fatty acids. J World Aquac Soc. 2007;38:309-13. https://doi.org/10.1111/j.1749-7345.2007.00101.x

20. Abd El-Wahab A, Meyer L, Kölln M, Chuppava B, Wilke V, Visscher C, et al. Insect larvae meal (Hermetia illucens) as a sustainable protein source of canine food and its impacts on nutrient digestibility and fecal quality. Animals. 2021;11:2525. https://doi.org/10.3390/ani11092525

21. Meyer H, Mundt H. Untersuchungen zum einsatz von knochenschrot in futterationen fur hunde. Deutsch Tierarztl Wochenschr. 1983;90:81-120.

22. Park S, Lee J, Yun W, Hong S, Oh H, Song D, et al. Supplementation of protease and different nutrient density diets in growing-finishing pigs. J Anim Sci Technol. 2024;66:326-39. https://doi: 10.5187/jast.2023.e55

23. Do S, Koutsos EA, McComb A, Phungviwatnikul T, de Godoy MRC, Swanson KS. Palatability and apparent total tract macronutrient digestibility of retorted black soldier fly larvae-containing diets and their effects on the fecal characteristics of cats consuming them. J Anim Sci. 2022;100:skac068. https://doi.org/10.1093/jas/skac068

24. Zuo Y, Fahey GC Jr, Merchen NR, Bajjalieh NL. Digestion responses to low oligosaccharide soybean meal by ileally-cannulated dogs. J Anim Sci. 1996;74:2441-9. https://doi.org/10.2527/1996.74102441x

25. Butowski CF, Thomas DG, Young W, Cave NJ, McKenzie CM, Rosendale DI, et al. Addition of plant dietary fibre to a raw red meat high protein, high fat diet, alters the faecal bacteriome and organic acid profiles of the domestic cat (Felis catus). PLOS ONE. 2019;14:e0216072.

https://doi.org/10.1371/journal.pone.0216072

26. Asaro NJ, Guevara MA, Berendt K, Zijlstra R, Shoveller AK. Digestibility is similar between commercial diets that provide ingredients with different perceived glycemic responses and the inaccuracy of using the modified atwater calculation to calculate metabolizable energy. Vet Sci. 2017;4:54. https://doi.org/10.3390/vetsci4040054

27. NRC (National Research Council). Nutrient requirements of dogs and cats. 10th ed. Washington, DC: National Academies Press; 2006.

28. Zheng J, Sun D, Li X, Liu D, Li C, Zheng Y, et al. The effect of fatty acid chain length and saturation on the emulsification properties of pork myofibrillar proteins. Lebensm Wiss Technol. 2021;139:110242. https://doi.org/10.1016/j.lwt.2020.110242

29. Daszkiewicz T, Murawska D, Kubiak D, Han J. Chemical composition and fatty acid profile of the pectoralis major muscle in broiler chickens fed diets with full-fat black soldier fly (Hermetia illucens) larvae meal. Animals. 2022;12:464. https://doi.org/10.3390/ani12040464

30. Peña-Saldarriaga LM, Fernández-López J, Pérez-Alvarez JA. Quality of chicken fat by-products: lipid profile and colour properties. Foods. 2020;9:1046. https://doi.org/10.3390/foods9081046

31. Toomer OT, Livingston M, Wall B, Sanders E, Vu T, Malheiros RD, et al. Feeding high-oleic peanuts to meat-type broiler chickens enhances the fatty acid profile of the meat produced. Poult Sci. 2020;99:2236-45. https://doi.org/10.1016/j.psj.2019.11.015

32. Price KL, Lin X, van Heugten E, Odle R, Willis G, Odle J. Diet physical form, fatty acid chain length, and emulsification alter fat utilization and growth of newly weaned pigs. J Anim Sci. 2013;91:783-92. https://doi.org/10.2527/jas.2012-5307

33. Detweiler KB, He F, Mangian HF, Davenport GM, de Godoy MR. Extruded feline diets formulated with high inclusion of soybean hulls: effects on apparent total tract macronutrient digestibility, and fecal quality and metabolites. J Anim Sci. 2019;97:1042-51. https://doi.org/10.1093/jas/skz014

34. Hu Y. Insect meals as novel protein sources in retorted pet food for adult cats [Master's thesis]. Urbana, IL: University of Illinois at Urbana-Champaign; 2020.

35. MacDonald ML, Rogers QR, Morris JG. Aversion of the cat to dietary medium-chain triglycerides and caprylic acid. Physiology & behavior, 1985.35.371-375.